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CD8\textsuperscript{+} T cells from patients with narcolepsy and healthy controls recognize hypocretin neuron-specific antigens

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Narcolepsy Type 1 (NT1) is a neurological sleep disorder, characterized by the loss of hypocretin/orexin signaling in the brain. Genetic, epidemiological and experimental data support the hypothesis that NT1 is a T-cell-mediated autoimmune disease targeting the hypocretin producing neurons. While autoreactive CD4\textsuperscript{+} T cells have been detected in patients, CD8\textsuperscript{+} T cells have only been examined to a minor extent. Here we detect CD8\textsuperscript{+} T cells specific toward narcolepsy-relevant peptides presented primarily by NT1-associated HLA types in the blood of 20 patients with NT1 as well as in 52 healthy controls, using peptide-MHC-I multimers labeled with DNA barcodes. In healthy controls carrying the disease-predisposing HLA-DQB1*06:02 allele, the frequency of autoreactive CD8\textsuperscript{+} T cells was lower as compared with both NT1 patients and HLA-DQB1*06:02-negative healthy individuals. These findings suggest that a certain level of CD8\textsuperscript{+} T-cell reactivity combined with HLA-DQB1*06:02 expression is important for NT1 development.
Narcolepsy type 1 (NT1) is a chronic neurological sleep disorder characterized by dysregulation of the sleep–wake cycle, leading to early occurring rapid eye movement (REM) sleep, excessive daytime sleepiness, and disrupted nighttime sleep. Another characteristic of NT1 is muscle tonus dysregulation during wakefulness, resulting in sudden loss of muscle tone (cataplexy). Furthermore, sleep paralysis, hypnagogic hallucinations, and REM sleep behavior disorder REM sleep without atonia are often seen1–3. NT1 is caused by disrupted signaling of the sleep-regulating neuropeptide hypocretin in the brain4 and it has been shown that this is owing to the loss of specific neurons in the hypothalamus that produce hypocretin5,6. An autoimmune basis for NT1 has long been suspected based on a strong association with the common HLA-DQ haplotype, DQA1*01:02/DQB1*06:02, which encodes the MHC class II DQ*0602 heterodimer7,8. This HLA association is one of the highest known: up to 98% of NT1 patients with demonstrated hypocretin deficiency carry DQ*0602 versus ~25% of the healthy population9,10.

Associations between several MHC class I molecules and narcolepsy have also been suggested by two independent studies10,11. HLA-A*11:01, HLA-B*51:01, and HLA-C*04:01 were found in both studies, whereas HLA-B*35:01 and HLA-B*35:03 were found in the study by Tafti et al.10 and Ollila et al.11, respectively; the discrepancy between the two subtypes is likely owing to ethnicity differences in the two cohorts. Ollila et al.11 further reported that HLA-B*18:01 is associated with narcolepsy, whereas HLA-B*07:02 had a weak protective effect.

Following the 2009/2010 H1N1 influenza vaccination campaigns with Pandemrix, as well as after the H1N1 epidemic itself, narcolepsy incidence dramatically increased in several countries12–14, further substantiating the role of the immune system in NT1 disease development.

Remarkably, even after the discovery of hypocretin-producing neurons as the putative autoimmune target, attempts to demonstrate narcolepsy-associated autoimmune responses have largely been unsuccessful (reviewed in ref. 15), until recently where autoreactive CD4+ T cells targeting hypocretin were detected in blood samples from narcolepsy patients16 and CD4+ T cells recognizing hypocretin were demonstrated to cross-react to the hemagglutinin protein from the 2009/2010 H1N1 influenza A virus17. As neurons express only MHC class I and not class II molecules under normal physiological conditions18, cytotoxic CD8+ T cells are the most likely effector cells in the autoimmune destruction of hypocretin neurons19. This is supported by the finding of post mortem hypothalamic CD8+ T-cell infiltration in a case of NT1 secondary to anti-Ma-associated diencephalitis20. The CD8+ T-cell infiltration was associated with a complete loss of hypocretinerigic neurons. Importantly, it has also been demonstrated in a mouse model that cytotoxic CD8+ T cells with reactivity toward hemagglutinin can specifically kill hypocretin neurons if these transgenically express hemagglutinin. This was not the case for CD4+ T cells targeting hemagglutinin. Even though these cells infiltrated the brain and caused local inflammation, this did not lead to loss of hemagglutinin-expressing hypocretin neurons21. Thus, even though autoreactive CD4+ T cells might initiate the disease process, we hypothesize that the presence of autoreactive CD8+ T cells could be necessary for the development of genuine NT1. In the recent study by Latorre et al.16 describing autoreactive CD4+ T cells, the researchers also searched for autoreactive CD8+ T cells. This was limited to reactivity toward hypocretin, and only 10 NT1 patients and 9 healthy controls were tested. Three of these patients, and two controls harbored CD8+ T cells responsive toward hypocretin epitopes. Even though hypocretin is the best-known marker of hypocretin neurons, other proteins specific to these neurons could also be the target of an autoimmune process. Testing for CD8+ reactivity toward other targets in NT1 blood samples is therefore an important next step for understanding NT1 pathogenesis.

We use a recently developed technique for detection of antigen specific CD8+ T cells that is especially valuable in identifying CD8+ T-cell populations of low frequency and affinity, as it does not rely exclusively on fluorescence separation like conventional tetramer methods22. Instead, it utilizes DNA barcode-labeled peptide-MHC (pMHC) complexes to identify CD8+ T cells specific for the peptide presented. Using this method, we were able to screen for CD8+ T-cell recognition of 1183 peptides expressed in hypocretin neurons and detect such cells in both NT1 patients and healthy controls. We observe a number of differences in the CD8+ T-cell recognition profile between NT1 patients and healthy controls expressing HLA-DQB1*06:02 and hypothesize that the combination of a certain level of CD8+ T-cell reactivity and HLA-DQB1*06:02 expression is necessary for NT1 development.

### Results

#### Experimental strategy and peptide selection

To test the hypothesis that NT1 is mediated by CD8+ T cells, we screened blood samples from 20 NT1 patients and 52 healthy controls for the presence of autoreactive CD8+ T cells targeting epitopes present in hypocretin neurons. NT1 was diagnosed according to the International Classification of Sleep Disorders, Edition 3 (ICSD-3)23. An overview of the experimental strategy is given in Fig. 1a and the core demographic, clinical, and sleep investigation parameters for the narcolepsy cohort are shown in Table 1.

As nearby neurons in the hypothalamus are not affected in NT1, the epitopes should derive from proteins unique to hypocretin neurons, at least locally but preferably globally. Several studies have aimed to characterize the signature of hypocretin neurons24–29. By combining data from these studies and also taking into account the peripheral tissue expression of each target30, we selected the following seven, potentially NT1-related proteins for our screening: the precursor protein for the hypocretin peptides (HCRT), the hypocretin receptor 2 (HCRTR2), Lim homeobox 9 (LHX9), prodynorphin (PDYN), the precursor protein for the pyroglutamylated RFamide peptides (QRFP), regulatory factor x4 (RFX4), and tribbles homolog 2 (TRIB2) (details listed in Table 2).

Using the NetMHCCons 1.1 in silico prediction algorithm for peptide-MHC class I binding (www.cbs.dtu.dk/services)31, we predicted 9- to 11-mer peptides from these seven selected proteins with the ability to bind to eight different HLA-I molecules. The HLA-I molecules were chosen based on reported associations with narcolepsy in two independent studies10,11, i.e., HLA-A*11:01, HLA-B*18:01, B*35:01, B*51:01, and HLA-C*04:01. Furthermore, HLA-A*02:01, -A*03:01, and HLA-B*07:02 were also included owing to their high prevalence in the available samples. Peptides with %Rank scores of two or below were defined as binders and included in the study, giving a total of 1183 peptides across the seven different proteins and eight different HLAs.

The distribution of peptides was not equal between the different proteins (Fig. 1b). The number of predicted HLA-binding peptides ranged from 36 in HCRT to 379 in RFX4. This is mostly an effect of the size differences between the proteins, ranging from 131 amino acids (aa) to 735 aa (Fig. 1c). Within each different protein, the number of predicted peptides with high affinity for each HLA also varied (Fig. 1d). For some proteins (RFX4, HCRTR2, and TRIB2), the distribution of peptides was fairly even between the eight different HLA types.
Screening for T-cell recognition of NT1-related peptides. The 1183 predicted HLA-binding peptides were synthesized and used to generate individual pMHC monomers using UV-mediated peptide exchange. We then multimerized the pMHC monomers onto a phycoerythrin (PE)-labeled polysaccharide backbone coupled to a DNA barcode that was unique to each specific pMHC. This yielded 1183 different pMHC multimers that were mixed according to donor HLA type and used to stain the relevant NT1 samples and healthy controls. CD8+ T cells with the ability to bind the pMHC multimers were sorted based on a positive PE signal, the associated DNA barcodes were amplified and the specificity of the CD8+ T cells could next be revealed by sequencing of the DNA barcodes. This strategy is depicted in Fig. 2a along with an example of a sorting plot from a healthy donor (#138) and the subsequent analysis of the peptide-specific CD8+ T-cell populations present in this sample. Gating strategy and control sorting of CD8− T cells with a PE-positive signal is shown in Supplementary figure 1 and 2, respectively.

The 1183 peptides included in this study are ligands of eight different HLA molecules. The eight HLA molecules cover the HLA haplotypes of the donor cohort for at least one and up to four HLA molecules per donor. All donors were screened only with the fraction of the library that matched their HLA type. For each protein and HLA type, Fig. 2b shows whether a given sample was screened (gray) or not (white) and whether CD8+ T cells specific toward peptides from the given protein in the given HLA-I molecule were detected (green/blue color gradient). We estimated the frequency of all multimer-positive CD8+ T-cell populations (as previously done in ref. 22) as the fraction of sequencing reads specific for a given pMHC out of total reads, multiplied by the percentage of sorted multimer-binding CD8+...
T cells out of total CD8+ T cells. The color gradient in Fig. 2b indicates the estimated frequency of the given multimer-positive population, if only one, or the sum of frequencies if more than one peptide-specific CD8+ T-cell population was detected in a sample. A full overview of all the multimer-positive populations found in each sample and the peptide sequences that they each recognize is given in Supplementary Figure 3. As can be seen from Fig. 2b, we observed CD8+ T-cell recognition of NT1-related peptides in both NT1 patients and healthy controls. All but one patient carried the NT1-associated HLA class II allele DQB1*06:02, whereas only 23 healthy controls out of 52 carried this allele. For this reason, the healthy controls were divided into two groups depending on their expression of this HLA molecule (Fig. 2b).

Enhanced T-cell recognition in NT1 patients. As some individuals had several CD8+ T-cell populations that recognized different peptides within the same protein in the context of more than one HLA type, we summed up the frequencies of all NT1-related CD8+ T-cell populations per protein in each individual donor (Fig. 3a, color gradient). As the library size used for T-cell screening depended on the HLA expression of the donor, we determined the proportional CD8+ T-cell recognition to correct for any potential biases based on differences in the pMHC library size. The proportional CD8+ T-cell recognition was determined as the fraction of peptides recognized by CD8+ T cells out of the total number of pMHCs used for screening the individual samples (Fig. 3a, circle size). In addition to the 1183 peptides potentially associated with NT1, all the samples were screened with a panel of known virus epitopes as positive controls. Similar to the NT1-relevant peptides, these were also selected to match the donor’s HLA type. For two healthy donors (#109 and #131), there was no overlap with the viral epitope-HLA complexes available and the HLA haplotype of the donors, and these samples were consequently not analyzed for T-cell recognition of viral antigens. The majority of patients (18/20; 90%) and healthy controls (42/50; 84%) displayed T-cell recognition of virus-derived epitopes (Fig. 3a lower panel). Furthermore, these virus-specific CD8+ T-cell populations were generally more frequent than those recognizing NT1-related peptides (Supplementary Figure 4). The virus epitopes were selected from a pool of

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### Table 1 Demographic, clinical, and paraclinical data of the Narcolepsy cohort

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Narcolepsy type 1 N = 20</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender (male), n (%)</td>
<td>9 (45%)</td>
</tr>
<tr>
<td>Age (years), mean (range)</td>
<td>25.6 (7–62)</td>
</tr>
<tr>
<td>Disease duration (years), mean (range)</td>
<td>5.3 (1–9)</td>
</tr>
<tr>
<td>HLA-DQB1*0602- positivity, n (%)</td>
<td>19 (95%)</td>
</tr>
<tr>
<td>CSF hcrt-1 ≤ 110 pg ml⁻¹, n (%)</td>
<td>20 (100%)</td>
</tr>
<tr>
<td>Daytime sleepiness, n (%)</td>
<td>20 (100%)</td>
</tr>
<tr>
<td>Epworth Sleepiness Scale, mean ± SEM</td>
<td>16.8 ± 0.5 N = 16</td>
</tr>
<tr>
<td>Cataplexy presence, n (%)</td>
<td>19 (95%)</td>
</tr>
<tr>
<td>Hypnagogic hallucinations, n (%)</td>
<td>15 (82%)</td>
</tr>
<tr>
<td>Sleep paralysis, n (%)</td>
<td>11 (62%)</td>
</tr>
<tr>
<td>Disrupted night sleep, n (%)</td>
<td>17 (87%)</td>
</tr>
<tr>
<td>MSLT sleep latency (min.) mean ± SD</td>
<td>3.28 ± 1.98 N = 20</td>
</tr>
<tr>
<td>SOREMPs (number) mean ± SD</td>
<td>3.95 ± 0.91 N = 20</td>
</tr>
</tbody>
</table>

MSLT multiple sleep latency test, SOREMPs sleep onset REM periods, CSF hcrt-1 hypocretin-1 levels in the cerebrospinal fluid.

### Table 2 Overview of the protein targets included in the study and potential targets of interest

<table>
<thead>
<tr>
<th>Protein</th>
<th>Full name</th>
<th>Size (amino acids)</th>
<th>Type</th>
<th>Relation to hypocretin neurons/narcolepsy</th>
<th>Reference</th>
<th>Allen brain atlas</th>
<th>Peripheral expression</th>
</tr>
</thead>
<tbody>
<tr>
<td>HCRT</td>
<td>Hypocretin neuropeptide precursor</td>
<td>131</td>
<td>Peptide precursor</td>
<td>Uniquely produced in the neurons</td>
<td>25,26</td>
<td>Specific</td>
<td>Very limited</td>
</tr>
<tr>
<td>PDYN</td>
<td>Prodynorphin</td>
<td>254</td>
<td>Peptide precursor</td>
<td>Missing post mortem in hypothalamus from NT1</td>
<td>25,26</td>
<td>Hypothalamus + other brain areas</td>
<td>Very limited</td>
</tr>
<tr>
<td>QRP</td>
<td>Pyroglutamylated RFamide peptide</td>
<td>136</td>
<td>Peptide precursor</td>
<td>Peptide related to sleep regulation</td>
<td>27</td>
<td>Only in hypothalamus All brain</td>
<td>Low, gut-intestinal tract</td>
</tr>
<tr>
<td>LHX9</td>
<td>Lim homeobox 9</td>
<td>397</td>
<td>Transcription factor</td>
<td>Transcription factor regulating hcrt expression.</td>
<td>25,28</td>
<td>Only in hypothalamus All brain</td>
<td>Very limited</td>
</tr>
<tr>
<td>RFX4</td>
<td>Regulatory factor x4</td>
<td>735</td>
<td>Transcription factor</td>
<td>Unknown but highly specific to hcrt neurons</td>
<td>25</td>
<td>Only in hypothalamus All brain</td>
<td>Yes</td>
</tr>
<tr>
<td>TRIB2</td>
<td>Tribbles pseudokinase 2</td>
<td>343</td>
<td>Cytosolic protein, pseudokinase Receptor, GPCR</td>
<td>Autoantibodies detected in NT1 HCRTR2 is not expressed in hcrt neurons</td>
<td>24</td>
<td>No signal</td>
<td>Low, kidney, heart</td>
</tr>
<tr>
<td>HCRTR2</td>
<td>Hypocretin receptor 2</td>
<td>444</td>
<td>Receptor, GPCR</td>
<td>Autoantibodies detected in NT1 HCRTR2 is not expressed in hcrt neurons</td>
<td>29</td>
<td>No signal</td>
<td>Low, kidney, heart</td>
</tr>
</tbody>
</table>

**Targets not included in the study**

<table>
<thead>
<tr>
<th>Protein</th>
<th>Full name</th>
<th>Size (amino acids)</th>
<th>Type</th>
<th>Relation to hypocretin neurons/narcolepsy</th>
<th>Reference</th>
<th>Allen brain atlas</th>
<th>Peripheral expression</th>
</tr>
</thead>
<tbody>
<tr>
<td>NPTX2</td>
<td>Neuronal pentraxin-2</td>
<td>431</td>
<td>Synaptic protein</td>
<td>Missing in hypothalamus from NT1. Also known as NARP</td>
<td>25,26</td>
<td>Hypothalamus + other brain areas</td>
<td>Medium, adrenal gland High, many organs</td>
</tr>
<tr>
<td>IGFBP3</td>
<td>Insulin-like growth factor-binding protein 3</td>
<td>297</td>
<td>Cytosolic protein</td>
<td>Regulates hypocretin expression</td>
<td>25,26</td>
<td>Hypothalamus + other brain areas</td>
<td>Medium, many organs Low, many organs</td>
</tr>
<tr>
<td>PLAGL1</td>
<td>PLAG1 like zinc finger 1</td>
<td>463</td>
<td>Zinc finger protein</td>
<td>Unknown</td>
<td>24,25</td>
<td>Hypothalamus + other brain areas</td>
<td>Hypothalamus + other brain areas</td>
</tr>
<tr>
<td>NR6A1</td>
<td>Nuclear receptor subfamily 6 group A member 1</td>
<td>480</td>
<td>Nuclear receptor</td>
<td>Regulates hypocretin expression</td>
<td>24,25</td>
<td>Hypothalamus + other brain areas</td>
<td>Hypothalamus + other brain areas</td>
</tr>
</tbody>
</table>

*Data from ref. 26*
peptides known to frequently generate T-cell responses in individuals carrying these common virus infections. Hence, the virus epitopes served merely as a positive control and to test for an overall similar immunological recognition capacity between the different cohorts. Consequently, for these highly selected peptides the proportional CD8\(^+\) T-cell recognition was much higher than seen for the predicted NT1-derived epitopes (Fig. 3a lower panel).
Although T-cell recognition was observed in all three cohorts both toward NT-relevant and virus-derived epitopes, comparison between the three different groups did reveal some interesting immunological characteristics for the NT1-related CD8+ T-cell recognition. When counting the number of different peptide-specific CD8+ T-cell populations found in each donor in the different cohorts, NT1 patients, HLA-DQB1*06:02-positive controls, and HLA-DQB1*06:02-negative controls, no significant difference was observed based on the average number of CD8+ T-cell populations detected (Fig. 3b). However, when evaluating the estimated frequency for each multimer-positive CD8+ T-cell population, we observed that the HLA-DQB1*06:02-positive controls had a significantly lower frequency of such T cells compared with both NT1 patients and DQB1*06:02-negative controls.
controls. NT1-related CD8\(^+\) T-cell populations in the latter two groups had estimated frequencies of similar levels (Fig. 3c). When summing-up the frequencies for all different multimer-positive CD8\(^+\) T-cell populations present in a given donor, there was no difference between the groups (Fig. 3d). We did not observe any differences in the number of protein targets subjected to CD8 T-cell recognition in the three different cohorts. There was no correlation between the duration of disease at the time of T-cell analyses and the number or frequency of NT1-related CD8\(^+\) T-cell populations detected in the NT1 patients (Supplementary Figure 5).

We analyzed the number and estimated frequency of the virus-specific CD8\(^+\) T-cell populations to test if the differences observed between the donor cohorts for NT1-related CD8\(^+\) T-cell recognition also applied to virus-specific CD8\(^+\) T-cell recognition. For the virus-specific CD8\(^+\) T-cell populations no differences were observed, neither in terms of number (Fig. 3e) nor estimated frequencies (Fig. 3f, g). For the summed frequencies, there was a tendency toward a higher frequency of virus-epitope recognition in the NT1 patient group, but this was not statistically significant (Fig. 3g).

The observed difference in the estimated frequency of NT1-related CD8\(^+\) T-cell populations between NT1 patients and HLA-DQB1*06:02-positive controls was more pronounced within certain types of proteins. As is evident from Fig. 3h–j, a significant difference was detected only for transcription factors (LHX9 and RXF4), whereas no difference in CD8\(^+\) T-cell recognition frequency was observed for the peptide precursor proteins (HCRT, PDYN, and QRFP) or the proteins that have previously been identified as targets for autoreactive antibodies (HCRT2 and TRIB2).

**T-cell recognition relates to predisposing HLA-I alleles.** We determined the fraction of the individuals in the three different cohorts carrying CD8\(^+\) T cells specific toward a given NT1-related protein for a given HLA type. Interestingly, the highest percentages of donors harboring NT1-related CD8\(^+\) T-cell populations were found within the groups of donors expressing NT1-associated HLA-I molecules (Fig. 4a). NT1-associated HLA-I molecules are given in red). The same pattern emerged when counting the number of different CD8\(^+\) T-cell populations recognizing each protein-HLA combination, normalized to the total number of possible peptide-HLA specific populations. The total number of possible peptide-specific CD8\(^+\) T-cell populations that could be detected was calculated as the number of samples with a given HLA type times the number of peptides from a given protein predicted for this HLA molecule (Supplementary Figure 6). Here again, the highest frequency of recognition was found with the NT1-associated HLA alleles.

Of the 20 patients included in this study, 9 were positive for an NT1-associated HLA allele and this was the case for 10/23 HLA-DQB1*06:02-positive controls and 15/29 HLA-DQB1*06:02-negative controls. Interestingly, NT1-related CD8\(^+\) T-cell recognition was detected in 9/9 NT1 patients carrying an NT1-associated HLA-I allele, whereas this was only true for 4/11 (36%) of the patients that did not (\(p = 0.005\), Fisher exact test). This was also observed for HLA-DQB1*06:02-negative controls where recognition was detected in 13/15 (87%) and 3/14 (21%) donors with and without an NT1-associated HLA-I allele, respectively (\(p = 0.0007\), Fisher exact test). No significant difference was observed for the HLA-DQB1*06:02-positive controls. It should be noted, however, that donors carrying an NT1-associated HLA allele generally matched more of the HLA class I alleles that we included in our study, than donors not carrying an NT1-associated HLA allele. Consequently, more peptides were evaluated for T-cell recognition in the individuals having an NT1-associated HLA-I allele, on average 399 versus 246 peptides. To account for this difference, the number of peptide-specific CD8\(^+\) T-cell populations in each donor was normalized to the total amount of peptides used to screen a given donor. The recognition percentage for patients (Fig. 4b) and HLA-DQB1*06:02-negative controls (Fig. 4d) was significantly different when stratifying the donors according to their expression of NT1-associated HLA class I alleles, whereas this was not the case for HLA-DQB1*06:02-positive controls (Fig. 4c).

Looking closer at only the CD8\(^+\) T-cell recognition of peptides that were restricted toward an NT1-associated HLA allele, we observed that the number of detected NT1-related CD8\(^+\) T-cell populations differed between the three different donor groups, with HLA-DQB1*06:02-positive controls having significantly fewer of such T-cell populations than NT1 patients and HLA-DQB1*06:02-negative controls (Fig. 4e). As such, 7/9 NT1 patients with NT1-related CD8\(^+\) T-cell recognition, recognized more than one peptide restricted to an NT1-associated HLA allele, whereas this was only the case for 1/6 of the HLA-DQB1*06:02-positive healthy controls. Thus, in this control group 5/6 donors harbored only one NT1-related CD8\(^+\) T-cell population restricted to NT1-associated HLA types.

**Ex vivo and in vitro verification of CD8\(^+\) T-cell specificity.** The detected CD8\(^+\) T-cell populations with specificity toward NT1-related peptides were, in general, of very low frequency and therefore challenging to detect using conventional fluorescently-labeled MHC multimers, in part owing to limited amounts of sample material. However, an NT1-related CD8\(^+\) T-cell population recognizing a peptide from LHX9 was detected and confirmed with MHC multimer staining (Supplementary Figure 7a).
It was not possible to further in vitro culture and expand these NT1-related CD8+ T cells, although virus-specific T cells from both patients and healthy controls could be expanded (Supplementary Figure 7b, c).

Immunogenic hotspots in NT1-relevant proteins. We mapped the location of the possible NT1-related T-cell recognition observed in this study, both for patients and healthy controls. Looking at each protein included in the study, a series of...
immunological hotspots seems to be present in these proteins, characterized by the colocalization of peptide sequences of different HLA restrictions that were identified as immunogenic across the cohort (Supplementary Figure 8).

Discussion
Here we present a comprehensive screening for CD8^+ T-cell recognition of NT1-relevant proteins in peripheral blood mononuclear cells (PBMCs) from well-characterized hypocretin-deficient NT1 patients and healthy controls. We used DNA barcode-labeled MHC multimers to screen for CD8^+ T-cell recognition of a large library of 1183 NT1-relevant peptides. These peptides were restricted to a range of HLA molecules (HLA-A*11:01, HLA-B*18:01, B*35:01, B*51:01, and HLA-C*04:01) based on their previously described association with NT1 and three other HLA alleles (HLA-A*02:01, A*03:01, and HLA-B*07:02) based on their high prevalence in the available samples. The samples were divided into three cohorts: NT1 patients (all but one being HLA-DQB1*06:02-positive), HLA-DQB1*06:02-positive healthy controls, and HLA-DQB1*06:02-negative healthy controls. We observed the expected levels of CD8^+ T-cell recognition of virus-derived epitopes in all three cohorts; no differences were observed in virus recognition frequency or magnitude. In all three cohorts, recognition of possible NT1-associated peptide epitopes was observed, although at a significantly lower level than for virus-derived epitopes. Interestingly, a number of differences in the CD8^+ recognition profile were observed between NT1 patients and the cohort of healthy controls also expressing HLA-DQB1*06:02. We found the estimated frequency of individual NT1-related CD8^+ T-cell populations to be significantly higher in NT1 patients compared with HLA-DQB1*06:02-positive healthy controls. Subgrouping the protein targets revealed that this difference was evident only for the transcription factors LHX9 and RFX4, indicating that these may be important targets for disease development. Furthermore, within the patient cohort, CD8^+ T-cell recognition was observed to a significantly higher fraction of the tested NT1-related peptides (recognition percentage) in the patients carrying NT1-associated HLA class I alleles (HLA-A*11:01, HLA-B*18:01, B*35:01, B*51:01, and HLA-C*04:01) compared with the patients that did not. Importantly, such a difference was not observed for HLA-DQB1*06:02-positive healthy controls. Finally, the NT1 patients carried a higher number of CD8^+ T-cell populations restricted toward NT1-associated HLA types than the HLA-DQB1*06:02-positive controls. This finding supports the role of such HLA-I restriction in disease development and indicates that the presence of these risk-associated HLA class I alleles may enhance the level of CD8^+ T-cell recognition of NT1-related peptides, even affecting those presented by non-risk HLA molecules.

Strikingly, the enhanced level of T-cell recognition observed in NT1 patients compared with healthy controls expressing HLA-DQB1*06:02 was not evident when compared with HLA-DQB1*06:02-negative healthy controls. In fact, for most of the analyses, this cohort resembled the patient cohort. This finding indicates a role for the level of autoreactive CD8^+ T cells, both in terms of the number and estimated frequency of individual peptide-specific populations, combined with HLA-DQB1*06:02 expression as important parameters for development of NT1. It could be speculated that the lack of HLA-DQB1*06:02 expression allows the existence of NT1-associated CD8^+ T-cell recognition without disease development, and that the lower level of CD8^+ T-cell recognition in the presence of HLA-DQB1*06:02 expression prevents disease development. Thus, the role of HLA-DQB1*06:02 in disease initiation should be further elucidated. It is possible, that autoreactive CD4^+ T cells, which have so far almost exclusively been detected in NT1 patients and not healthy controls, are necessary for the function of the autoreactive CD8^+ T cells.

In the recent study by Latorre et al., detecting NT1-relevant autoreactive CD4^+ T cells, only controls expressing HLA-DQB1*06:02 were included. Thus, whether the same differences between HLA-DQB1*06:02-positive and negative controls are evident for autoreactive CD4^+ T cells can only be speculated.

Latorre et al. detect very limited CD8^+ T-cell reactivity in both patient and healthy donor cohorts, when screening for T-cell reactivity toward HCRT. Here, we included a broader spectrum of seven proteins with preferential expression in hypocretin-producing neurons. Although we also observe limited CD8^+ T-cell recognition toward HCRT (in one patient and three HLA-DQB1*06:02-negative controls), the CD8^+ T-cell recognition was substantially larger for the total pool of proteins. The methods used in the two studies differ with regards to the type of T cells that can be detected. Whereas Latorre et al. detected only memory T cells with the ability to proliferate in response to HCRT peptides, we detected all CD8^+ T cells with the ability to recognize and bind to NT1-relevant peptides. We did not include phenotype markers or functional analyses to our screening and can therefore not determine whether the autoreactive CD8^+ T cells found in patients and healthy controls will differ in terms of phenotype or functionality.

Another challenge for the present study was the low level of CD8^+ T-cell recognition, which compromised our ability to further study these autoreactive T cells by tracking and expanding them, e.g., using fluorescently labeled MHC multimers. Consequently, functional analysis of the autoreactive CD8^+ T cells that we detected was not possible in our study, although it would have been very interesting to investigate whether any differences between NT1 patients and healthy controls would have been detected. Furthermore, autoreactive T cells are likely to have lower affinity for their target pMHC than virus-specific T cells, consequently making MHC multimer-based detection difficult. As mentioned earlier, we have previously demonstrated that the DNA barcoding-based strategy is especially valuable for detecting low-affinity T cells, and hence the preferable choice in this case. Despite these limitations, in one case we were able to detect NT1-relevant CD8^+ T cells by a direct MHC multimer staining, demonstrating the presence of such T cells.

A potential limitation in the present study is the fact that patient samples were collected from 1 to 9 years after disease onset. It is possible, that the autoimmune response in NT1 is not strong enough to establish a long-lasting memory response in all patients. We did not observe a correlation between the time from NT1 diagnosis and the number or frequency of detected NT1-related CD8^+ T-cell populations, but it is possible that most CD8^+ T-cell recognition toward hypocretin neurons, will fade over time and be undetectable after 1 year. Hence, screening samples from patients very close to disease onset would be a very interesting next step.

In addition to the finding of autoreactive CD8^+ T cells in NT1 patients, the present study suggests that low levels of autoreactive CD8^+ T cells of relevance for NT1 can be detected in most individuals, independent of their disease status and HLA types. Similar observations have been made in several studies of other autoimmune diseases. Disease relevant autoreactive CD8^+ T cells are found to be present in both multiple sclerosis (MS) patients and healthy controls in several studies of MS33,34 and the same is true for type 1 diabetes.35-37. This points to the presence of an external trigger for the activation of autoreactive CD8^+ T cells as necessary for disease development. In NT1, external triggers, e.g., H1N1 infection or vaccination, could be hypothesized to boost preexisting autoreactive CD8^+ T cells.
In a recent study of autoreactive CD8+ T cells relevant for type 1 diabetes, the frequency and functional capacity of these cells was found to be different between diabetes patients and healthy controls. What did, however, distinguish these two donor cohorts was the presence of autoreactive CD8+ T cells inside the target organ, which was found mainly in the diabetes patients. In the present study we only investigated the presence of NT1-relevant autoreactive CD8+ T cells in blood samples from patients and controls, but this may not reflect the true level and distribution of CD8+ T-cell recognition relevant for NT1. Detection of such T cells in the cerebrospinal fluid (CSF) could be interesting and potentially more relevant as a measure of disease-initiating T cells. In the study by Laterre et al., detection of autoreactive T-cell clones in the CSF was attempted for a few NT1 and NT2 patients, but no such CD4+ or CD8+ T-cell clones were detected in NT1 patients or controls. Two HCRT reactive CD8+ T-cell clones were, however, detected in the CSF from an NT2 patient who was recently diagnosed, again underlining the potential importance of screening for T-cell recognition closer to disease onset.

Some HLA class II DQB1 alleles have been suggested to be protective in NT1. These are 05:01, 06:01, and 06:03, and less strong all DQB1*02 alleles. We speculated whether the presence of these alleles differed between controls with or without CD8+ T-cell recognition. Only 7 of the 23 HLA-DQB1*06:02-positive controls carried a putatively protective HLA class II type. Three of these showed multimarker-positive CD8+ T cells. Among the remaining HLA-DQB1*06:02-positive controls that were not protected by their HLA class II type, nine individuals still had multimarker-positive CD8+ T cells. Furthermore, six of the 20 NT1 patients in this cohort carried a protective HLA class II type, indicating that the protective effect of these HLA types is likely only minor.

In this study, we find that NT1 patients have enhanced CD8+ T-cell recognition of NT1-related proteins compared with HLA-DQB1*06:02-positive healthy controls, both in terms of the frequency and the breadth of their CD8+ T-cell recognition. Although the observation of a higher level of autoreactive CD8+ T cells in patients, compared with HLA-DQB1*06:02-positive healthy controls, points to some involvement of CD8+ T cells in disease development, the autoimmune nature of NT1 is still not indisputably proven. CD8+ T cells reside in a complex interplay between many different cells of the immune system, and further studies are needed to fully elucidate the etiology of NT1.

### Methods

**Patient and healthy control material.** Twenty ICSD-323 diagnosed Type 1 narcolepsy patients (age range: 7–62 y, mean age: 25.6 y; 9 male) were selected for blood donation from the known patient cohort at the Danish Centre for Sleep Medicine. Inclusion criteria were hypocretin de

**PBMC extraction.** To collect PBMCs, a blood donation (450 ml) was obtained via the blood bank of Rigshospitalet, Denmark. The blood donations from the Danish blood bank were transported to the laboratory the same day and PBMCs were fractionated into plasma,uffy coat, and red blood cells. PBMCs from theuffy coat were next extracted using Ficoll gradient reagent, frozen in 10% dimethyl sulfoxide (DMSO) ( Sigma) in fetal bovine serum (Gibco), and cryopreserved at −140 °C until use.

### Peptide prediction analysis

The online prediction algorithm NetMHCCons 1.1 (http://www.cbs.dtu.dk/services/NetMHCcons/), was used to predict 9–11-mer MHC class I T-cell epitopes within seven different protein sequences associated with hypocretin neurons. Peptides were predicted from the following proteins: HCRT (O43612–1), TRIB2 (3 variants Q92519–1, F8W18A, B5CMX4), cox5a (10 variants Q3ES49–1, Q3ES94–2, Q3ES94–3, Q3ES49–4, R4GM3J, F8VKD4, F8VZC4, B4DBZ7, F8WT19, F8VX50), PDYN (P01213–1), LHDCX (8 variants Q9NNQ6–1, Q9NNQ6–2, Q9NNQ6–3, Q9NNQ6–4, HYOTL5, HOY330, A0A087X083, A0A04DAG4Y), QRFP (P38597–1), and HCRTR2 (O43614–1).

Eight different HLA alleles were included in the analysis: HLA-A*0201, HLA-A*0301, HLA-B*0702, HLA-A*1101, HLA-B*1801, HLA-B*3501, HLA-B*5101, and HLA-C*0401. This yielded a total of 1183 predicted T-cell epitopes spread on the eight different HLA alleles and seven different proteins. The peptides were synthesized and purchased from Pepscan (Pepscan Presto, The Netherlands) and dissolved to 10 μM in DMSO.

### HLA analysis across the cohort

All donors included in the study were HLA- and MHC encoded generation sequencing of the full exomes of HLA class I A, B, and -C. The full exome of HLA class II DQA was also sequenced, but only exon 2 and 3 of HLA-DQB1 were sequenced (GenDx, Netherlands). MiSeq from Illumina was used for all sequence analyses. The full HLA type of each donor is presented in Supplementary Table 1. The highlighted HLA subtypes are associated with an increased risk of NT110,11.

### Generation of DNA barcodes and dextran conjugation

Oligonucleotides containing distinct 25mer nucleotide sequences (from Xu et al. 2009) were purchased from LGC Biosearch Technologies (Denmark). All oligos carried a 6-nt unique molecular identifier. Oligonucleotides modified with a 5’ biotin tag (oligo A) were joined to unmodified, partially complementary oligonucleotides (oligo B) to generate a > 1000 dextran double-stranded AxBy DNA barcodes. Combining a and B oligos (one of each) were mixed with 5 μmol Sequense Reaction Buffer mix (PN 70702, Affymetrix) to final concentrations of 26 μmol (Oligo A) and 52 μmol (Oligo B), respectively; heated to 65 °C for 2 min; and allowed to anneal by cooling slowly to <35 °C over 15–30 min. The annealed oligo As and Bs were elongated to create double-stranded AxBy DNA barcodes by adding Sequense polymerase (70775Y, Affymetrix), 20 μmol DTT and 800 μM or 72 μM dNTPs, followed by incubation for 5–10 min at RT. Elongated AxBy barcodes were diluted in nuclease-free water +0.1% Tween to 2.17 μM (with respect to the A oligo) and stored at 20 °C.

Attachment of 5’ biotinylated AxBy DNA barcodes to PE- and streptavidin-conjugated dextran (provided by Innudex, Denmark) was performed by mixing the two components at final concentrations of 14 × 10^–8 μmol dextran backbone and 2.8 × 10^–3 μmol in order to obtain ~2 barcodes for each dextran backbone and subsequent incubation for 30 min at 4 °C.

### Generation of pMHC-I multimers labeled with DNA barcode

A total of 1183 pMHC complexes were generated through UV-mediated conditional ligand exchange. In short, HMC monomers carrying UV-sensitive ligands were mixed with matching peptides at a final concentration of 50 μg ml−1 monomer and 100 μM peptide and exposed to UV light for 60 min. The pMHC complexes were then coupled to DNA barcode- and PE-labeled dextran backbones to a final concentration of 35 μg ml−1 monomer and 4.2 × 10^–8 μM barcode- and PE-labeled dextran backbone and incubated for 20 min on ice. Finally a 15x freeze buffer was added to reach phosphate-buffered saline (PBS) +0.5% bovine serum albumin (BSA) +100 μg ml−1 herring DNA +2 μM ethylenediaminetetraacetic acid (EDTA) +5% glycerol and 995 μM biotin and after 20 min on ice, the pMHC multimers were stored at −20 °C until use.

### T-cell staining with barcode-labeled MHC multimers

Cryopreserved cells were thawed, washed twice in Roswell Park Memorial Institute (RPMI) +10% fetal calf serum (FCS) and then washed in barcode-cytometry buffer (PBS +0.5% BSA +100 μg ml−1 herring DNA +2 μM EDTA). On the day of staining, barcode-labeled MHC multimers were thawed on ice, centrifuged for 5 min at 3300 g, and 1 μM according to 0.043 μM of each distinct PMA from the top of each well, to avoid any potential aggregates, and pooled according to the HLA type of the donors. The volume of the reagent pool was reduced by ultrafilteration to obtain a final volume of ~80 μL of MHC multimers. Centrifugal concentrators (Vivaspin 6, 100,000 Dalton, Sartorius) were saturated with BSA before use. Following ultrafiltration, the pool of multimers was spun at 10,000g for 2 min to sediment potential aggregates. An aliquot of −5 μL of the MHC multimer reagent pool was stored at −20 °C for baseline analysis. Prior to staining...
with the MHC multimers, 50 ns dasatinib was added to up to 10 × 10^6 cells. The MHC multimer pool was then added to the cells and these were incubated for 15 min at 37 °C in a total volume of 80 μl. Following incubation, the cells were stained with an antibody mix containing CD8-BV480 (BD 566121, clone RPA-T8), CD4-FITC (BD 345768) (final dilution 1/80), CD14-FITC (BD 345784) (final dilution 1/32), CD19-FITC (BD 345776) (final dilution 1/40), CD56-FITC (BD 345777) (final dilution 1/100), and a dead cell marker (LIVE/DEAD Fixable Near-IR; Invitrogen L10119) (final dilution 1/1000) and incubated for 30 min at 4 °C. Cells were washed three times in barcode-cytometry buffer and fixed in 1% paraformaldehyde (PFA) for 0.5–24 h before they were washed twice and resuspended in barcode-cytometry buffer. Cells were acquired within a week after multimer staining.

Sorting of MHC multimer-positive T cells. Multimer-binding CD8 T cells were sorted on a FACSAriaFusion (BD) into BSA saturated tubes containing 100 μl of barcode-cytometry buffer. Using FACSdiva software, we gated on single, live, CD8-positive, and dump channel (CD4, 14, 16, 19, and 40)-negative lymphocytes. The binding strategy is exemplified in Supplementary Figure 1. As tested and described in Bentzen et al., inclusion of CD8-positive, multimer negative cells in the sorting gate does not have an impact on the final results because the fluorescence signal is used only for sorting out the relevant cells. Determination of antigen specificity is done solely based on the DNA barcode. The sorted cells were centrifuged for 1 min at 8000 × g and the supernatant was removed. The cell pellet was stored at −80 °C in a minimal amount of residual buffer (< 20 μl).

DNA barcode amplification. DNA barcode amplification was performed using Taq PCR Master Mix Kit (Qiagen, 201443) and 3 μl of forward and reverse primers (LGCh Biosearch Technologies). PCR amplification was conducted on sorted multimer-binding T cells (in < 20 μl of buffer) and on the stored aliquot of the MHC multimer precipitation pool (diluted 50,000 × in the final PCR) under the following conditions: 95 °C 10 min; 36 cycles: 95 °C 30 s, 60 °C 45 s, 72 °C 30 s, and 72 °C 4 min. The multimer reagent pool was used as the baseline to determine the number of DNA barcode reads within a non-processed MHC multimer reagent library. PCR products were purified with a QIAquick PCR Purification Kit (Qiagen, 28104) and the amplified DNA barcodes were sequenced at Sequetech (USA) using an Ion Torrent PGM 316 or 318 chip (Life Technologies).

Processing of sequencing data from DNA barcodes. Sequencing data were processed by the software package "Barracoda," available online at (http://www.cbs.dtu.dk/services/barracoda)22. This tool identifies the barcodes used in a given experiment, assigns sampleID and pHMC specificity to each barcode, and counts the total number of reads and clonally reduced reads for each pHMC-associated DNA barcode. Furthermore, it accounts for barcode enrichment based on methods designed for the analysis of RNA-seq data, implemented in the R package edgeR: specifically, log, fold changes in read counts mapped to a given sample relative to the mean read counts mapped to triplicate baseline samples are estimated using normalization factors determined by the trimmed mean of M-values method.23 Barcodes with fold change with a p < 0.01, which equals a false discovery rate < 0.1 estimated using the Benjamini–Hochberg method, and with clonal read counts above 100 were considered to represent true CD8+ T-cell recognition. The cutoff of 100 reads was chosen based on evaluation of this particular data set, in order to ensure that background events were not included in the analysis. We have previously tested sorting and sequencing of the CD8 negative, dump channel positive cells and did not detect any CD8+ T-cell recognition in this population (Supplementary Figure 2a). Also, the inclusion of high-levels of MHC complexes that were generated with non-matching peptides under UV-induced peptide exchange, hence producing unstable, partly defocused complexes, did not substantially impact the background of the MHC multimer staining (Supplementary Figure 2b).

Verification of specificity with flow cytometry. A small cohort of 40 selected peptides, to which specific T cells had been detected either in several donors or with a high number of reads in single donors, were used to generate fluorescently-labeled MHC multimers. Again, UV-mediated peptide exchange was used and pHMC complexes were multimerized using streptavidin-conjugated PE (405204, Biolek-gend), APC (405207, BioLegend), PE-Cy7 (405206, BioLegend), PE-CF594 (562284, BD), BV421 (563259, BD), BV605 (563260, BD), BV650 (563855, BD), BV403 (564176, BD), and BV747 (564293, BD). The final concentrations of monomer and peptide were 100 μg ml−1 and 200 nM, respectively. Each pMHC was made in two different colors and mixed before the MHC multimers were stained at −20 °C with 0.5% bovine serum albumin (Sigma Aldrich) and 5% glycerc (Fluka). For ex vivo T-cell staining, samples from cells were thawed as described above and 2 × 10^6 cells were stained with 1 μl of each pHMC, according to HLA type, for 15 min at 37 °C. Cells were then stained with a 5× antibody mix as described for staining with DNA barcode-labeled MHC multimers. After two washing steps and fixation in PFA, cells were acquired on an LSR Fortessa (BD). The gating strategy is identical to the one described above and presented in Supplementary Figure 1.

Enrichment of specific T cells. Six donors (three patients and three healthy controls) with detected NTI-relevant CD8+ T-cell recognition were selected for further specific T cells. PE-labeled MHC multimers were generated in the same fashion as described for the verification. The cells were thawed and washed twice in RPMI + 10% FCS and 1 × 10^6 cells were taken to a new tube and irradiated at −20 Gy for 6 min and 15 sec. These cells were washed twice and kept for later use as feeder cells. From the remaining cells, 20 × 10^6 cells were stained with 1 μl per pHMC of the PE-labeled MHC multimers for 60 min at 4 °C. Following staining, the cells were washed twice and stained with α-PE beads (Miltenyi Biotec) for 15 min at 4 °C. The cells with where then washed twice again and loaded onto a magnetic column placed in the magnetic field of a MACS separator (Miltenyi Biotec), which captured the cells with a PE label through the α-PE beads. After washing the column twice, the column was removed from the separator and the specific cells were washed into a new tube containing X-vivo media (Lonza) + 5% human serum (HS; Sigma Aldrich). The specific cells were counted, washed, and cultured in X-vivo + 5% HS supplemented with 23.8 U ml−1 IL-15 (PeproTech, #200–15–10UG) and 100 U ml−1 IL-2 (PeproTech, #200–02–50UG). Approximately 5000 specific cells were cultured with 50,000 feeder cells/well as well as 11,000 CD3/CD8 Dynabeads (Gibco). After 1.5 weeks of culturing, 2 × 10^6 cells were stained with the above-described fluorescently labeled MHC multimers and acquired on an LSR Fortessa (BD).

Graph generation and statistical data analysis. Graphs were generated using either GraphPad Prism 7.0d for mac or ggplot2 in R and edited using Omnigraffe version 7.8.1. Statistical analysis of DNA barcoding data was performed using the software package "Barracoda" as described above. Significant differences between the number and frequency of detected NTI-related CD8+ T-cell populations in Figs. 3, 4, and Supplementary Figure 4 were calculated in GraphPad Prism 7.0b. For Fig. 3, and Supplementary figure 4, log transformation of data was performed, and parametric tests used to determine significant differences, whereas for Fig. 4 non-parametric tests were used. Normal distribution was tested with D´Agostino normality test. Fisher exact tests were used for contingency analysis of over- or under-representation of detected recognition within the subgroups of donors. The single patient (14) which is HLA-DQB1*06:02 positive is removed from comparisons between patients and HLA-DQB1*06:02-positive controls. For all figures, *: p < 0.05, **: p < 0.01, ***: p < 0.001, ****: p < 0.0001.

Ethical approval. All donor material was collected with the approval of the Scientific Ethics Committee of the Capital Region, Denmark (Protocol H–3–2013–054) and written informed consent was obtained according to the Declaration of Helsinki.

Code availability. All relevant code is available from the authors. For DNA barcode analysis, the tool Barracoda is available online at http://www.cbs.dtu.dk/services/barracoda.

Data availability. The data that support the findings of this study are available from the corresponding author upon reasonable request.

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