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Published in:
Journal of Biomedical Optics

Link to article, DOI:
10.1117/1.JBO.22.3.037003

Publication date:
2017

Document Version
Publisher's PDF, also known as Version of record

Link back to DTU Orbit

Citation (APA):

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Abstract. We report on highly accurate femtosecond (fs) laser micromachining of a compound parabolic concentrator (CPC) fiber tip on a polymer optical fiber (POF). The accuracy is reflected in an unprecedented correspondence between the numerically predicted and experimentally found improvement in fluorescence pickup efficiency of a Förster resonance energy transfer-based POF glucose sensor. A Zemax model of the CPC-tipped sensor predicts an optimal improvement of a factor of 3.96 compared to the sensor with a plane-cut fiber tip. The fs laser micromachined CPC tip showed an increase of a factor of 3.5, which is only 11.6% from the predicted value. Earlier state-of-the-art fabrication of the CPC-shaped tip by fiber tapering was of so poor quality that the actual improvement was 43% lower than the predicted improvement of the ideal CPC shape.© The Authors. Published by SPIE under a Creative Commons Attribution 3.0 Unported License. Distribution or reproduction of this work in whole or in part requires full attribution of the original publication, including its DOI. [DOI: 10.1117/1.JBO.22.3.037003]

Keywords: fiber optics sensors; polymers; blood or tissue constituent monitoring; nonimaging optics.

1 Introduction

Fluorescence-based polymer optical fiber (POF) glucose sensors for continuous glucose monitoring have significant potential for optimum control of diabetes. One limitation of fluorescence-based sensors is that the detected fluorescence signal is often weak, which results in low signal-to-noise ratio, and significantly compromises the sensor performance. This problem can be overcome in fiber-optical fluorescence sensors by shaping the fiber tip to increase the numerical aperture (NA) of the fiber thereby collecting more of the fluorescence signal, as demonstrated by Gao et al. by linear and step-wise tapering of silica fibers. In our previous work on POF-based glucose sensors, we have shaped the POF tip as a compound parabolic concentrator (CPC), which is a well-known profile that provides optimal light pickup efficiency, as demonstrated in solar energy systems for thermal and photovoltaic applications and for coupling light emitting diode (LED) light into fibers.

In our original work, CPC tips were fabricated by parabolically tapering of the POF using the heat-and-pull method and demonstrated to increase the NA and fluorescence light pickup efficiency of the fiber. However, the CPC fiber tips fabricated by the heat-and-pull method were not geometrically close to the ideal CPC shape, as shown in Fig. 1, but had imperfections, such as an increase in the local diameter at the start of the CPC tip. Such imperfections led to a 43% reduction in the maximum fluorescence that could have been achieved by the ideal CPC shape, as predicted by Zemax modeling. To take full advantage of the CPC to achieve maximum increase in the fluorescence pickup efficiency, an alternative approach is needed to allow better control and make fiber tips with a shape closer to the ideal CPC shape. One such powerful technique to shape fiber tips is femtosecond (fs) laser micromachining. fs lasers have been used for micromachining and for fabricating fiber Bragg gratings, optical waveguides, and several photonics devices in transparent materials, such as glass. Several groups have also used fs lasers for micromachining in polymers, to write POF FBGs and fabricate photonic devices. fs lasers have the ability to precisely structure different materials in three dimensions, which make it a powerful technique to micromachine and shape the optical fiber tips.

In this paper, we report for the first time the fs laser micromachining of a CPC shaped POF tip with a shape that is close to the ideal CPC, as shown in Fig. 1. We further demonstrate that the fs laser machined high-quality CPC tip gives an improvement of the pickup efficiency of a factor of 3.5, as compared to the corresponding plane-cut fiber tip. This is only an 11.6% deviation from the increase a perfect CPC shape of that length should give. The previous state of the art in pickup efficiency improvement with a CPC tip was 43% lower than the predicted optimum.

2 Femtosecond-Laser Micromachining of Compound Parabolic Concentrator

The CPC in two-dimensional geometry is formed by two identical and oppositely aligned parabolas truncated at their focal point, as shown in Fig. 2(a). The CPC profile is traced by the coordinates y and z, as defined in Ref. and given by

\[ y = \frac{2a_1(1 + \sin \theta_i) \sin(\phi - \theta_i)}{1 - \cos \phi} - a_1; \]

\[ z = \frac{2a_1(1 + \sin \theta_i) \cos(\phi - \theta_i)}{1 - \cos \phi}, \]

where \( a_1 \) and \( a_2 \) are the radii of the input and output apertures, \( \theta_i \) is the acceptance angle of the CPC, and \( \phi \) is an angle that varies...
from 2θ to 90 deg + θ. The geometrical parameters of the CPC are related through the following equation:

\[ a_1 = a_2 \sin \theta_1; \quad L = (a_1 + a_2) / \tan \theta_1, \quad (2) \]

where \( L \) denotes the CPC length. A CPC conserves the entrance angle \( a_1, NA_1 = a_2 NA_2 \) which means that the NA at the output is increased due to the reduced output aperture. Therefore, the CPC tip can increase the fluorescence pickup efficiency in POF-based glucose sensors, as reported in our previous work\(^6\) for a 35-mm long sensor.

The fiber used to manufacture the CPC tip is the Super ESKA fiber with a polymethyl methacrylate (PMMA) core and polyvinylidene difluoride (PVDF) cladding. The core diameter is 240 μm and the outer diameter is 250 μm. The refractive index of the core and cladding is \( n_{\text{core}} = 1.49 \) and \( n_{\text{clad}} = 1.40 \), respectively, which results in an NA of 0.5. It is known from Ref. 9 that all of the light collected by the CPC can be coupled perfectly to the straight part of the fiber only if the acceptance angle of the CPC (\( \theta_i \)) is within the acceptance angle of the straight fiber (\( \theta_{\text{max}} \)), i.e., if \( \theta_i \leq \theta_{\text{max}} \). The CPC acceptance angle is then \( \theta_i = \theta_{\text{max}} = \cos^{-1}(n_{\text{clad}} / n_{\text{core}}) \). The CPC output aperture \( 2a_2 = 240 \mu m \) and the CPC length \( L = 445 \mu m \). These CPC parameters were used in our previously defined Zemax model of the sensor,\(^6\) which showed that the ideal CPC shape of the tip would increase the detected fluorescence by a factor of 3.96 compared to the plane-cut fiber tip.

Before micromachining this fiber tip, the fiber PVDF cladding was removed using triethyl phosphate amide (solvent for PVDF),\(^26\) for a length of 450 μm, which was the closest we could get to the ideal 445 μm. The fiber was then machined using the fs-laser setup shown in Fig. 2(b). The setup consists of an fs-laser system (HighQ laser femtoREGEN) operating at 517 nm, with a 250-fs pulse duration, 380-nJ pulse energy, and 100-kHz repetition rate. The fiber was mounted as shown in Fig. 2 on an air bearing translation system (Aerotech) for a nanometer accurate two-axis motion in the \( x \)- and \( y \)-directions and the laser beam was focused from above using a long working distance 50× objective (Mitutoyo). The CPC shape is then carved out from the top-down in 63 steps of 7 μm in the direction of the fiber axis, as shown in Fig. 2(c), with the first tip diameter set at 82 μm. For each step, the carving was undertaken in circles in a number of rounds given by the laser beam width (2 μm) and the desired tip diameter, increasing the radius of the circle by 1 μm each round. The increase in tip diameter was made such as to follow Eq. (2).

### 3 Glucose Sensor

The fs-laser micromachined CPC POF tips were tested in the POF-based glucose sensor, whose sensor principle is detailed in Fig. 3. The glucose sensor, which is the same as the one also used in our previous work,\(^6\) uses Förster resonance energy transfer (FRET)\(^27\) as the sensing mechanism. FRET is a radiationless energy transfer between an excited donor fluorophore and a proximal ground state acceptor (a fluorophore or a dye) through a long range dipole–dipole interaction and can be used to measure intermolecular distances in the range of angstroms (10 to 100 Å).\(^27\)

The assay chemistry of the sensor is filled in a compartment, which is formed by bonding a glucose permeable membrane onto the fiber. Glucose molecules enter into the assay through the permeable membrane, when the sensor is in place in the glucose solution. The assay chemistry consists of a glucose binding protein labeled with a fluorophore (Alexa fluor 594; \( \lambda_{\text{excitation}} = 590 \text{nm}, \lambda_{\text{emission}} = 618 \text{nm} \)) and a glucose analog (green chain) labeled with crystal violet dye (blue dots). Together they make an FRET pair resulting in the quenching of the fluorophore, thus reducing the emitted fluorescence intensity. When the glucose molecules enter the chemistry, they compete with the glucose analog molecules to attach to the protein. This competition results in breaking the FRET pair and changing their quantity.

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**Fig. 1** (a) Ideal CPC shape. (b) Nonoptimal CPC fabricated by the heat-and-pull method. (c) fs laser micromachined CPC close to the ideal shape.

**Fig. 2** (a) Geometry of a CPC. (b) fs laser setup with stages to micromachine the fiber tip. (c) Illustration of carving process.
in the chemistry. When equilibrium is reached, the reduced number of FRET pairs can be detected as an increase in the overall intensity of the fluorescence collected by the fiber. Therefore, the glucose concentration can be correlated to the detected fluorescence and used to measure the glucose concentration.

The sensor chemistry also contains a reference fluorophore (Alexa fluor 700; $\lambda_{\text{excitation}} = 633 - 647$ nm, $\lambda_{\text{emission}} = 723$ nm), which is unaffected by the glucose concentration. This is to eliminate any unwanted fluctuations in the detected fluorescence intensity caused either by the fluctuations in the light source used to excite the assay or in the coupling between the assay and the light source. The ratio between the assay and the reference fluorescence determines the absolute glucose concentration.

The sensors are characterized by an epifluorescence setup and a fiber-optical spectrometer (USB2000+, Ocean Optics, Florida), as shown in Fig. 3(c). In the setup, the light from the LED source (HLMP-EL30-MQ000), with central wavelength 590 nm, passes through a 55-nm excitation filter centered at 560 nm and then through a beam splitter to finally be coupled to the fiber by a lens, in order to excite the assay chemistry. The resulting fluorescence from the assay is collected by the fiber and passes through the beam splitter and a long-pass emission filter with 610-nm cut-off wavelength to finally reach the spectrometer.

### 4 Results and Discussions

The final fs laser machined CPC shape is shown in Fig. 4(a). To make the surface of the CPC smoother, it is dipped into dibromomethane, a solvent for PMMA, for 20 s and then cleaned with distilled water.

The fluorescence coupling efficiency of the CPC-tipped and plane-cut fiber sensors were both characterized in a dummy sensor configuration, using a miniature cuvette made of a nonfluorescing transparent Tygon tube sealed in one end. The glucose chemistry is filled into the tube and the 35-mm long fiber is inserted into the tube [see Fig. 3(c)], and characterized using the optical setup described in Sec. 3. To avoid any effect of misalignment and coupling variations from the setup, three measurements are taken for each sensor, each using a new dummy sensor.

The detected average spectra from the CPC-tipped and plane-cut fiber sensors are shown in Fig. 4(b); each has been averaged over three measurements. We define the increment factor as the ratio of the average spectral intensity of the CPC-tipped sensor and the plane-cut sensor at 618 nm, which is where the peak of the fluorescence spectrum is located. The experimental results showed that the increment factor achieved with the CPC tip is 3.5. Numerical modeling with Zemax, using the ideal shape of the CPC with the same length and input and output diameters, predicts an increment factor of 3.96.

The question is of course whether the Zemax predicted value of 3.96 is to be trusted, i.e., can it all be reached. After all, it uses several assumptions, such as one-scatter event per ray. To demonstrate the accuracy of the Zemax model, the actual, fabricated

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**Fig. 3** (a) Real glucose sensor. (b) Illustration of the assay chemistry and FRET sensing principle resulting in low and high intensity. (c) Optical setup for sensor characterization.

**Fig. 4** (a) From top to bottom: fs-laser micromachined CPC, ideal CPC, and reconstructed CPC shape in Zemax. (b) Average fluorescence spectra from plane-cut and micromachined CPC tips, each averaged over three measurements.
CPC shape is reconstructed in Zemax using freeform optics. The ideal and actual reconstructed CPC forms are compared in Fig. 4(a), which clearly demonstrates the accuracy of the fs-laser micromachining in generating the desired shape. Zemax calculations with the reconstructed actual fabricated shape then gave an increment factor of 3.62. The fact that this Zemax obtained values close to the experimentally found 3.5 allows us to conclude that the fs-laser micromachined CPC-tipped sensor has indeed an improved pickup efficiency close to the ideal value. It also allows us to trust that the predicted ideal value of 3.96 should be reachable with further optimization.

5 Conclusion
We have micromachined a CPC-shaped POF tip for enhancing the fluorescence pickup efficiency of fluorescence-based fiber-optical biosensors, such as the glucose sensor we have demonstrated here. The micromachined CPC tip showed an increment factor of 3.5 which is in close agreement with the Zemax predicted value of 3.62 obtained using the reconstructed manufactured CPC shape. This allowed us to conclude that the CPC shape obtained using the fs-laser micromachining is close to the ideal CPC shape and thus this method can be used to fabricate high-quality CPC tips in a controlled and precise manner. In terms of further improvement of the increment factor, it is important to note that Zemax predicts a value that is 0.12 higher than the measured value. In our similar comparison for CPC tips fabricated by heat-and-pull tapering using a soldering iron, we measured an increment factor of 2.1 where, using the reconstructed actual fabricated CPC shape, Zemax predicted a factor of around 2.3, so 0.2 higher. First, this shows that fs-laser micromachining is much more accurate that the heat-and-pull tapering. Second, it shows that it would be reasonable to assume that an increment factor of about 3.8 could be achieved with further optimization, since this is a reasonable 0.16 below the Zemax predicted ideal increment factor of 3.96.

Disclosures
Authors declare that there is no conflict of interest regarding the publication of this paper.

Acknowledgments
The research leading to these results has received funding from the People Programme (Marie Curie Actions) of the European Union’s Seventh Framework Programme FP7/2007-2013/under REA Grant Agreement No. 608382.

References

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