Danger in the reef
proteome, toxicity, and neutralization of the venom of the olive sea snake, Aipysurus laevis

Laustsen, Andreas Hougaard; Gutiérrez, José Maria; Redsted Rasmussen, Arne; Engmark, Mikael; Gravlund, Peter; Sanders, Kate L.; Lohse, Brian; Lomonte, Bruno

Published in:
Toxicon

Link to article, DOI:
10.1016/j.toxicon.2015.07.008

Publication date:
2015

Document Version
Peer reviewed version

Link back to DTU Orbit

Citation (APA):
Danger in the reef: Proteome, toxicity, and neutralization of the venom of
the olive sea snake, *Aipysurus laevis*

Andreas H. Laustsen¹, José María Gutiérrez², Arne R. Rasmussen³, Mikael Engmark⁴,
Peter Gravlund⁵, Kate L. Sanders⁶, Brian Lohse¹, Bruno Lomonte²

¹ Department of Drug Design and Pharmacology, Faculty of Health and Medical
Sciences, University of Copenhagen, Denmark
² Instituto Clodomiro Picado, Facultad de Microbiología, Universidad de Costa Rica,
San José, Costa Rica
³ Royal Danish Academy of Fine Arts, School of Conservation, Denmark
⁴ Department of Systems Biology, Technical University of Denmark, Denmark
⁵ National Aquarium of Denmark (Den Blå Planet), Denmark
⁶ School of Earth & Environmental Sciences, University of Adelaide, Australia

**Running title:** *Aipysurus laevis* venom proteomics

**Keywords:** *Aipysurus laevis*; olive sea snake; snake venom; proteomics; toxicity;
venomics

**Address for correspondence:**
Dr Bruno Lomonte
Professor
Instituto Clodomiro Picado
Facultad de Microbiología
Universidad de Costa Rica
San José, COSTA RICA
bruno.lomonte@ucr.ac.cr
Highlights

- The venom proteome of the olive sea snake, *Aipysurus laevis*, is presented
- Most abundant venom components are phospholipases A₂ and short neurotoxins
- Lethality screening coupled to abundance estimation identified the medically relevant toxins
- BioCSL Sea Snake Antivenom neutralizes venom lethality
- ICP Anti-Coral Antivenom cross-recognizes phospholipases A₂ from *A. laevis* venom
Abstract

Four specimens of the olive sea snake, *Aipysurus laevis*, were collected off the coast of Western Australia, and the venom proteome was characterized and quantitatively estimated by RP-HPLC, SDS-PAGE, and MALDI-TOF-TOF analyses. *A. laevis* venom is remarkably simple and consists of phospholipases A2 (71.2%), three-finger toxins (3FTx; 25.3%), cysteine-rich secretory proteins (CRISP; 2.5%), and traces of a complement control module protein (CCM; 0.2%). Using a Toxicity Score, the most lethal components were determined to be short neurotoxins. Whole venom had an intravenous LD_{50} of 0.07 mg/kg in mice and showed a high phospholipase A2 activity, but no proteinase activity *in vitro*. Preclinical assessment of neutralization and ELISA immunoprofiling showed that BioCSL Sea Snake Antivenom was effective in cross-neutralizing *A. laevis* venom with an ED_{50} of 821 μg venom per mL antivenom, with a binding preference towards short neurotoxins, due to the high degree of conservation between short neurotoxins from *A. laevis* and *Enhydrina schistosa* venom. Our results point towards the possibility of developing recombinant antibodies or synthetic inhibitors against *A. laevis* venom due to its simplicity.

(176 words)
1. Introduction

The viviparous sea snakes are a diverse clade of more than 60 species that are phylogenetically nested within the front-fanged Australo-Melanesian terrestrial elapids (Hydrophiinae) (Rasmussen et al., 2011). They are highly aquatic and occupy most shallow-marine habitats throughout the tropical and subtropical Indo-West Pacific, yet are estimated to share a common ancestor dated at only 6–8 million years ago (Sanders et al., 2008; Lukoschek et al., 2012). The amphibious sea kraits (Hydrophiinae: Laticauda) represent an independently aquatic and earlier diverging lineage that is the sister to terrestrial and viviparous marine hydrophiines (Keogh, 1998; Scanlon and Lee, 2004; Sanders et al., 2008). Two major clades are recognised within the viviparous marine group: An ‘Aipysurus’ lineage comprising ten species found primarily in the Australo-Papuan region, and a ‘Hydrophis’ lineage containing at least 50 species distributed throughout the Indo-West Pacific (Rasmussen et al., 2011).

In the Aipysurus group, the olive sea snake, Aipysurus laevis, has a large muscular head and is the most robustly built and longest species recorded, reaching more than 170 cm in total length (Smith, 1926; Cogger, 1975). A. laevis has been recorded from Aru Archipelago and Kai Islands (Indonesia) in the west and from the northern coast of Australia and southern coast of New Guinea (Timor Sea and Arafura Sea) to New Caledonia in the east (Coral Sea) (Cogger, 1975; Ineich and Rasmussen, 1997; Sanders et al., 2014). A. laevis is found in shallow marine habitats – coral reefs as well as sandy, rocky, and mud-bottom habitats, and is often one of the most abundant species throughout its range (Cogger, 1975; Lukoschek et al., 2007; Sanders et al., 2014). It hunts primarily in crevices on the sea floor, and the following fish families have been found as prey items in A. laevis: Acanthuridae, Apogonidae, Carangidae, Clupeidae, Engraulidae, Labridae, Lutjanidae, Pempheridae, Pomacentridae, Scaridae,
Scorpaeidae and Serranidae (McCosker, 1975; Voris and Voris, 1983). Fish eggs, crabs, shrimp and pelecypod (Limiidae) have also been found in stomach content (McCosker, 1975; Voris and Voris, 1983).

During mating season A. laevis is more prone to defensive attacks than at other times of the year (Heatwole, 1975). However, normally A. laevis will ignore a diver even if the diver approaches quite close (Heatwole, 1975). A. laevis has up to at least 5 mm long fangs and the venom is known for being extremely toxic (Limpus, 1978; Minton, 1983; Mackessy and Tu, 1993; Greer, 1997). A. laevis is commonly caught as by-catch, and commercial trawler fishers and recreational fishers handling nets are therefore the typical bite victims of A. laevis.

The venoms of sea snakes, typically containing α-neurotoxins and phospholipases A₂ (PLA₂s), are known to be generally more potent than the venoms from terrestrial snakes in terms of lethality (Minton, 1983; Takasaki, 1998). In contrast to the latter, however, only few studies have been focused on determining the comprehensive composition of sea snake venoms by means of proteomic analyses, i.e. venomics. The venom of A. laevis has been shown to be neurotoxic, nephrotoxic, and myotoxic in mice, causing acute renal tubular degeneration, proliferative glomerulonephritis, local muscle degeneration, necrosis, enlarged spleen, inflammation, and lymphadenopathy (Zimmerman et al., 1992a, 1992c; Ryan and Yong, 1997, 2002).

Regarding the venom components of A. laevis, a total of four short-chain neurotoxin isoforms with minor amino acid sequence variations (P19958, P19959, P19960, and P32879) and one PLA₂ (P08872) have been fully sequenced (Maeda and Nobuo, 1976; Ducancel et al., 1988, 1990). The short α-neurotoxins display a high affinity towards the acetylcholine receptor (Ishikawa et al., 1977), which is in agreement with the very low LD₅₀ observed for the whole venom (Tamiya, 1973; Maeda and Nobuo, 1976).
Toxicity of the venom has additionally been tested in different fish species, showing variations in responses (Berman, 1983; Zimmerman et al., 1990, 1992a, 1992c). It has been suggested that several components of the venom may act in a synergistic manner to potentiate toxic effects (Ryan and Yong, 1997). Finally, antivenoms raised against tiger snake (Notechis scutatus) or common sea snake (Enhydrina schistosa) venoms have been shown to have some cross-reactivity towards the venom of A. laevis, although the efficacies of these antivenoms are lower than against the venoms of homologous species (Baxter and Gallichio, 1974).

Aiming to further develop understanding of sea snake venoms and to expand knowledge of venom intra-species variability, this study presents the proteomic analysis of the venom of A. laevis, together with an assessment of variability in three different specimens, and of toxicity of all its main protein components in mice. In addition, the ability to cross-recognize and neutralize A. laevis venom was evaluated for two antivenoms against coral snakes and sea snakes.

2. Materials and Methods

2.1 Snake venom

Aipysurus laevis venom was obtained from four specimens (“Mifisto”, “Medusa”, “His”, and “Nessi”) kept at the National Aquarium, Den Blå Planet, Denmark. All specimens were collected at night by Kate L. Sanders from a boat using spotlights and dip nets. The boat was operating at shallow water close to Broome, Australia. The venom, collected in the National Aquarium of Denmark, was immediately frozen, lyophilized, and kept at -20 °C. In order to assess individual variability, a small sample of venom from each snake was kept separated, while the remaining material was pooled.
2.2 Venom separation by reverse-phase HPLC and SDS-PAGE

The pooled venom of *A. laevis* was fractionated by sequential RP-HPLC and SDS-PAGE separation steps, following the ‘snake venomics’ analytical strategy (Calvete, 2011) under conditions described previously (Lomonte et al., 2014). Venom load for the RP-HPLC step on C\textsubscript{18} (4.6 x 250 mm column, 5 μm particle diameter; Teknokroma) was 2 mg. Protein fractions were monitored at 215 nm, manually collected, dried by vacuum centrifugation, and electrophoretically separated under reducing conditions. Resulting bands were stained with colloidal Coomassie blue G-250, and digitally recorded on a ChemiDoc\textsuperscript{®} imager using ImageLab\textsuperscript{®} software (Bio-Rad).

2.3 Protein identification by tandem mass spectrometry of tryptic peptides

Protein bands were excised from gels, destained with 50% acetonitrile in 25 mM ammonium bicarbonate, and then subjected to reduction (10 mM dithiothreitol), alkylation (50 mM iodoacetamide), and overnight in-gel digestion with sequencing grade trypsin (Sigma), in 50 mM ammonium bicarbonate at 37°C. The resulting tryptic peptides were extracted with 50% acetonitrile containing 1% trifluoroacetic acid (TFA), and analyzed by MALDI-TOF-TOF on an AB4800-Plus Proteomics Analyzer (Applied Biosystems), under conditions previously described (Lomonte et al., 2014). In each run, CalMix\textsuperscript{®} standards (ABSciex) spotted onto the same plate were used as external calibrants. Resulting spectra were searched against the UniProt/SwissProt database using ProteinPilot\textsuperscript{®} v.4 and the Paragon\textsuperscript{®} algorithm (ABSciex) for protein identification at ≥ 95% score confidence, or manually interpreted. Few peptide sequences with lower confidence scores were manually searched using BLAST (http://blast.ncbi.nlm.nih.gov)
for protein similarity and assignment to protein families.

2.4 Relative protein abundance estimations

Areas of the RP-HPLC chromatographic peaks at 215 nm were integrated using ChemStation® (Agilent) in order to estimate relative protein abundances (Calvete, 2011). For peaks containing several electrophoretic bands, percentage distributions were assigned by densitometry, using ImageLab® (Bio-Rad).

2.5 Phospholipase A₂ and proteolytic enzyme activities

Enzymatic activities of A. laevis venom were tested comparatively with samples obtained from other elapid snakes (Dendroaspis polylepis, Naja kaouthia; obtained from Latoxan, France; and Micrurus nigrocinctus, obtained from Instituto Clodomiro Picado) or the viperid Bothrops asper (Instituto Clodomiro Picado). PLA₂ activity was assayed on the chromogenic 4-nitro-3-octanoyloxybenzoic acid (NOBA) synthetic substrate, as described (Lomonte et al., 2015). Venoms (20 μg, dissolved in 25 μL of 10 mM Tris, 10 mM CaCl₂, 0.1 M NaCl, pH 8.0, buffer) were mixed with 200 μL of the same buffer and 25 μL of NOBA to achieve a final substrate concentration of 0.32 mM. Plates were incubated for 60 min at 37 °C, and absorbance was recorded at 405 nm in a microplate reader. Proteolytic activity was determined on azocasein, according to Wang et al. (2004). Venoms (40 μg, dissolved in 50 mM Tris–HCl, 0.15 M NaCl, 5 mM CaCl₂ buffer, pH 8.0) were added to 100 μL of azocasein (10 mg/mL in the same buffer), and incubated for 90 min at 37 °C. The reaction was stopped by addition of 200 μL of 5% trichloroacetic acid, and after centrifugation (5 min, 13,000 rpm), 150 μL of supernatants were mixed with 100 μL of 0.5 M NaOH, and absorbance was recorded at 450 nm. All samples in these assays were run in triplicate wells, and controls of solvents
without venoms were included.

2.7 Lethality screening

Lethality assays were conducted in CD-1 mice, supplied by Instituto Clodomiro Picado, following protocols approved by the Institutional Committee for the Use and Care of Animals (CICUA), University of Costa Rica. The lethality of the whole venom and fractions or isolated toxins was tested by intravenous (i.v.) injection in groups of four mice (18–20 g body weight). Various amounts of venom or fractions/toxins were dissolved in phosphate-buffered saline (PBS; 0.12 M NaCl, 0.04 M sodium phosphate buffer, pH 7.2), and injected in the caudal vein, using a volume of 100 µL. Deaths occurring within 24 h were recorded, and the LD$_{50}$ values were calculated by probits (Finney, 1971), using the BioStat® software (AnalySoft).

The toxicity of venom fractions was initially screened by selecting a dose based on fraction abundance. The dose was selected to assess whether the fraction would score above or below 1 according to the Toxicity Score defined by Laustsen et al. (2015a) as the toxin abundance (%) divided by its LD$_{50}$. Fractions that were not lethal at this dose (yielding a Toxicity Score <1) were considered as having insignificant toxicity, whereas fractions, which did kill mice at this level, were further evaluated, and LD$_{50}$ values were determined for them.

2.8 Myotoxicity of phospholipases A$_2$

A pool of all the PLA$_2$ fractions was prepared, and doses of 30 µg, dissolved in 50 µL PBS, were injected intramuscularly, either in the right gastrocnemius, the thigh or the soleus, to groups of five mice (18-20 g). In another experiment, mice received 30 µg of the PLA$_2$ fractions in the soleus muscle. Injection of PBS was used for the control
group. Blood was collected after 3 h from the tip of the tail into heparinized capillaries. Plasma creatine kinase (CK) activity was determined using an UV kinetic assay (CK-Nac®, Analyticon). After blood collection, mice were sacrificed by CO₂ inhalation and a sample of muscles were obtained and immediately fixed in 10% formalin solution. After routine processing, tissues were embedded in paraffin, sectioned, and stained with hematoxylin-eosin for histological observation. In addition, in order to assess the acidic or basic nature of the various PLA₂s of the venom, chromatographic peaks 9-18 were analyzed by zone electrophoresis under native conditions, using a 1% agarose gel dissolved in 0.1 M Tris, 0.3 M glycine, pH 8.6 buffer. The gel was run at 75 V for 90 min, and protein migration was detected by Coomassie R-250 staining.

2.9 Antivenom neutralization studies

Two antivenoms were used: (a) BioCSL Sea Snake Antivenom, manufactured by BioCSL Limited (Melbourne, Victoria, Australia) (batch 054908201; expiry date: 04/2015); (b) Monospecific *Micrurus nigrocinctus* Anticoral Antivenom from Instituto Clodomiro Picado (batch 5310713ACLQ, expiry date 07/2016), for comparison. Mixtures containing a fixed amount of venom and several dilutions of antivenoms were prepared using PBS as diluent, and incubated at 37 °C for 30 min. Controls included venom incubated with PBS instead of antivenom. Aliquots of 100 µL of the solutions, containing 4×LD₅₀ of venom (11.2 µg/mouse) were then injected i.v. into groups of four mice (18-20 g). Deaths occurring within 24 h were recorded for assessing the neutralizing capacity of antivenoms. Neutralization was expressed as the Median Effective Dose (ED₅₀) of antivenom, defined as the ratio µg venom/mL antivenom at which 50% of the injected mice were protected. ED₅₀s were estimated by probits, as described in Section 2.7.
2.10 Antivenom immunoprofiling by ELISA

Wells in MaxiSorp plates (NUNC, Roskilde, Denmark) were coated overnight with 0.6 μg of each HPLC venom fraction, dissolved in 100 μL PBS. Then, wells were blocked by adding 100 μL PBS containing 2% (w:v) bovine serum albumin (BSA, Sigma) at room temperature for 1 h, and washed five times with PBS. A dilution of each antivenom in PBS containing 2% BSA was prepared such that the protein concentration was 86 μg/mL (as measured by their absorbance at 280 nm on a NanoDrop® 2000c instrument, Thermo Scientific), and 100 μL were added to the wells for 2 h. After five washings with PBS, 100 μL of a 1:2000 dilution of rabbit anti-horse IgG (whole molecule)-alkaline phosphatase conjugated antibodies (Sigma A6063, in PBS, 2% BSA) was added to each well for 2 h, and then washed five times with FALC buffer (0.05 M Tris, 0.15 M NaCl, 20 μM ZnCl$_2$, 1 mM MgCl$_2$, pH 7.4). Color was developed by adding 100 μL of p-nitrophenyl phosphate (1 mg/mL in 9.7% v/v diethanolamine buffer, pH 9.8), and the absorbances at 405 nm were recorded at several time intervals in a microplate reader (Multiskan FC, Thermo Scientific).

3.0 Results and Discussion

3.1 Venomics

A detailed proteomics characterization was performed on the pooled venom from *A. laevis*. From 20 fractions resolved by RP-HPLC, 35 peptidic bands were obtained after SDS-PAGE separation (Fig.1). By in-gel digestion and MALDI-TOF-TOF analysis, 99.2% of the protein bands could be assigned to toxin families. As shown in Fig.2, the predominant family of proteins in this venom corresponds to PLA$_2$s.
(71.2%), followed by a significant proportion of three-finger toxins (3FTx; 25.3%). A small amount of cysteine-rich secretory proteins (CRISP; 2.5%) and traces of a complement control module protein (CCM; 0.2%) were also detected. These results highlight the simple protein family composition of A. laevis venom, which essentially relies on a relatively small group of PLA2 and 3FTx isoforms to exert its trophic role. Also, these findings are in agreement with the trend emerging from recent proteomic studies on sea snake venoms, which have revealed that their venoms are much simpler than their terrestrial elapid counterparts in terms of the number of dominant protein families and diversification of isoforms, typically within the PLA2 and 3FTx families (Fry et al., 2003; Li et al., 2005). Thus far, sea snake venom proteomes have been deciphered for Hydrophis cyanocinctus (Calvete et al., 2012) and Pelamis platura (Lomonte et al., 2014). Similar to these, A. laevis venom contains few toxin families. However, in contrast to the venoms of P. platura and H. cyanocinctus, where the main toxin families are three-finger toxins (50% and 81% of all venom proteins, respectively) followed by PLA2s (33% and 19% of all venom proteins, respectively), A. laevis venom displays the opposite relative venom composition, being dominated by PLA2s (71.2%) followed by 3FTxs (25.3%). Furthermore, whereas the three-finger toxins of A. laevis venom are all short neurotoxins, P. platura and H. cyanocinctus venoms contain both short and long neurotoxins. The current findings on A. laevis venom composition differ from a previous study, where three of the short neurotoxins were reported to represent 22%, 33%, and 21% of the venom (76% altogether), respectively (Maeda and Tamiya, 1976). The reasons behind these discrepant results are difficult to determine, although they may reflect possible intraspecies differences in venom composition in specimens collected in different geographical locations: Maeda and Tamiya (1976) used A. laevis collected from Ashmore Reef, which is separated from our collection localities near
Broome by more than 600 km of mostly unsuitable (deep water) habitat. Potentially, the observed differences could also be explained by interspecific hybridization, which has previously been observed for *A. laevis* and closely related *A. fuscus* on Ashmore Reef, where hybrid individuals closely resemble *A. laevis* in phenotype (Sanders et al., 2014). In similarity with the predominance of PLA$_2$s over 3FTxs herein reported for *A. laevis* venom, a transcriptomic study on the venom glands of *Aipysurus eydouxi* revealed the existence of as many as sixteen unique PLA$_2$ transcripts, in contrast to a single transcript corresponding to a 3FTx (Li et al., 2005). This could suggest that both *Aipysurus* species share the same venom compositional predominance. However, assessment of this possibility would require a direct examination of the *A. eydouxi* venom proteins, in addition to its venom gland transcripts.

Individual variations of toxin expression in snake venoms are not uncommon (Chippaux et al., 1991). To investigate the possible individual variability in *A. laevis*, samples from three specimens (“Mifisto”, “Medusa”, and “Nessi”) were compared by RP-HPLC (Fig.3). This analysis revealed that some qualitative variation in toxin expression was indeed present, although most fractions did not show significant deviation in abundance between specimens or pooled venom.

Unlike several terrestrial elapids (Aird, 2002; Laustsen et al., 2015a), nucleosides were not detected in *A. laevis* venom. On the other hand, its high content of PLA$_2$s suggests that this venom might induce myotoxicity, as previously shown in experimental studies (Zimmerman et al., 1992c; Ryan and Yong, 1997, 2002). Systemic myotoxicity, i.e. rhabdomyolysis, with myoglobinuria characterizes envenomings by some species of sea snakes in humans (Reid, 1961), and is responsible for acute kidney injury. However, when a pool of PLA$_2$ fractions of *A. laevis* venom was tested for myotoxicity in mice, only a mild effect was observed, as judged by increments in
plasma CK activity. Mice receiving PBS had CK activity of 215 ± 10 U/L, and mice injected in the gastrocnemius, thigh, or soleus muscles with 30 µg of the PLA₂ fraction pool had plasma CK activities of 926 ± 160 U/L, 1196 ± 119 U/L, and 764 ± 182 U/L (mean ± SEM), respectively. Increments in CK were significant only when the PLA₂ fraction pool was injected in the gastrocnemius and thigh muscles (p<0.05). Thus, _A. laevis_ PLA₂s only induced a mild myotoxic effect. In agreement, histological analysis of the soleus muscle 3 h after injection of PLA₂ fraction pool showed only few scattered necrotic fibers (Fig.4). These observations contrast with the prominent increment in plasma CK activity described for other elapid venoms, such as that of _Micrurus nigrocinctus_ (Fernández et al., 2011). By using native zone electrophoresis, it was observed that all PLA₂ fractions (peaks 9-18) migrated towards the anode, indicating that they were of acidic nature (not shown). This observation could explain the low myotoxic effect of the PLA₂ pool tested, since commonly PLA₂s having potent myotoxic effects are of a basic nature (Montecucco et al., 2008). Our results suggest that myotoxicity is unlikely to be a significant effect in envenomings by _A. laevis_.

In agreement with its proteomic composition showing an abundance of PLA₂s, high PLA₂ activity of the venom was confirmed _in vitro_ (Fig.5A), whereas no proteinase activity was detected (Fig.5B), in line with the absence of these enzymes in the venom proteome. Three-finger toxins were shown to represent the second major group of venom proteins in terms of abundance (25.3%), and all of them were identified as short neurotoxin isoforms (Table 1), previously characterized by Maeda and Tamiya (1976) and Ducancel et al. (1990). These short neurotoxins have been shown to bind with high affinity to nicotinic receptors at the motor end-plate of muscle fibers, leading to flaccid paralysis, which may result in respiratory failure and death (Maeda and Nobuo, 1976; Ducancel et al., 1990).
All venom fractions were examined for acute toxicity in CD1 mice, and LD$_{50}$ values were determined for most of those having a Toxicity Score below 1 (Table 2). All fractions containing short neurotoxins (fractions 1-4) and some fractions containing PLA$_2$s (fractions 5-18) induced lethality in mice, although the LD$_{50}$ values of the short neurotoxins were 10-40 fold lower than those of the PLA$_2$s. Evaluated on the basis of their Toxicity Score, the short neurotoxins of *A. laevis* venom are the most relevant toxins to target in order to counteract the main clinical manifestations of the venom. The venom of *A. laevis* is remarkably simple compared to terrestrial elapids, such as *Dendroaspis polylepis* (Laustsen et al., 2015a) and *Naja kaouthia* (Laustsen et al., 2015b), which display a more diverse arsenal of toxins, although also being dominated by only two main toxin families.

The concept of a Toxicity Score for acute toxicity was presented for the first time in Laustsen et al. (2015a), and this score can be used to rank the importance of the individual toxins for acute toxicity in the given *in vivo* model (typically rodents). By examining the difference between the Toxicity Score of whole venom and the Accumulated Toxicity Score for all venom components (the sum of the Toxicity Scores for all the for the individual venom components), an indication of how the toxins in whole venom interact can be deduced. For a venom displaying synergism, the Toxicity Score for whole venom will be higher than the sum of the Toxicity Score for the individual components, since the synergistic effects between toxins will lead to an increased potency of the venom. For *A. laevis* there seems to be an indication that the Toxicity Score of whole venom (TS = 676) is almost the double of the Accumulated Toxicity Score of the venom components (ATS = 357) (Table 2), indicating that synergistic effects may exist. This observation is supported by previous studies indicating the presence of synergism (Ryan and Yong, 1997), which is quite fascinating.
given the simplicity of this venom, being dominated by only a few very similar isoforms of short neurotoxins responsible for the main neurotoxic effects.

3.2 Venom neutralization and antivenom profiling

The ability of BioCSL Sea Snake Antivenom and ICP Anti-Coral Antivenom to neutralize *A. laevis* venom was investigated in CD-1 mice. The BioCSL Sea Snake Antivenom was effective in neutralizing lethality with an ED$_{50}$ of 821 µg venom per mL antivenom (95% confidence limits: 478–1439 µg/mL), whereas no neutralization was observed for ICP Anti-Coral Antivenom at a level of 100 µg venom per mL. Our observations are in agreement with previous findings on the ability of BioCSL Sea Snake Antivenom to neutralize the neuromuscular blocking activity of *A. laevis* and other sea snake venoms (Chetty et al., 2004). To further investigate the antivenoms, both BioCSL Sea Snake Antivenom and ICP Anti-Coral Antivenom were profiled by ELISA to determine the extent of binding between antivenom antibodies and toxins in *A. laevis* venom (Fig.6). Two general trends present themselves: BioCSL Sea Snake Antivenom displays significantly higher binding to fractions containing short neurotoxins (fractions 1-4), whereas the ICP Anti-Coral Antivenom displays either similar or even increased binding against PLA$_2$ containing fractions (fractions 5-18). This finding further supports that the short neurotoxins are responsible for the main toxic effects of *A. laevis* venom.

The underlying reason for the differences in binding preference between the two antivenoms may be explained by the venom compositions of *Micrurus nigrocinctus* and *Enhydrina schistosa*, which are used in the immunization mixtures of BioCSL Sea Snake Antivenom and ICP Anti-Coral Antivenom, respectively (Fig.7A). It must be noted, however, that it is not unlikely that horses hyper-immunized with several
different snake venoms were used for production of BioCSL Sea Snake Antivenom, and that the “monovalence” of this antivenom is primarily due to the horses being boosted with *E. schistosa* venom immediately before bleeding (Chetty et al., 2004; O’Leary and Isbister, 2009; Herrera et al., 2014). Therefore, unexpected cross-reactivity is not an unlikely event. *E. schistosa* venom has a high abundance of 3FTxs with a high degree of conservation relative to the short neurotoxins found in *A. laevis* (Fig. 7B), and it is therefore not surprising that the BioCSL Sea Snake Antivenom has a strong preference for fractions 1-4, containing short neurotoxins from *A. laevis*. In comparison, the PLA$_2$s found in *M. nigrocinctus* venom are not more similar to the PLA$_2$s found in *A. laevis* venom than the PLA$_2$ reported for *E. schistosa* venom (Fig. 7C) (Fohlman and Eaker, 1977). However, it is speculated that the much higher abundance of PLA$_2$s in the immunization mixture used for producing ICP Anti-Coral Antivenom in itself drives the immunological response towards a stronger recognition against PLA$_2$s in general. Given that BioCSL Sea Snake Antivenom readily cross-recognizes the neurotoxic components having the highest Toxicity Scores, and since this antivenom was shown to neutralize whole venom in rodents, BioCSL Sea Snake Antivenom should be useful for treating human snakebite accidents inflicted by *A. laevis*.

The venom of *A. laevis* is remarkably simple. It could therefore be feasible to develop modern antivenoms based on human(ized) monoclonal antibodies or peptide-based inhibitors against this venom, since it is likely that only few antibodies are needed to obtain its full neutralization. The degree of conservation, especially in the clinically relevant short neurotoxins is high, and it is therefore likely that a potent, cross-reactive antibody or peptide-based inhibitor capable of neutralizing all of these components can be developed.
4.0 Concluding remarks and outlook

A proteomic analysis and functional study of *A. laevis* venom was carried out, revealing that this venom is remarkably simple and dominated by PLA2s (71.2% of venom protein content) followed by short neurotoxins of the three-finger toxin family (25.3% of venom protein content). Also, cysteine-rich secretory proteins (CRISP) and a complement control module (CCM) were detected. Based on thorough toxicity testing of the individual fractions obtained from whole venom, the most relevant toxins to target for an effective antivenom against acute toxicity are the short neurotoxins. Based on their Toxicity Scores, the toxins present in *A. laevis* venom seem to interact in a slightly synergistic manner, possibly due to the short neurotoxins all targeting the nicotinic receptors at the motor end-plate of muscle fibers. BioCSL Sea Snake Antivenom was capable of neutralizing *A. laevis* venom in CD-1 mice when venom and antivenom were preincubated and administered by i.v. injection. ELISA-based immunoprofiling indicated that the BioCSL Sea Snake Antivenom has a binding preference for short neurotoxins. Therefore, this antivenom should be of clinical use for treating bites inflicted by *A. laevis*. Finally, given the simplicity of *A. laevis* venom, a potential for developing a modern antivenom based on human(ized) monoclonal antibodies or peptide-based inhibitors may be a possibility in the future.

Acknowledgments

The authors thank Dr. Ken Winkel and Dr. David Williams (University of Melbourne, Australia) for kindly providing the sample of BioCSL Sea Snake Antivenom. The authors further thank the Department of Drug Design and Pharmacology (University of Copenhagen), the Instituto Clodomiro Picado (Universidad de Costa Rica), and Den Blå Planet for supporting the research. Finally,
we thank the following foundations for financial support: Drug Research Academy (University of Copenhagen), Novo Nordisk Fond, Dansk Tennis Fond Oticon Fonden, Knud Højgaards Fond, Rudolph Als Fondet, Henry Shaws Legat, Læge Johannes Nicolai Krigsgaard of Hustru Else Krogsgaards Mindelegat for Medicinsk Forskning og Medicinske Studenter ved Københavns Universitet, Lundbeckfonden, Torben of Alice Frimodts Fond, Frants Allings Legat, Christian og Ottilia Brorsons Rejselegat for Yngre Videnskabsmænd- og kvinder, and Fonden for Lægevidenskabens Fremme.

Ethical statement

The authors declare that there are no conflicts of interest related to this study.

J.M. Gutiérrez and B. Lomonte work at the Instituto Clodomiro Picado (Universidad de Costa Rica), where the anti-coral snake antivenom used in this study is produced. Sources that provided financial support were not involved in the collection, analysis, or interpretation of data, nor in writing the report and submitting it for publication.
References


Berman, D.M., 1983. The toxicities of snake venoms to goldfish (Carassiusauratus) and the susceptibilities of reef fish and crabs to olive sea snake (Aipysurus laevis) venom. Toxicon 21, 37–40.


26


Figure legends

Figure 1: Separation of *A. laevis* (A) venom proteins by RP-HPLC (C), followed by SDS-PAGE (B). Two mg of venom were fractionated on a C$_{18}$ column and eluted with an acetonitrile gradient (dashed line), as described in Methods. Fractions were further separated by SDS-PAGE under reducing conditions. Molecular weight markers (Mw) are indicated in kDa. Coomassie-stained bands were excised, in-gel digested with trypsin, and subjected to MALDI-TOF/TOF analysis for assignment to protein families, as shown in Table 1.

Figure 2: Composition of *A. laevis* venom according to protein families, expressed as percentages of the total protein content. 3FTx: three-finger toxin; PLA$_2$: phospholipase A$_2$; CRISP: cysteine-rich secretory protein; CCM: complement control module. (see Table 1).

Figure 3: Comparison of the chromatographic profiles of the venoms from three individual *A. laevis* specimens (“Mifisto”, “Medusa”, and “Nessi”). The patterns of the individual venoms are similar, although some differences do occur in abundance for certain fractions. Fraction peaks representing more than 1% of total venom protein, which have an abundance deviating more than 50% from the pool, are marked with *.

Figure 4: Light micrographs of sections of mouse soleus muscles collected 3 h after injection of either phosphate-buffered saline (PBS) solution (A) or 30 µg of a pool of all the PLA$_2$ fractions of the venom of *A. laevis* (B) (see Methods for experimental details). A normal histological pattern is observed in (A), whereas few scattered necrotic muscle...
fibers (arrows) are observed in (B), thus evidencing the mild myotoxic activity of this venom. Hematoxylin-eosin staining. Bar represents 100 µm.

**Figure 5:** (A) Comparison of the phospholipase A$_2$ activity between the venoms of *Aipysurus laevis*, *Dendroaspis polylepis*, *Naja kaouthia*, and *Micrurus nigrocinctus*. *A. laevis* displays high enzymatic activity, although lower than *M. nigrocinctus*. (B) Comparison of the proteolytic activity between the venoms of *A. laevis*, *D. polylepis*, *N. kaouthia*, *M. nigrocinctus*, and *Bophrops asper*, evaluated on azocasein. *A. laevis* shows negligible proteinase activity.

**Figure 6:** ELISA-based immunoprofiling of antivenoms (**CSL Sea Snake**: BioCSL Sea Snake Antivenom from BioCSL Limited, **ICP Micrurus**: Monospecific *Micrurus nigrocinctus* Anticoral Antivenom from Instituto Clodomiro Picado, for comparison, and a negative control (**Horse negative**: normal serum from non-immunized horses from Instituto Clodomiro Picado) to all fractions of *A. laevis* venom separated by RP-HPLC (see Methods for details). For identification of venom fractions refer to Table 2. Each bar represents mean ± SD of triplicate wells.

**Figure 7:** (A) Comparison between the venom profiles and similarities of toxins from *Aipysurus laevis*, *Enhydrina schistosa* (*Hydrophis schistosa*), and *Micrurus nigrocinctus*. *E. schistosa* venom is used for production of BioCSL Sea Snake Antivenom, while *M. nigrocinctus* venom is used for production of ICP’s Monospecific *M. nigrocinctus* Anticoral Antivenom. *Venom composition is based on venomics studies of *M. nigrocinctus* (Fernández et al., 2011) and the reported study of *Hydrophis cyanocinctus* (Calvete et al., 2012), as such studies have not been performed on *E.*
schistosa. Notice that the PLA₂ content of the venoms differ. The green frames highlight observations used in discussion of Fig.6 (see text). (B) Alignment of all known three-finger toxins (3FTx) from A. laevis with the most similar toxins from M. nigrocinctus and E. schistosa. A high degree of conservation exists between short neurotoxins from A. laevis and the most similar E. schistosa toxin, possibly explaining the observed cross-reactivity of BioCSL Sea Snake Antivenom. (C) Alignment of the single known PLA₂ from A. laevis with the most similar toxins from M. nigrocinctus and E. schistosa, showing only a limited degree of conservation.
Table 1: Assignment of the RP-HPLC isolated fractions of *A. laevis* venom to protein families by MALDI-TOF-TOF of selected reproducible ions from in-gel trypsin-digested protein bands.

<table>
<thead>
<tr>
<th>Peak</th>
<th>% Mass</th>
<th>Peptide Ion</th>
<th>Cont</th>
<th>Sc</th>
<th>Related Protein, code</th>
<th>Protein family</th>
<th>MS/MS-derived sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>07</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>08</td>
<td>0.4</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>09</td>
<td>0.1</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>10</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>11</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>12</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>13</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>14</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>15</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>16</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>17</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>18</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
<tr>
<td>19</td>
<td>0.3</td>
<td>P19960</td>
<td>1</td>
<td>1</td>
<td>11010000</td>
<td>short neurotoxin D</td>
<td>TTTDCADDSCYBK XTCCNBBSSBPK GCGCPBVBPGXK</td>
</tr>
</tbody>
</table>

*Note: The data represents the assignment of the RP-HPLC isolated fractions of *A. laevis* venom to protein families by MALDI-TOF-TOF of selected reproducible ions from in-gel trypsin-digested protein bands.*
<table>
<thead>
<tr>
<th>Protein Name</th>
<th>Accession</th>
<th>Mass (Da)</th>
<th>pI</th>
<th>Percent Identity</th>
<th>Percent Similarity</th>
<th>SCWRL4 Score</th>
<th>Tm (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>phospholipase A</td>
<td>Q5DNE1</td>
<td>2091.0</td>
<td>12</td>
<td>99</td>
<td>91</td>
<td>0.2</td>
<td>11</td>
</tr>
<tr>
<td>phospholipase A</td>
<td>Q8QFW3</td>
<td>2337.1</td>
<td>11</td>
<td>99</td>
<td>98</td>
<td>9.1</td>
<td>10</td>
</tr>
<tr>
<td>phospholipase A</td>
<td>P08872</td>
<td>2912.3</td>
<td>12</td>
<td>99</td>
<td>95.3</td>
<td>3.3</td>
<td>10</td>
</tr>
<tr>
<td>phospholipase A</td>
<td>P08872</td>
<td>2895.4</td>
<td>11</td>
<td>99</td>
<td>97.3</td>
<td>7.2</td>
<td>10</td>
</tr>
<tr>
<td>phospholipase A</td>
<td>P08872</td>
<td>2835.4</td>
<td>11</td>
<td>99</td>
<td>96.1</td>
<td>6.2</td>
<td>10</td>
</tr>
<tr>
<td>phospholipase A</td>
<td>P08872</td>
<td>2091.0</td>
<td>10</td>
<td>99</td>
<td>97.3</td>
<td>1.7</td>
<td>9</td>
</tr>
</tbody>
</table>

Note: SCWRL4 score ranges from 0.0 to 1.0, with higher scores indicating better structure prediction. Tm represents the thermal stability of the protein in °C.
<table>
<thead>
<tr>
<th></th>
<th>Unknown</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
<th>13</th>
<th>14</th>
<th>15</th>
<th>16</th>
<th>17</th>
<th>18</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Proteins</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hydrophis hodgkini</td>
<td>-66</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
<td>16</td>
<td>17</td>
<td>18</td>
</tr>
<tr>
<td>CRISP</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hydrophis hodgkini</td>
<td>-66</td>
<td>159</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
<td>16</td>
<td>17</td>
</tr>
<tr>
<td>CRISP</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Hydrophis hodgkini</em></td>
<td>-66</td>
<td>190</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
<td>16</td>
<td>17</td>
</tr>
<tr>
<td><strong>Homology</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Hydrophis hodgkini</em></td>
<td>-66</td>
<td>191</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
<td>16</td>
<td>17</td>
</tr>
<tr>
<td><strong>Homology</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Hydrophis hodgkini</em></td>
<td>-66</td>
<td>192</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
<td>16</td>
<td>17</td>
</tr>
<tr>
<td><strong>Homology</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Hydrophis hodgkini</em></td>
<td>-66</td>
<td>193</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
<td>13</td>
<td>14</td>
<td>15</td>
<td>16</td>
<td>17</td>
</tr>
</tbody>
</table>
Cysteine residues are carbamidomethylated. Confidence (Conf) and Score (Sc) values are calculated by the Paragon® algorithm of ProteinPilot®. **▼:** reduced SDS-PAGE mass estimations, in kDa. X: Leu/Ile; B: Lys/Gln; Z: pyroglutamate (2-oxo-pyrrolidone carboxylic acid). Possible, although unconfirmed/ambiguous amino acid modifications suggested by the automated identification software are shown in parentheses, with the following abbreviations: ox: oxidized; da: deamidated; dt: dethiomethyl; pa: propionamide; fo: formylated.

Protein family abbreviations: 3FTx: three-finger toxin; PL-A2: phospholipase A2; CRISP: cysteine-rich secretory protein; CCM.
<table>
<thead>
<tr>
<th>Peak</th>
<th>Protein Family</th>
<th>Toxicity Score</th>
<th>% LD&lt;sub&gt;50&lt;/sub&gt; (95% C.L.)</th>
<th>Toxicity Score</th>
<th>% LD&lt;sub&gt;50&lt;/sub&gt; (95% C.L.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1&gt;</td>
<td></td>
<td>3.2</td>
<td>(0.01 - 0.15)</td>
<td>2&gt;</td>
<td>(0.04 - 0.08)</td>
</tr>
<tr>
<td>1&gt;</td>
<td></td>
<td>1.3</td>
<td>(0.09 - 0.15)</td>
<td>1.3</td>
<td>(0.09 - 0.15)</td>
</tr>
<tr>
<td>1&gt;</td>
<td></td>
<td>3.3</td>
<td>(0.08 - 0.15)</td>
<td>3.3</td>
<td>(0.08 - 0.15)</td>
</tr>
<tr>
<td>1&gt;</td>
<td></td>
<td>3.4</td>
<td>(0.10 - 0.15)</td>
<td>3.4</td>
<td>(0.10 - 0.15)</td>
</tr>
<tr>
<td>1&gt;</td>
<td></td>
<td>1.0</td>
<td>(0.08 - 0.15)</td>
<td>1.0</td>
<td>(0.08 - 0.15)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>0.2</td>
<td>(0.03 - 0.09)</td>
<td>0.2</td>
<td>(0.03 - 0.09)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>0.5</td>
<td>(0.05 - 0.18)</td>
<td>0.5</td>
<td>(0.05 - 0.18)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>0.9</td>
<td>(0.10 - 0.28)</td>
<td>0.9</td>
<td>(0.10 - 0.28)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>0.12</td>
<td>(0.07 - 0.19)</td>
<td>0.12</td>
<td>(0.07 - 0.19)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>0.3</td>
<td>(0.03 - 0.15)</td>
<td>0.3</td>
<td>(0.03 - 0.15)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>0.3</td>
<td>(0.03 - 0.15)</td>
<td>0.3</td>
<td>(0.03 - 0.15)</td>
</tr>
<tr>
<td>1&gt;</td>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;: Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
<td>1.0</td>
<td>(0.08 - 0.20)</td>
<td>1.0</td>
<td>(0.08 - 0.20)</td>
</tr>
</tbody>
</table>

**Table 2**: LD<sub>50</sub> values of *Aipysurus laevis* venom and the RP-HPLC isolated fractions.
Toxicity Score was defined as the ratio of protein fraction abundance (%) in the venom divided by its estimated median lethal dose (LD$_{50}$) for CD-1 mice by i.v. injection.  

Mix indices imply that the fraction did not contain a pure, isolated toxin, but instead a mixture of ≥ 4 different toxins in variable ratios indicated in the table.

N.t.: not tested

<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>&lt;0.5</td>
<td>CRISP</td>
<td></td>
<td>0.1</td>
<td>2.0</td>
<td>0.3</td>
<td>3.05 (1.92-4.67)</td>
<td>3.6</td>
</tr>
<tr>
<td>1</td>
<td>&gt;0.5</td>
<td>CRISP</td>
<td></td>
<td>0.3</td>
<td>2.5</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
</tr>
<tr>
<td>1</td>
<td>&gt;0.3</td>
<td>CRISP</td>
<td></td>
<td>0.3</td>
<td>2.5</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
</tr>
<tr>
<td>1</td>
<td>&gt;0.2</td>
<td>CRISP</td>
<td></td>
<td>0.3</td>
<td>2.5</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
</tr>
<tr>
<td>1</td>
<td>&gt;0.1</td>
<td>CRISP</td>
<td></td>
<td>0.3</td>
<td>2.5</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
</tr>
</tbody>
</table>
Figure 1

Absorbance 215 nm (mU)

Time (min)

Acetone (20-80%)

A

B

C
Figure 2
Figure 3

A. laevis 1
“Mifisto”

A. laevis 2
“Medusa”

A. laevis 3
“Nessi”
Figure 5

A

\[ A \text{ (405 nm)} \]

\[ \text{Venom (20 \( \mu \)g/well)} \]

- \( D. \text{ polylepis} \)
- \( N. \text{ kaouthia} \)
- \( A. \text{ laevis} \)
- \( M. \text{ nigrocinctus} \)

B

\[ A \text{ (450 nm)} \]

\[ \text{Venom (40 \( \mu \)g/well)} \]

- \( D. \text{ polylepis} \)
- \( N. \text{ kaouthia} \)
- \( A. \text{ laevis} \)
- \( M. \text{ nigrocinctus} \)
- \( B. \text{ asper} \)
### Figure 7

#### A

<table>
<thead>
<tr>
<th>Antivenom</th>
<th>BioCSL sea snake</th>
<th>ICP Anticoral</th>
</tr>
</thead>
<tbody>
<tr>
<td>Venom used for Immunization</td>
<td><em>Enhydrina schistosa</em></td>
<td><em>Micrurus nigrocinctus</em></td>
</tr>
<tr>
<td>Venom composition*</td>
<td>3FTx</td>
<td>PLA₂</td>
</tr>
<tr>
<td>Similarity of 3FTx to <em>A. laevis</em> 3FTx</td>
<td>Highly conserved, identity 90%</td>
<td>Well conserved, identity 63%</td>
</tr>
<tr>
<td>Similarity of PLA₂ to <em>A. laevis</em> PLA₂</td>
<td>Partially conserved, identity 58%</td>
<td>Partially conserved, identity 51%</td>
</tr>
</tbody>
</table>

#### B

**Alignment 3FTxs (all short-chain, type 1 α-neurotoxins)**

- **E. schistosa**
- **A. laevis**
- **M. nigrocinctus**

<table>
<thead>
<tr>
<th>Accession</th>
<th>Identity</th>
</tr>
</thead>
<tbody>
<tr>
<td>P06415</td>
<td></td>
</tr>
<tr>
<td>P12979</td>
<td></td>
</tr>
<tr>
<td>P19950</td>
<td></td>
</tr>
<tr>
<td>P19952</td>
<td></td>
</tr>
<tr>
<td>P19956</td>
<td></td>
</tr>
<tr>
<td>P09950</td>
<td></td>
</tr>
<tr>
<td>P09940</td>
<td></td>
</tr>
</tbody>
</table>

#### C

**Alignment PLA₂s (all group 1, D49 sub-subfamily members)**

- **E. schistosa**
- **A. laevis**
- **M. nigrocinctus**

<table>
<thead>
<tr>
<th>Accession</th>
<th>Identity</th>
</tr>
</thead>
<tbody>
<tr>
<td>P06113</td>
<td></td>
</tr>
<tr>
<td>P09973</td>
<td></td>
</tr>
<tr>
<td>P09944</td>
<td></td>
</tr>
</tbody>
</table>
Graphical Abstract

Danger in the reef

*Aipysurus laevis*