Peptide Nucleic Acids Having Enhanced Binding Affinity and Sequence Specificity

Buchardt, Ole; Engholm, Michael; Nielsen, Peter E.; Berg, Rolf Henrik

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PEPTIDE NUCLEIC ACIDS HAVING ENHANCED BINDING AFFINITY AND SEQUENCE SPECIFICITY

Inventors: Ole Buchardt, deceased, late of 3500 Vaelose, Denmark; Michael Egholm, 1231 Lexington Ridge Dr., Lexington, Mass. 02173; Peter Eigel Nielsen, Hjotrevanget 509, 2980 Kokkedal; Rolf Henriek Berg, Strandvaengen 6 DK 2960 Rungsted Kyst, both of Denmark; by Dorte Buchardt, executor, Sodergardesvej 73, 3500 Vaerlose, Germany

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FOREIGN PATENT DOCUMENTS
WO 86/05518 9/1986 WIPO
WO 90/02749 3/1990 WIPO
WO 92/20702 11/1992 WIPO

OTHER PUBLICATIONS

ABSTRACT
A novel class of compounds, known as peptide nucleic acids, bind complementary DNA and RNA strands more strongly than a corresponding DNA strand, and exhibit increased sequence specificity and binding affinity. Methods of increasing binding affinity and sequence specificity of peptide nucleic acids are provided wherein some peptide nucleic acids comprise ligands selected from a group consisting of naturally-occurring nucleobases and non-naturally-occurring nucleobases attached to a polyamide backbone, while other peptide nucleic acids contain at least one 2,6-diaminopurine nucleobase and at least one C1-C8 alkylamine side chain.

24 Claims, 11 Drawing Sheets
OTHER PUBLICATIONS


Nielsen, P.E., "Sequence-Specific Transcription Arrest by peptide Nucleic Acid Bound to the NDA Template Strand", Gene, 1994, 149, 139–145.


Mizutani et al., "Oligo(DT)–glycercyl porous glass, a better support for the preparation of mRNA". *J. Chromatogr.*, 1986, 356, 202-205.


5,766,855

Page 5


Fig. 1(b)
Fig. 2
**Fig. 3**
Fig. 6
Fig. 7(a)
Fig. 7(b)
Fig. 7(c)
HCV RNA/PNA IVT

Fig. 8
PEPTIDE NUCLEIC ACIDS HAVING ENHANCED BINDING AFFINITY AND SEQUENCE SPECIFICITY

CROSS-REFERENCE TO RELATED APPLICATIONS


FIELD OF THE INVENTION

The present invention is directed to methods for enhancing sequence specificity, binding affinity of peptide nucleic acids (PNAs) in which naturally-occurring nucleobases or non-naturally-occurring nucleobases are covalently bound to a polyamide backbone. One preferred naturally-occurring nucleobase is 2,6-diaminopurine. Some PNAs of the present invention comprise at least one 2,6-diaminopurine nucleobase while other PNAs comprise at least one 2,6-diaminopurine nucleobase and at least one C1-C6 alkylamine sidechain resulting in enhanced binding affinity to nucleic acids and sequence specificity as well as other beneficial qualities.

BACKGROUND OF THE INVENTION

The function of a gene starts by transcription of its information to a messenger RNA (mRNA). By interacting with the ribosomal complex, mRNA directs synthesis of the protein. This protein synthesis process is known as translation. Translation requires the presence of various cofactors, building blocks, amino acids and transfer RNAs (tRNAs), all of which are present in normal cells.

Most conventional drugs exert their effect by interacting with and modulating one or more targeted endogenous proteins, e.g., enzymes. Typically, however, such drugs are not specific for targeted proteins but interact with other proteins as well. Thus, use of a relatively large dose of drug is necessary to effectively modulate the action of a particular protein. If the modulation of a protein activity could be achieved by interaction with or inactivation of mRNA, a dramatic reduction in the amount of drug necessary, and the side-effects of the drug, could be achieved. Further reductions in the amount of drug necessary and the side-effects could be obtained if such interaction is site-specific. Since a functioning gene continually produces mRNA, it would be even more advantageous if genetic transcription could be arrested in its entirety. Oligonucleotides and their analogs have been developed and used as diagnostics, therapeutics and research reagents. One example of a modification to oligonucleotides is labeling with non-isotopic labels, e.g., fluorescein, biotin, digoxigenin, alkaline phosphatase, or other reporter molecules. Other modifications have been made to the ribose phosphate backbone to increase the resistance to nucleases. These modifications include use of linkages such as methyl phosphonates, phosphorothioates and phosphorodithioates, and 2'-O-methyl ribose sugar moieties. Other oligonucleotide modifications include those made to modulate uptake and cellular distribution. Phosphorothioate oligonucleotides are presently being used as antisense agents in human clinical trials for the treatment of various disease states. Although some improvements in diagnostic and therapeutic uses have been realized with these oligonucleotide modifications, there exists an ongoing demand for improved oligonucleotide analogs.

In the art, there are several known nucleic acid analogs having nucleobases bound to backbones other than the naturally-occurring ribonucleic acids or deoxyribonucleic acids. These nucleic acid analogs have the ability to bind to nucleic acids with complementary nucleobase sequences. Among these, the peptide nucleic acids (PNAs), as described, for example, in WO 92/20702, have been shown to be useful as therapeutic and diagnostic reagents. This may be due to their generally higher affinity for complementary nucleobase sequence than the corresponding wild-type nucleic acids.

PNAs are compounds that are analogous to oligonucleotides, but differ in composition. In PNAs, the deoxyribose backbone of oligonucleotide is replaced by a peptide backbone. Each subunit of the peptide backbone is attached to a naturally-occurring or non-naturally-occurring nucleobase. One such peptide backbone is constructed of repeating units of N-(2-aminoethyl)glycine linked through amide bonds.

PNAs bind to both DNA and RNA and form PNA/DNA or PNA/RNA duplexes. The resulting PNA/DNA or PNA/RNA duplexes are bound tighter than corresponding DNA/DNA or DNA/RNA duplexes as evidenced by their higher melting temperatures (Tm). This high thermal stability of PNA/DNA(RNA) duplexes has been attributed to the neutrality of the PNA backbone, which results elimination of charge repulsion that is present in DNA/DNA or RNA/RNA duplexes. Another advantage of PNA/DNA(RNA) duplexes is that Tm is practically independent of salt concentration. DNA/DNA duplexes, on the other hand, are highly dependent on the ionic strength.

Homopyrimidine PNAs have been shown to bind complementarily DNA or RNA forming PNA/DNA(RNA) triplexes of high thermal stability (Egholm et al., Science, 1991, 254, 1497; Egholm et al., J. Am. Chem. Soc., 1992, 114, 1895; Egholm et al., J. Am. Chem. Soc., 1992, 114, 9677).

In addition to increased affinity, PNAs have increased specificity for DNA binding. Thus, a PNA/DNA duplex mismatch show 8° to 20° C. drop in the Tm relative to the DNA/DNA duplex. This decrease in Tm is not observed with the corresponding DNA/DNA duplex mismatch (Egholm et al., Nature 1993, 365, 566).

A further advantage of PNAs, compared to oligonucleotides, is that the polyamide backbone of PNAs is resistant to degradation by enzymes.

Considerable research is being directed to the application of oligonucleotides and oligonucleotide analogs that bind to complementary DNA and RNA strands for use as diagnostics, research reagents and potential therapeutics. For many applications, the oligonucleotides and oligonucleotide analogs must be transported across cell membranes or taken up by cells to express their activity.

PCT/EP/01219 describes novel PNAs which bind to complementary DNA and RNA more tightly than the corresponding DNA. It is desirable to append groups to these PNAs which will modulate their activity, modify their membrane permeability or increase their cellular uptake property. One method for increasing amount of cellular uptake property of PNAs is to attach a lipophilic group. U.S. application Ser. No. 117,363, filed Sep. 3, 1993, describes several alkylamino functionalities and their use in the attachment of such pendant groups to oligonucleotides.
U.S. application Ser. No. 07/943,516, filed Sep. 11, 1992, and its corresponding published PCT application WO 94/06815, describe other novel amine-containing compounds and their incorporation into oligonucleotides for, inter alia, the purposes of enhancing cellular uptake, increasing lipophilicity, causing greater cellular retention and increasing the distribution of the compound within the cell.

U.S. application Ser. No. 08/116,801, filed Sep. 3, 1993, describes nucleosides and oligonucleosides derivatized to include a thiolalkyl functionality, through which pendant groups are attached.

Despite recent advances, there remains a need for a stable compound that enhances or modulates binding to nucleic acids, stabilizes the hybridized complexes and increases the aqueous solubility.

SUMMARY OF THE INVENTION

The present invention provides peptide nucleic acids (PNAs) with higher binding affinity to complementary DNA and RNA than corresponding DNA. The PNAs of the present invention comprise ligands linked to a polyamide backbone. Representative ligands include the four major naturally-occurring DNA nucleobases (i.e. thymine, cytosine, adenine and guanine), other naturally-occurring nucleobases (e.g. inosine, uracil, 5-methylcytosine, thioracil or 2,6-diaminopurine) or artificial bases (e.g. bromothymine,azaadenines or azaguainines) attached to a polyamide backbone through a suitable linker.

The present invention provides a method for enhancing the DNA or RNA sequence specificity of a peptide nucleic acid by incorporating a 2,6-diaminopurine nucleobase in said peptide nucleic acid having formula (I):

![Chemical Structure](image)

wherein:

- each L is independently selected from a group consisting of naturally-occurring nucleobases and non-naturally-occurring nucleobases, provided that at least one L is 2,6-diaminopurine nucleobase;
- each R^L is independently hydrogen or C_1-C_8 alkylamyl; R^L is OH, NH_2 or NH-LysNH_2;
- R^H is H, COCH_3 or t-butoxycarbonyl; and n is an integer from 1 to 30.

Preferably, R^L is C_2-C_6 alkylamine. More preferably, R^L is C_2-C_4 alkylamine. Still more preferably, R^L is butylamine.

Preferably, at least one of the R^L is C_1-C_8 alkylamine. Preferably, at least one of the R^L is C_2-C_4 alkylamine. More preferably, at least one of the R^L is C_2-C_4 alkylamine. Still more preferably, at least one of the R^L is butylamine. Even more preferably, substantially all of the R^L is butylamine.

Preferably, the carbon atom to which substituent R^L is attached is stereochemically enriched. Hereinafter, "stereochemically enriched" means that one stereoisomer is present more than the other stereoisomer in a sufficient amount so as to provide a beneficial effect. Preferably, one stereoisomer is present by more than 50%. More preferably, one stereoisomer is present by more than 80%. Even more preferably, one stereoisomer is present by more than 90%. Still more preferably, one stereoisomer is present by more than 95%. Even more preferably, one stereoisomer is present by more than 99%. Still even more preferably, one stereoisomer is present in substantially quantitatively. Preferably, the stereochemical enrichment is of R configuration.

Preferably, the peptide nucleic acid (I) of the present invention is derived from an amino acid. More preferably, the peptide nucleic acid (I) of the present invention is derived from D-lysine.

The present invention also provides a method for enhancing the DNA or RNA binding affinity of a peptide nucleic acid by incorporating a 2,6-diaminopurine nucleobase in said peptide nucleic acid having formula (I).

The PNAs of the present invention are synthesized by adaptation of standard peptide synthesis procedures, either in solution or on a solid phase.

Monomer subunits of the present invention or their activated derivatives, protected by standard protecting groups, are specially designed amino acid compounds having formula (II):

![Chemical Structure](image)

wherein:

- is a 2,6-diaminopurine nucleobase;
- R^7 is hydrogen or C_1-C_8 alkylamine;
- E is COOH or an activated or protected derivative thereof; and
- Z is NH_2 or NH-Pg, where Pg is an amino-protecting group.

Preferably, R^7 is C_4-C_6 alkylamine. More preferably, R^7 is C_4-C_5 alkylamine. Even more preferably, R^7 is butylamine.

The carbon atom to which substituent R^7 is attached (identified by an asterisk) is stereochemically enriched. Preferably, the stereochemical enrichment is of R configuration.

Preferably, the compound (II) of the present invention is derived from an amino acid.

More preferably, the compound (II) of the present invention is derived from D-lysine.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1(a) and (b) provide examples of naturally-occurring and non-naturally-occurring nucleobases for RNA recognition.

FIG. 2 shows the Acr^1 ligand and a PNA, Acr^1-(Tae,)^10-Lys-NH_2.

FIG. 3 provides a general scheme for solid phase PNA synthesis illustrating the preparation of linear unprotected PNA amides.

FIG. 4 provides a procedure for the synthesis of protected PNA synthons.

FIG. 5 provides a procedure for synthesis of thymine monomer synthons with side chains corresponding to the common amino acids.

FIG. 6 provides a procedure for synthesis of an aminoethyl-β-alanine analogue of thymine monomer synthon.

FIGS. 7(a), 7(b) and 7(c) are schematics showing the synthesis of PNA monomers containing l-lysine.

FIG. 8 is a graph showing inhibition of HCV protein translation in an in vitro translation assay.
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DETAILED DESCRIPTION OF THE
INVENTION

In the PNAs of the present invention having the formula (I), nucleobase L is a naturally-occurring nucleobase
attached at the position found in nature, i.e., position 9 for
adenine or guanine, and position 1 for thymine or cytosine,
a non-naturally-occurring nucleobase (nucleobase analog)
or a nucleobase-binding moiety, at least one of said ligands
1. being a 2,6-diaminopurine nucleobase. Exemplary nucleo-
bases include adenine, guanine, thymine, cytosine, 2,6-
diaminopurine, uracil, thiouracil, inosine, 5-methylcytosine,
bromothymine, azadenines and azaguanines. Some typical
nucleobases and illustrative synthetic nucleobases are
shown in FIGS. 1(a) and 1(b).

In monomer subunits according to the present invention
having the formula (II), L is a naturally-occurring nucleo-
base or a non-naturally-occurring nucleobase which may be
protected with one or more protecting groups. Exemplary
protecting groups include t-butoxycarbonyl (BOC), fluore-
nethylmethoxy carbonyl (FMOC) or 2-nitrobenzyl (2Nb).
Accordingly, such protecting groups may be either acid,
base, hydrogenolytic or photolytically labile.

Preferably R₇ in the monomer subunit is hydrogen or
C₁₋₆ alkylamine.

Preferably, E in the monomer subunit is COOH or an
activated derivative thereof. Activation may, for example,
be achieved using an acid anhydride or an active ester deriva-
tive.

The amino acids which form the polyamide backbone
may be identical or different. We have found that those based
on 2-aminoethyglycine are particularly useful in the present
invention.

The PNAs of the present invention may be linked to low
molecular weight effector ligands, such as ligands having
nuclease activity or alkylating activity or reporter ligands
(e.g., fluorescent, spin labels, radioactive, protein recogni-
tion ligands, for example, biotin or hapten). PNAs may also
be linked to peptides or proteins, where the peptides have
signaling activity. Exemplary proteins include enzymes,
transcription factors and antibodies. The PNAs of the
present invention may also be attached to water-soluble
polymers, water-insoluble polymers, oligonucleotides or
carbohydrates. When warranted, a PNA oligomer may be
synthesized onto a moiety (e.g., a peptide chain, reporter,
terpolymer or other type of ligand-containing group) attached
to a solid support.

In monomer subunits according to the present invention
having the formula (II), L is a naturally-occurring nucleo-
base or a non-naturally-occurring nucleobase which may be
protected with one or more protecting groups.

A compound according to the present invention having
general formula (I), L is a 2,6-diaminopurine nucleobase
which may be protected with one or more protecting groups.
Exemplary protecting groups include t-butoxycarbonyl (BOC), fluore-
nethylmethoxy carbonyl (FMOC) or 2-nitrobenzyl (2Nb). Accordingly, such protecting groups
may be either acid, base, hydrogenolytic or photolytically
labile.

Preferably R₇ is independently hydrogen or C₁₋₆ alkyl-
amine.

Preferably, E in the monomer subunit is COOH or an
activated derivative thereof. Activation may, for example, be
achieved using an acid anhydride or an active ester deriva-
tive.

The amino acids which form the polyamide backbone
may be identical or different. We have found that those based
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The PNAs of the present invention may be linked to low
molecular weight effector ligands, such as ligands having
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tion ligands, for example, biotin or hapten). PNAs may also
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signaling activity. Exemplary proteins include enzymes,
transcription factors and antibodies. The PNAs of the
present invention may also be attached to water-soluble
polymers, water-insoluble polymers, oligonucleotides or
carbohydrates. When warranted, a PNA oligomer may be
synthesized onto a moiety (e.g., a peptide chain, reporter,
terpolymer or other type of ligand-containing group) attached
to a solid support.

The PNAs of the present invention may be used for gene
modulation (e.g., gene targeted drugs), diagnostics, biotech-
nology and other research purposes. The PNAs may also
be used to target RNA and single stranded DNA (ssDNA) to
produce both antisense-type gene regulating moieties and as
hybridization probes, e.g., for the identification and purifi-
cation of nucleic acids. Furthermore, the PNAs may be
modified in such a way that they form triple helices with
double stranded DNA (dsDNA). Compounds that bind
sequence-specifically to dsDNA have applications as gene
targeted drugs. These compounds are extremely useful drugs
for treating diseases such as cancer, acquired immune defi-
cency syndrome (AIDS) and other virus infections and
genetic disorders. Furthermore, these compounds may be
used in research, diagnostics and for detection and isolation
of specific nucleic acids.

Gene-targeted drugs are designed with a nucleobase
sequence (preferably containing 10–20 units) complementary
to the regulatory region (the promoter) of the target
gene. Therefore, upon administration, the gene-targeted
drugs bind to the promoter and prevent RNA polymerase
from accessing the promoter. Consequently, no mRNA, and
thus no gene product (protein), is produced. If the target is
within a vital gene for a virus, no viable virus particles will
be produced. Alternatively, the target region could be down-
stream from the promoter, causing the RNA polymerase to
terminate at this position, thus forming a truncated mRNA/protein which is nonfunctional.

Sequence-specific recognition of ssDNA by base com-
plementary hybridization can likewise be exploited to target
specific genes and viruses. In this case, the target sequence
is contained in the mRNA such that binding of the drug to the
target hinders the action of ribosomes and, consequently,
translation of the mRNA into a protein. The PNAs of the
present invention have higher affinity for complementary
ssDNA than other currently available oligonucleotide ana-
logs. Also PNAs of the present invention need not possess
a net charge and can bear substituents that enhance aqueous
solubility, which facilitates cellular uptake. In addition, the
PNAs of the present invention contain amides of non-
biochemical amino acids, which make them biostable and
resistant to enzymatic degradation.

The PNAs of the present invention comprising C₁₋₆ alkyl-
amine side chains exhibit enhanced binding affinity.
This is demonstrated by increased thermal stability of the
complex formed between said compounds of the present
invention and a complementary DNA strand. The PNAs of
the present invention also exhibit enhanced solubility and
sequence specificity in binding to complementary nucleic
acids.

A synthesis of PNAs according to the present invention is
discussed in detail below.
Synthesis of PNA Oligomers

The principle of anchoring molecules during a reaction onto a solid matrix is known as Solid Phase Synthesis or Merrifield Synthesis (see Merrifield, *J. Am. Chem. Soc.*, 1963, 85, 2149 and *Science*, 1986, 232, 341). Established methods for the stepwise or fragment-wise solid-phase assembly of amino acids into peptides normally employ a bead matrix of cross-linked styrene-divinylbenzene copolymer. The cross-linked copolymer is formed by the pearl polymerization of styrene monomer to which is added a mixture of divinylbenzenes. Usually, 1-2% cross-linking is employed. Such a matrix may be used in solid phase PNA synthesis of the present invention (FIG. 3).

More than fifty methods for initial functionalization of the solid phase have been described in connection with traditional solid phase peptide synthesis (see Barany and Merrifield in "The Peptides" Vol. 2. Academic Press, New York. 1979, pp. 1-284, and Stewart and Young, "Solid Phase Peptide Synthesis", 2nd Ed., Pierce Chemical Company, Ill., 1984). Reactions for the introduction of chloromethyl functionality (Merrifield resin; via a chloromethyl methyl ether, SnCl4 reaction), aminomethyl functionality (via an N-hydroxymethylphthalimide reaction; Mitchell et al., *Tetrahedron Lett.*, 1976, 3795) and benzhydrylamino functionality (Pieta et al., *J. Chem. Soc., 1970, 650*) are most widely used. Regardless of its nature, the purpose of introducing a functionality on the solid phase is to form an anchoring linkage between the copolymer solid support and the C-terminus of the first amino acid to be coupled to the solid support. As will be recognized, anchoring linkages may also be formed between the solid support and the amino acid N-terminus. The "concentration" of a functional group present in the solid phase is generally expressed in millimoles per gram (mmol/g). Other reactive functionalities which have been initially introduced include 4-methylbenzhydrylamine and 4-methoxybenzhydrylaminogroups. All of these established methods are, in principle, useful within the context of the present invention.

A preferred method for PNA synthesis employs aminomethyl as the initial functionality. Aminomethyl is particularly advantageous as a "spacer" or "handle" group because it forms amide bonds with a carboxylic acid group in nearly quantitative amounts. A vast number of reactive spacer- or handle-forming bifunctional reagents have been described (see Barany et al., *Int. J. Peptide Protein Res.*, 1987, 30, 705). Representative bifunctional reagents include 4-(haloalky)aryl-lower alkanolic acids such as 4-[(bromomethyl)phenylacetic acid; BOC-aminocarbonyl-4-(oxymethyl)aryl-lower alkanolic acids such as BOC-aminocarbonyl-4-(oxymethyl)phenylacetic acid; N-BOC-p-acylbenzhydrylamines such as N-BOC-p-glutaroylbenzhydrylamine; N-BOC-4'-lower alkyl-p-acylbenzhydrylamines such as N-BOC-4'-methyl-p-glutaroylbenzhydrylamine; N-BOC-4'-lower alkoxy-p-acylbenzhydrylamines such as N-BOC-4'-methoxy-p-glutaroylbenzhydrylamine; and 4-hydroxymethylphenoxycetic acid. One type of spacer group particularly relevant within the context of the present invention is fluroacetic acid (FMA) handle (Mitchell and Merrifield, *J. Org. Chem.*, 1976, 41, 2015) which, deriving from the electron withdrawing effect of the 4-phenylacetamidomethyl group, is about 100 times more stable than a benzyl ester linkage towards the BOC-amin depredication reagent trifluoroacetic acid (TFA).

Certain functionalities (e.g., benzhydrylamo, 4-methylbenzhydrylamo and 4-methoxybenzhydrylamo), which may be incorporated for the purpose of cleavage of a synthesized PNA chain from the solid support such that the C-terminal of the PNA chain is released as an amide, require no introduction of a spacer group. Any such functionality may advantageously be employed in the context of the present invention.

An alternative strategy concerning the introduction of spacer or handle groups is the so-called "preformed handle" strategy (see Tam et al., *Synthesis*, 1979, 955-957), which offers complete control over coupling of the first amino acid and excludes the possibility of complications arising from the presence of undesired functional groups not related to the peptide or PNA synthesis. In this strategy, spacer or handle groups, of the same type as described above, are reacted with the first amino acid desired to be bound to the solid support, the amino acid being N-protected and optionally protected at the other side chains which are not relevant with respect to the growth of the desired PNA chain. Thus, in those cases in which a spacer or handle group is desirable, the first amino acid to be coupled to the solid support can either be coupled to the free reactive end of a spacer group which has been bound to the initially introduced functionality (for example, an aminomethyl group) or can be reacted with the spacer-forming reagent. The space-forming reagent is then reacted with the initially introduced functionality. Other useful anchoring schemes include the "multidetachable" resins (see Tam et al., *Tetrahedron Lett.*, 1979, 4935 and *J. Am. Chem. Soc.*, 1980, 102, 611; Tam, J. Org. Chem., 1985, 50, 5291), which provide more than one mode of release and thereby allow more flexibility in synthetic design.


In addition to such amino-protecting groups, nonurethane-type of amino-protecting groups are also applicable when assembling PNA molecules. Thus, not only the above-mentioned amino-protecting groups (or those derived from any of these groups) are useful within the context of the present invention, but so are virtually any amino-protecting groups which largely fulfill the following requirements: (1) stable to mild acids (not significantly attacked by carboxyl groups); (2) stable to mild bases or nucleophiles (not significantly attacked by the nucleophile); (3) resistant to acylation (not significantly attacked by activated amino acids); (4) can be substantially removed without any serious side reaction; and (5) preserves the optical integrity, if any, of the incoming amino acid upon coupling.

The choice of side chain protecting groups, in general, depends on the choice of the amino-protecting group, because the side chain protecting group must withstand the conditions of the repeated amino depredication cycles. This is
true whether the overall strategy for chemically assembling PNA molecules relies on, for example, different acid stability of amino and side chain protecting groups (such as is the case for the above-mentioned "BOC-benzyl" approach) or employs an orthogonal, that is, chemoselective, protection scheme (such as is the case for the above-mentioned "FMC-c-But" approach).

Polymerization of the first amino acid, the next stage of solid phase synthesis is the systematic elaboration of the desired PNA chain. This elaboration involves repeated deprotection/coupling cycles. A temporary protecting group, such as BOC or FMC, on the last coupled amino acid is quantitatively removed by a suitable treatment, for example, by acidolysis, such as with trifluoroacetic acid in the case of BOC, or by base treatment, such as with piperidine in the case of FMC, so as to liberate the N-terminal amine function.

The next desired N-protected amino acid is then coupled to the N-terminal of the last coupled amino acid. This coupling of the C-terminal of an amino acid with the N-terminal of the last coupled amino acid can be achieved in several ways. For example, it can be achieved by providing the incoming amino acid in a form with the carboxyl group activated by any of several methods, including the initial formation of an active ester derivative such as a 2,4,5-trichlorophenyl ester (Pliss et al., Helv Chim Acta, 1963. 46, 1609), a phthalimido ester (Nefkens et al., J. Am. Chem. Soc., 1961. 83. 1263), a pentachlorophenyl ester (Kupyszewski, Rocq. Chem., 1969. 35. 595), a pentfluorophenyl ester (Kovacs et al., J. Am. Chem. Soc., 1963. 85. 183), an o-nitrophenyl ester (Bodanszky, Nature, 1955. 175, 685), an amidazole ester (Li et al., J. Am. Chem. Soc., 1970. 92. 7608), and a 3-hydroxy-4-oxo-3,4-dihydroquinazoline (Dht(OH) ester (Konig et al., Chem. Ber., 1973. 103, 2024 and 2034), or the initial formation of an anhydride such as a symmetrical anhydride (Wieland et al. Angew. Chem., Int. Ed. Engl., 1971. 10. 336). Alternatively, the carboxyl group of the incoming amino acid can be reacted directly with the N-terminus of the last coupled amino acid with the assistance of a condensation reagent such as, for example, dicyclohexylcarbodiimide (Sheehan et al., J. Am. Chem. Soc., 1957. 77, 1067) derivatives thereof. Benzotraziolyl N-oxytridimethylammonophenonium hexafluorophosphat (BOP), "Castro's reagent" (see Rivaille et al., Tetrahedron, 1980. 36, 3413), is recommended when assembling PNA molecules containing secondary amino groups. Finally, activated PNA monomers analogous to the recently-reported amino acid fluorides (Carpino, J. Am. Chem. Soc., 1990. 112. 9651) hold considerable promise to be used in PNA synthesis as well.

Following the assembly of the desired PNA chain, including protecting groups, the next step will normally be deprotection of the amino acid moieties of the PNA chain and cleavage of the synthesized PNA from the solid support. These processes can take place substantially simultaneously, thereby providing the free PNA molecule in the desired form. Alternatively, in cases in which condensation of two separately synthesized PNA chains is to be carried out, it is possible, by choosing a suitable spacer group at the start of the synthesis, to cleave the desired PNA chains from their respective solid supports (both peptide chains still incorporating their side chain-protecting groups) and finally removing the side chain-protecting groups after, for example, coupling the two side chain-protected peptide chains to form a longer PNA chain.

In the above-mentioned "BOC-benzyl" protection scheme, the final deprotection of side chains and release of the PNA molecule from the solid support is most often carried out by the use of strong acids such as hydroxyl HF (Sakakibara et al. Bull. Chem. Soc. Jpn., 1965. 38, 4921), boron tris (trifluoroacetate) (Pliss et al., Helv Chim Acta, 1973. 46. 1609) and sulfonic acids, such as trifluoromethanesulfonic acid and methanesulfonic acid (Yajima et al., J. Chem. Soc. Chem. Comm., 1974. 1077). A strong acid (e.g. anhydrous HF) deprotection method may produce very reactive carbocations that may lead to alkylation and introduction of sensitive residues in the PNA chain. Such side reactions are only partly avoided by the presence of scavengers such as anisole, phenol, dimethyl sulfoxide, and mercaptoethanol. Thus, the sulfide-assisted acidoletic S,2 deprotection method (Tam et al., J. Am. Chem. Soc., 1983. 105. 6442 and J. Am. Chem. Soc., 1986. 108. 5242), the so-called "low" method, which removes the precursors of harmful carbocations to form inert sulfonium salts, is frequently employed in peptide and PNA synthesis. Other methods for deprotection and/or final cleavage of the PNA solid support bond may include base-catalyzed alcoholysis (Barton et al., J. Am. Chem. Soc., 1973. 95, 4501), ammonolysis, hydrazinolysis (Bodanszky et al., Chem. Ind., 1964 1423), hydrolyslolysis (Jones, Tetrahedron Lett, 1977 2853 and Schlatter et al., Tetrahedron Lett. 1977 2861) and photolysis (Rich and Guwara, J. Am. Chem. Soc., 1975 97. 1575).

Finally, in contrast with the chemical synthesis of conventional peptides, stepwise chain building of chaedral PNAAs such as those based on aminooxyethylglycol backbone units, can start either from the N-terminus or the C-terminus. Those skills in the art will recognize that synthesis commencing at the C-terminus typically employ protected amine groups and free or activated acid groups, and syntheses commencing at the N-terminus typically employ protected acid groups and free or activated amine groups.

Based on the recognition that most operations are identical in the synthetic cycles of solid phase peptide synthesis (as is also the case for solid phase PNA synthesis), a new matrix, PEPS, was recently introduced (Berg et al., J. Am. Chem. Soc., 1989. 111, 8024 and International Patent Application WO 94(02749) to facilitate the preparation of a large number of peptides. This matrix is comprised of a polyethylene (PE) film with pendant long-chain polystyrene (PS) grafts (molecular weight on the order of 10^6 Daltons). The loading capacity of the film is as high as that of a beaded matrix, but PEPS has the additional flexibility to suit multiple syntheses simultaneously. Thus, in a new configuration for solid phase peptide synthesis, the PEPS film is fashioned in the form of discrete, labeled sheets, each serving as an individual compartment. During all the identical steps of the synthetic cycles, the sheets are kept together in a single reaction vessel to permit concurrent preparation of a multitude of peptides at a rate close to that of a single peptide synthesis by conventional methods. It is believed that the PEPS film support, comprising linker or spacer groups adapted to the particular chemistry will be particularly valuable in the synthesis of multiple PNA molecules. The synthesis of PNAAs are conceptually simple because only four different reaction compartments are normally required, one for each of the four "pseudo-nucleotide" units. The PEPS film support has been successfully tested in a number of PNA syntheses carried out in a parallel and substantially simultaneous fashion. The yield and quality of the products obtained from PEPS are comparable to those obtained by using the traditional polystyrene bead support. Also, experiments with other geometries of the PEPS polymer, for example, non-woven felt, knitted net, sticks and
microwell plates, have not indicated any limitations of the 
synthetic efficacy.

Two other methods for the simultaneous synthesis of large 
numbers of peptides also apply to the preparation of 
multiple, different PNA molecules. The first of these meth-
3958) utilizes acrylic acid-grafted polyethylene-rod- and 
96-microtool wells to immobilize the growing peptide 
chains and to perform the compartmentalized synthesis. 
While effective, this method is only applicable on a micro-
gram scale. The second method (Houghten, Proc. Natl. 
Acad. Sci. USA, 1985. 82. 5131) utilizes a "tea bag" con-
taining traditionally-used polymer beads. Other methods 
for multiple peptide or PNA synthesis in the context of the 
present invention include the simultaneous use of two dif-
f erent supports with different densities (Tregear in "Che-
Arbor Sci. Publ. Ann Arbor, 1972, pp. 175-178), combining 
reaction vessels via a manifold (Gorman, Anal. Biochem., 
1984. 136. 397), multicolumn solid phase synthesis (Krcn-
ak et al., Int. J. Protein Res., 1989. 33. 209, 
15 and Holm and Meldal in "Proceedings of the 20th European 
Peptide Symposium", G. Jung and E. Bayer, Eds., Walter 
de Gruyter & Co., Berlin, 1989, pp. 208-210) and the use of 

Conventional cross-linked styrene/divinylbenzene 
copolymer matrix and the PEPS support are preferred in 
the context of solid phase PNA synthesis. Other exemplary solid 
supports include (1) particles based upon copolymers of 
dimethylacrylamide cross-linked with N,N'- 

bisacryloylhexamethylene diamine. (2) solid supports based 
on silica-containing particles such as porous glass beads and 
silica gel. (3) composites that contain two major ingredients: 
a resin and another material that is also substantially inert 
to the reaction conditions employed (see Scott et al., J. Chrom.
1978. 17. 243; and van Riet-voschoten in "Peptides 1974", Y.
113-116) and (4) contiguous solid supports other than PEPS, 
such as cotton sheets (Lebl and Echler, Peptide Res., 1989. 
2. 232) and hydroxypropylacrylate-coated polypropylene 

Whether manually or automatically operated, solid phase 
PNA synthesis, in the context of the present invention, 
is normally performed batchwise. However, most of the syn-
theses may be carried out equally well in the continuous-
flow mode, where the support is packed into columns (Bayer 
et al., Tetrahedron Lett., 1970. 4503; and Scott et al., J.
Chromatogr. Sci., 1971. 9. 577). With respect to continuous-
flow solid phase synthesis, the rigid poly 
(dimethylacrylamide)-Kieselguhr support (Atherton et al. J.
particularly useful. Another useful configuration is the one 
worked out for the standard copoly(styrene-1-
divinylbenzene) support (Krcnak et al., Tetrahedron Lett., 

While the phase technique is preferred in the present 
invention, other methodologies or combinations thereof may 
also be used. Exemplary methodologies include (1) the 
classical solution phase methods for peptide synthesis (Bodansky, "Principles of Peptide Synthesis", Springer-
Verlag, Berlin-New York, 1984), either by stepwise assem-
by or by segment/fragment condensation, (2) the "liquid 
phase" strategy, which utilizes soluble polymeric supports 
such as linear polystyrene (Shemyakin et al., Tetrahedron 
Lett., 1965. 2323) and polyethylene glycol (PEG) (Mutter 
random polymerization (O’don, "Principles of 
mixtures of many molecular weights ("polydisperse") pept-
deptide PNA molecules and (4) a technique based on the use 
of polymer-supported amino acid active esters (Friskin et al.
J. Am. Chem. Soc., 1965. 87. 4646), sometimes referred to 
as "inverse Merrifield synthesis" or "polymeric reagent 
synthesis". In addition, it is envisaged that PNA molecules 
may be assembled enzymatically by enzymes such as pro-
teases or derivatives thereof with novel specificities 
(obtained, for example, by artificial means such as protein 
engineering). Also, one can envision the development of 
PNA ligases" for the condensation of a number of PNA 
fragments into very large PNA molecules. Also, since anti-
body can be generated to virtually any molecule of interest, 
the recently developed catalytic antibodies (enzymes), dis-
covered simultaneously by Tramontano et al. Science, 
1986. 234. 1566 and Pollack et al., Science, 1986. 234. 1570, 
should also be considered as potential candidates for assem-
bling PNA molecules. Thus, there has been considerable 
success in producing enzymes catalyzing acyl-transfer reac-
tions (see Shokat et al. Nature, 1989. 338. 269, and refer-
ences therein). Finally, completely artificial enzymes, very 
recently pioneered by Hahn et al. (Science, 1990. 248. 
1544), may be developed for PNA synthesis. The design of 
generically applicable enzymes, ligases, and catalytic 
antibodies, capable of mediating specific coupling reactions, 
should be more readily achieved for PNA synthesis than for 
"normal" peptide synthesis since PNA molecules will often 
be comprised of only four different amino acids (one for 
each of the four native nucleobases).

Likely therapeutic and prophylactic targets include herpes 
simplex virus (HSV), human papillomavirus (HPV), human 
immunodeficiency virus (HIV), candida albicans, influenza 
virus, cytomegalovirus (CMV), intercellular adhesion 
molecules (ICAM), 5-lipoxigenase (5-LO), phospholipase A, 
(protein kinase C (PKC), and the ras oncogene. Potential 
treatment of such targeting include ocular, labial, 
genital, and systemic herpes simplex I and II infections; 
genital warts; cervical cancer; common warts; Kaposi's 
sarcoma; AIDS; skin and systemic fungal infections; flu; 
pneumonia; reinitis and pneumonitis in immunosuppressed 
patients; mononucleosis; ocular, skin and systemic inflam-
mation; cardiovascular disease; cancer; asthma; psoriasis; 
cardiac valve collapse; cardiac infection; gastrointestinal 
disease; kidney disease; rheumatoid arthritis; osteoarthritis; 
acute pancreatitis; septic shock; and Crohn's disease.

In general, for therapeutic or prophylactic treatment, a 
patient suspected of requiring such therapy is administered 
a compound of the present invention, commonly in a phar-
caceutical acceptably carrier, in amounts and for periods 
of time which will vary depending upon the nature of the 
particular disease, it's severity and the patient's overall 
condition. The peptide nucleic acids of this invention can be 
formulated in a pharmaceutical composition, which may 
include carriers, thickeners, diluents, buffers, preservatives, 
surface-active agents and the like. Pharmaceutical compo-
sitions may also include one or more active ingredients such 
as antimicrobial agents, anti-inflammatory agents, anesthet-
ics and the like, in addition to the peptide nucleic acids.

The pharmaceutical composition may be administered in a 
number of ways depending upon whether local or systemic 
treatment is desired, and upon the area to be treated. Admin-
istration may be topical (including ophthalmic, vaginal, 
rectal, intranasal, transdermal), oral or parenteral, for 
example, by intravenous drip, subcutaneous, intraperitoneal
or intramuscular injection or intrathecal or intraventricular administration.

Formulations for topical administration may include transdermal patches, ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable. Coated condoms, gloves and the like may also be useful.

Compositions for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets, or tablets. Thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders may be added.

Compositions for intrathecal or intraventricular administration may include sterile aqueous solutions which may also contain buffers, diluents and other suitable additives.

Formulations for parenteral administration may include sterile aqueous solutions which may also contain buffers, diluents and other suitable additives.

Dosing is dependent on severity and responsiveness of the condition to be treated, but will normally be one or more doses per day, with the course of treatment lasting from several days to several months or until a cure is effected or a diminution of disease state is achieved. Persons of ordinary skill can easily determine optimum dosages, dosing methodologies and repetition rates.

The present invention also pertains to the advantageous use of PNA molecules in solid phase biochemistry (see "Solid Phase Biochemistry—Analytical and Synthetic Aspects", W. H. Scouf, Ed., John Wiley & Sons, New York, 1983), notably solid phase biosystems, especially bioassays or solid phase techniques for diagnostic detection/quantitation or affinity purification of complementary nucleic acids (see "Affinity Chromatography—A Practical Approach", P. D. G. Dean, W. S. Johnson and F. A. Middle, Eds., IRL Press Ltd. Oxford, 1986; "Nucleic Acid Hybridization—A Practical Approach", B. D. Hanes and S. J. Higgins, IRL Press Ltd., Oxford, 1987). Current methods for performing such bioassays or purification techniques almost exclusively utilize "normal" or slightly modified oligonucleotides either physically adsorbed or bound through a substantially permanent covalent anchoring linkage to beaded solid supports such as cellulose, glass beads, including those with controlled porosity (Mizutani et al., J. Chromatog, 1986, 356, 202), "Sephadex", "Sephacore", agarose, polyacrylamide, porous particulate alumina, hydroxyalkyl methacrylate gels, diol-bonded silica, porous ceramics, or contiguous materials such as filter discs of nylon and nitrocellulose.

All the above-mentioned methods are applicable within the context of the present invention. However, when possible, covalent linkage method is are preferred over the physical adsorption method, because the latter approach may result in some of the immobilized molecules being washed out (desorbed) during the hybridization or affinity process. The severity of this problem will, of course, depend to a large extent on the rate at which equilibrium between adsorbed and "free" species is established. In certain cases it may be virtually impossible to perform a quantitative assay with acceptable accuracy and/or reproducibility. The amount of loss of adsorbed species during the treatment of the support with body fluids, aqueous reagents or washing media will, in general, be expected to be most pronounced for species of relatively low molecular weight.

In contrast with oligonucleotides, PNA molecules are easier to attach onto solid supports because they contain strong nucleophilic and/or electrophilic centers. In addition, a direct assembly of oligonucleotides onto solid supports suffers from an extremely low loading of the immobilized molecule (Beaucage and Caruthers, Tetrahedron Lett., 1981, 22, 1859; and Caruthers, Science, 1985, 232, 281). In addition, because it uses the alternative phosphite triester method (Letsinger and Mahadevan, J. Am. Chem. Soc., 1976, 98, 3655), which is suited for solid supports with a high surface loading capacity, only relatively short oligonucleotides can be obtained.

As for conventional solid phase peptide synthesis, however, the latter supports are excellent materials for building up immobilized PNA molecules. It allows the side chain-protecting groups to be removed from the synthesized PNA chain without cleaving the anchoring linkage holding the chain to the solid support. They also can be loaded onto solid supports in large amounts, thus further increasing the capacity of the solid phase technique.

Furthermore, certain types of studies concerning the use of PNA in solid phase biochemistry can be conducted, facilitated, or greatly accelerated by use of the recently-reported "light-directed, spatially addressable, parallel chemical synthesis" technology (Fodor et al., Science, 1991, 251, 767), a technique that combines solid phase chemistry and photolithography to produce thousands of highly diverse, but identifiable, permanently immobilized compounds (such as peptides) in a substantially simultaneous way.

Synthesis of monomer synths

The monomer subunits preferably are synthesized by the general scheme outlined in FIG. 4. This involves preparation of either the methyl or ethyl ester of (BOC-aminoethyl) glycline, by a protection/deprotection procedure as described in Examples 20-22. The synthesis of thymine monomer is described in Examples 23-24, and the synthesis of protected tyrosine monomer is described in Example 25.

The synthesis of a protected adenine monomer involves alkylation of adenine with ethyl bromoacetate (Example 26) and verification of the position of substitution (i.e. position 9) by X-ray crystallography. The N9-amino group is then protected with the benzoyloxy carbonyl group by the use of the reagent N-ethyl-benzoyloxy carbonylimidazole tetrafluoroborate (Example 27). Simple hydrolysis of the product ester (Example 28) gave N9-benzoyloxy carbonyl-9-carboxymethyl adenine, which was used in the standard procedure (Examples 29-30). The adenine monomer has been built into two different PNA oligomers (Examples 52, 53, 56 and 57).

For the synthesis of the protected G-monomer, the starting material, 2-amino-6-chloropurine, was alkylated with bromoacetic acid (Example 31) and the chlorine atom was then substituted with a benzoyloxy group (Example 32). The resulting acid was coupled to the (BOC-aminoethyl)glycine methyl ester (from Example 22) with agent PyBropTM and the resulting ester was hydrolyzed (Example 33). The O5'-benzyl group was removed in the final HF-cleavage step in the synthesis of the PNA oligomer. Cleavage was verified by mass spectrometry of the final PNA oligomer, upon incorporation into a PNA oligomer using diisopropyl carbodiimide as the condensation agent (Examples 51 and 56).

Additional objects, advantages, and novel features of the present invention will become apparent to those skilled in the art upon examination of the following examples thereof, which are not intended to be limiting.

General Remarks

The following abbreviations are used in the experimental examples: DMF, N.N-dimethylformamide; Tyr, tyrosine;
Lys, lysine; DCC, N,N-dicyclohexyl-carbodiimide; DUC, N,N-dicyclohexylurea; THF, tetrahydrofuran; aeg, N-acetyl (2-aminoethyl)glycine; Pfp, pentafluorophenyl; BOC, tert-butoxy carbonyl; Z, benzyloxycarbonyl; NMR, nuclear magnetic resonance; s, singlet; d, doublet; dd, doublet of doublets; t, triplet; q, quartet; m, multiplet; b, broad; δ, chemical shift; ppm, parts per million (chemical shift).

NMR spectra were recorded on JEOL FX 90Q spectrometer or a Bruker 250 MHz with tetramethylsilane as an internal standard. Mass spectrometry was performed on a MassLab VG 12-250 quadrupole instrument fitted with a VG FAB source and probe. Melting points were recorded on a Buchi melting point apparatus and are uncorrected. N,N-Dimethylformamide was dried over 4 Å molecular sieves, distilled and stored over 4 Å molecular sieves. Pyridine (HPLC quality) was dried and stored over 4 Å molecular sieves. Other solvents used were either the highest quality obtainable or were distilled prior to use. Dioxane was passed through basic alumina prior to use. BOC-anhydride, 4-nitrophenol, methyl bromoacetate, benzyloxycarbonyl chloride, pentafluorophenol were all obtained from Aldrich Chemical Company. Thymine, cytosine, adenine were all obtained from Sigma.

Thin layer chromatography (tlc) was performed using the following solvent systems: (1) chloroform:triethyl amine:methanol. 7:1:2; (2) methylene chloride:methanol, 9:1; (3) chloroform:methanol:acetic acid 85:10:5. Spots were visualized by UV (254 nm) and/or spraying with a ninhydrin solution (3 g ninhydrin in 1000 mL of 1-butanol and 30 mL of acetic acid), after heating at 120° C. for 5 minutes and, after spraying, heating again.

EXAMPLE 1
Synthesis of tert-Butyl-4-nitrophenyl carbonate

Sodium carbonate (29.14 g, 0.275 mol) and 4-nitrophenol (12.75 g, 91.6 mmol) were mixed with dioxane (250 mL). BOC-anhydride (2 g, 91.6 mmol) was transferred to the mixture with dioxane (50 mL). The mixture was refluxed for 1 h. cooled to 0° C. filtered and concentrated to a third of the volume, and then poured into water (350 mL) at 0° C. After stirring for 0.5 h, the product was collected by filtration, washed with water, and then dried over secapent. in vacuo. Yield 21.3 g (97%). M.p. 73.0°-74.5° C. (lit. 78.5°-79.5° C.). Anal. for C_{14}H_{13}NO_{2}. Found (calc.) C: 55.20 (55.23); H: 5.62 (5.48); N: 5.82 (5.85).

EXAMPLE 2
Synthesis of N-BOC-2'-aminoethyglycine (2)
The title compound was prepared by a modification of the procedure by Heimer et al. Int. J. Pept., 1984, 23, 203-211. N-(2-Aminoethyglycine (1.3 g, 25.4 mmol) was dissolved in water (50 mL). dioxane (50 mL) was added, and the pH was adjusted to 11.2 with 2N sodium hydroxide. tert-Butyl-4-nitrophenyl carbonate (7.29 g, 30.5 mmol) was dissolved in dioxane (40 mL) and added dropwise over a period of 2 h, during which time the pH was maintained at 11.2 with 2N sodium hydroxide. The pH was adjusted periodically to 11.2 for three more hours and then the solution was allowed to stand overnight. The solution was cooled to 0° C. and the pH was carefully adjusted to 3.5 with 0.5M hydrochloric acid. The aqueous solution was washed with chloroform (3×200 mL), the pH adjusted to 9.5 with 2N sodium hydroxide and the solution was evaporated to dryness, in vacuo (14 mm Hg). The residue was extracted with DMSO (25×2×10 mL) and the extracts filtered to remove excess salt. This results in a solution of the title compound in about 60% yield and greater than 95% purity by tlc (system 1 and visualised with ninhydrin, Rf=0.3). The solution was used in the following preparations of BOC-aeg derivates without further purification.

EXAMPLE 3
Synthesis of N-1-Carboxymethylthymine (4)
This procedure is different from the literature synthesis, but is easier, gives higher yields, and leaves no unreacted thymine in the product. To a suspension of thymine (3.40 g, 0.317 mol) and potassium carbonate (87.7 g, 0.634 mmol) in DMSO (990 mL) was added methyl bromoacetate (30 mL, 0.317 mol). The mixture was stirred vigorously overnight under nitrogen. The mixture was filtered and evaporated to dryness, in vacuo. The solid residue was treated with water (350 mL) and 4N hydrochloric acid (12 mL), stirred for 15 minutes at 0° C., filtered, and washed with water (2×75 mL). The precipitate was treated with water (120 mL) and 2N sodium hydroxide (60 mL), and was refluxed for 10 minutes. The mixture was cooled to 0° C. filtered, and pure title compound was precipitated by the addition of 4N hydrochloric acid (70 mL). The yield after drying, in vacuo over secapent was 37.1 g (64%). "H-NMR: (90 MHz; DMSO-d_{6}); 11.33 ppm (s, 1H, NH); 7.49 (d, J=0.92 Hz, 1H, ArH); 4.38 (s, 2H, CH_{2}); 1.76 (d, J=0.92 Hz, T-CH_{3}).

EXAMPLE 4
Synthesis of N-1-Carboxymethylthymine pentfluorophenyl ester (5)
N-1-Carboxymethylthymine (4.10 g, 54.3 mmol) and pentafluorophenol (10 g, 54.3 mmol) were dissolved in DMSO (100 mL) and cooled to 5° C. in ice water. DCC (13.45 g, 65.2 mmol) was added. When the temperature decreased below 5° C., the ice bath was removed and the mixture was stirred for 3 h at ambient temperature. The precipitated DCC was removed by filtration and washed twice with DMF (2×10 mL). The combined filtrate was poured into ether (1400 mL) and cooled to 0° C. Petroleum ether (1400 mL) was added and the mixture was left overnight. The title compound was isolated by filtration and washed with petroleum ether. Yield: 14.8 g (78%). The product was pure enough to carry out the next reaction, but an analytical sample was obtained by recrystallization from 2-propanol. M.p. 200°-206° C. Anal. for C_{14}H_{7}F_{5}NO_{2}. Found (calc.) C: 44.79(44.59); H: 2.14(2.01) N: 8.13(8.00). FAB-MS: 443 (M+1; glycerol), 351 (M+1). "H-NMR (90 MHz; DMSO-d_{6}); 11.52 ppm (s, 1H, NH); 7.64 (s, 1H, ArH); 4.99 (s, 2H, CH); 1.76 (s, 3H, CH_{3}).

EXAMPLE 5
Synthesis of 1-(BOC-aeg)thymine (6)
To a DMF solution of product of Example 2 was added triethyl amine (7.08 mL, 50.8 mmol) followed by N-1-carboxymethylthymine pentafluorophenyl ester (5. 4.45 g, 12.7 mmol). The resultant solution was stirred for 1 h. The solution was cooled to 0° C. and treated with cation exchange material ("Dowex 50W X-8", 40 g) for 20 minutes. The cation exchange material was removed by filtration, washed with dichloromethane (2×15 mL), and the filtrate was diluted further with dichloromethane (150 mL). The resulting solution was washed with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, in vacuo, first by a water aspirator and then by an oil pump. The residue was shaken with water (50 mL) and evaporated to dryness. This procedure was repeated again.
The residue then was dissolved in methanol (75 mL) and poured into ether (600 mL) and petroleum ether (1400 mL). After stirring overnight, the white solid was isolated by filtration and was washed with petroleum ether. Drying over dichromate in vacuo, the mass was 35.0 g (71.7% of the title compound). M.p. 142°-147° C. Anal. for C_{14}H_{23}N_{2}O_{5} Found (calc.) C: 49.59(50.00) H: 6.34(6.29) N: 14.58(14.58). 1H-NMR (250 MHz, DMSO-d_{6}): Due to the limited rotation around the secondary amide bond several of the signals were doubled in the ratio 2:1 (indicated in the list by m for major and s for minor); 12.73 ppm (t, 1H, CO-CH₃); 11.27 ppm (s, m, imide); 7.30 ppm (s, m, ArH); 3.72 ppm (s, m, ArH); 6.92 ppm (unres. t, m, BOC-NH); 3.53 ppm (unres. t, m, BOC-NH); 4.63 ppm (s, m, T-CH$_2$-CO-); 4.47 ppm (s, m, T-CH$_2$-CO-); 4.19 ppm (s, m, CONRCH$_2$COH); 3.97 ppm (s, m, CONH$_2$COH); 3.41-2.89 ppm (unres. m, CH$_2$CH$_2$- and water); 1.75 ppm (s, 3H, T-CH$_3$); 1.36 ppm (s, 9H, t-Bu). 31P-NMR: 170.68 ppm (CO); 170.34 (CO); 167.47 (CO); 167.06 (CO); 164.29 (CO); 159.0 (C$_5$'); 141.92 (C$_6$'); 106.34 (C$_7$'); 77.96 and 77.68 (Thy-CH$_3$); 48.90, 47.45 and 46.70 (THF, CH$_2$); and NCH$_2$COH); 37.98 (THF, CH$_3$); 28.07 (t-Bu). FAB-MS: 407 (M+Na$^+$); 385 (M+H$^+$).

EXAMPLE 6
Synthesis of 1-(BOC-ethylamine) pentafluorophenyl ester (7, BOC-TaqbOPFp)

1-(BOC-ethylamine) (6) (2 g, 5.20 mmol) was dissolved in DMF (5 mL) and methylene chloride (15 mL) was added. Pentafluorophenol (1.05 g, 5.72 mmol) was added and the solution was cooled to 0° C. After 2 minutes, the precipitated DCU was removed by filtration and washed with methylene chloride. The combined filtrate was washed with saturated sodium carbonate and washed with water, evaporated to dryness, and the residue was dissolved in dioxane (150 mL) and poured into water (200 mL) at 0° C. The title compound was isolated by filtration, washed with water, and dried (60% yield). The title compound was recrystallized from 2-propanol. M.p. 174°-175.5° C. Analysis for C$_{14}$H$_{26}$N$_{2}$O$_{5}$P (calc.) C: 48.22(48.01); H: 4.64(4.21); N: 9.67(10.18). 1H-NMR (250 MHz, CDCl$_3$): Due to the limited rotation around the secondary amide bond several of the signals were doubled in the ratio 6:1 (indicated in the list by m for major and s for minor); 7.01 ppm (s, m, ArH); 6.99 ppm (s, m, ArH); 5.27 ppm (unres. t, BOC-NH); 4.67 ppm (s, m, T-CH$_2$-CO-); 4.60 ppm (s, m, T-CH$_2$-CO-); 4.45 ppm (s, m, CONRCH$_2$COOPf); 4.42 ppm (s, m, CONRCH$_2$COOPf); 3.64 ppm (t, 2H, BOC-NHCH$_2$CH$_2$-); 3.87 ppm ("q", 2H, BOC-NHCH$_2$CH$_2$-); 1.44 ppm (5H, t-Bu). FAB-MS: 551 (10); M+1); 495 (10); M+1Bu); 451 (80); -BOC.

EXAMPLE 7
Synthesis of N$_2$-Benzoylcarbonyl cytosine (9)

Over a period of about 1h, benzoylcarbonyl chloride (52 mL, 0.36 mol) was added dropwise to a suspension of cytosine (8, 20 g, 0.18 mol) in dry pyridine (1000 mL) at 0° C. Under nitrogen in oven-dried equipment. The solution was then stirred overnight, after which the pyridine suspension was evaporated to dryness in vacuo. Water (200 mL) and 4N hydrochloric acid were added to reach pH 1. The resulting white precipitate was filtered off, washed with water and partially dried by air suction. The moist precipitate was reflushed with absolute ethanol (500 mL) for 10 minutes, cooled to 0° C, filtered, washed with ether, and dried in vacuo. Yield 24.7 g (54%). M.p. 250° C. Anal. for C$_{14}$H$_{13}$N$_{2}$O$_{7}$ Found (calc.) C: 58.59(58.77); H: 4.55(4.52); N: 17.17(17.13). No NMR spectra were recorded since it was not possible to get the product dissolved.

EXAMPLE 8
Synthesis of N$_2$-Benzoylcarbonyl-N$_2$-carboxymethyl cytosine (10)

In a three-necked round bottom flask equipped with a mechanical stirrer and a nitrogen inlet was placed methyl bromoacetate (7.82 mL, 82.6 mmol) and a suspension of N$_2$-benzoylcarbonyl-cytosine (9, 21 g, 82.6 mmol) and potassium carbonate (11.4 g, 82.6 mmol) in dry DMF (900 mL). The mixture was stirred vigorously overnight, filtered, and evaporated to dryness, in vacuo. Water (300 mL) and 4N hydrochloric acid (10 mL) were added, the mixture was stirred for 15 minutes at 0° C, filtered, and washed with water (2x75 mL). The isolated precipitate was treated with water (120 mL). 2N sodium hydroxide (60 mL), stirred for 30 minutes, filtered, cooled to 0° C, and 4N hydrochloric acid (35 mL) was added. The title compound was isolated by filtration, washed with water, recrystallized from methanol (1000 mL) and washed with ether. This afforded 7.70 g (31%) of pure title compound. The mother liquor from recrystallization was reduced to a volume of 200 mL and cooled to 0° C. This afforded an additional 2.30 g of a material that was pure by tlc but had a reddish color. M.p. 266°-274° C. Anal. for C$_{14}$H$_{13}$N$_{2}$O$_{7}$ Found (calc.) C: 55.41 (55.45); H: 4.23(4.32); N: 14.04(13.86). 1H-NMR (90 MHz; DMSO-d$_6$): 8.02 ppm (d, J=7.32Hz, 1H, H-6); 7.39 (s, 5H, Ph); 7.01 (d, J=7.32Hz, 1H, H-5); 5.19 (s, 2H, PhCH$_2$-); 4.52 (s, 2H).

EXAMPLE 9
Synthesis of N$_2$-Benzoylcarbonyl-N$_2$-carboxymethyl-cytosine pentafluorophenyl ester (11)

N$_2$-Benzoylcarbonyl-N$_2$-carboxymethyl-cytosine (10, 4 g, 13.2 mmol) and pentafluorophenol (2.67 g, 14.5 mmol) were mixed with DMF (70 mL), cooled to 0° C, with ice-water, and DCC (3.27 g, 15.8 mmol) was added. The ice bath was removed after 3 minutes and the mixture was stirred for 3 h at room temperature. The precipitated DCU was removed by filtration, washed with DMF, and the filtrate was evaporated to dryness, in vacuo (0.2 mm Hg). The solid residue was treated with methylene chloride (250 mL). Stirred vigorously for 15 minutes, filtered, washed twice with diluted sodium hydrogen carbonate and once with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness. The solid residue was recrystallized from 2-propanol (150 mL) and the crystals were washed with ether. Yield 3.40 g (55%). M.p. 241°-245° C. Anal. for C$_{14}$H$_{13}$N$_{2}$F$_{5}$O$_{2}$ Found (calc.) C: 51.56(51.18); H: 2.77(2.58); N: 9.24(8.95). 1H-NMR (90 MHz; CDCl$_3$); 7.66 ppm (d, J=7.63Hz, 1H, H-6); 7.37 (s, 5H, Ph); 7.31 (d, J=7.63Hz, 1H, H-5); 5.21 (s, 2H, PhCH$_2$-); 4.97 (s, 2H, NCH$_2$-). FAB-MS: 470 (M+41).

EXAMPLE 10
Synthesis of N$_2$-Benzoylcarbonyl-1-BOC-aeg-cytosine (12)

To a solution of (N-BOC-2-aminoethylic)glycine (2) in DMF, prepared as described above, was added triethylamine.
amine (7 mL, 50.8 mmol) and N^4-benzoyloxycarbonyl-N^1-carboxymethyl-cytosine pentfluorophenyl ester (11, 2.7 g, 5.75 mmol). After stirring the solution for 1 h at room temperature, methylene chloride (150 mL), saturated sodium chloride (250 mL), and 4N hydrochloric acid to pH 1 were added. The organic layer was separated and washed twice with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, in vacuo, first with a water aspirator and then with an oil pump. The oily residue was treated with water (25 mL) and was again evaporated to dryness, in vacuo. This procedure was then repeated. The oily residue (2.8 g) was then dissolved in methylene chloride (100 mL), petroleum ether (250 mL) was added, and the mixture was stirred overnight. The title compound was isolated by filtration and washed with petroleum ether. TLC (system 1) indicated substantial quantities of pentfluorophenol, but no attempt was made to remove it. Yield: 1.72 g (59%). M.p. 156° C. (decomp.). 1H-NMR (250 MHz, CDCl3). Due to the limited rotation around the secondary amide bond several of the signals were doubled in the ratio 2:1 (indicated in the list by mj for major and mi for minor): 7.88 ppm (dd, 1H, H-6); 7.39 (m, 5H, Ph); 7.00 (dd, 1H, H-5); 6.92 (1H, BOC-NH); 6.74 (1H, ZNH-2); 5.19 (s, 2H, Ph-CH2); 4.81 ppm (s, mj, Cyt-CH2-CO—); 4.62 ppm (s, mi, Cyt-CH2-CO—); 4.23 (s, mi, CONHCO2H); 3.98 ppm (s, mj, CONRCH2CO2H); 3.42-3.02 (unres. m, —CH2N— and water); 1.37 (s, 9H, t-Bu). FAB-MS: 504 (M+1); 448 (M+1-t-Bu).

EXEMPLARY 11

Synthesis of N^4-Benzoyloxycarbonyl-1-BOC-aeg-cytosine pentfluorophenyl ester (13)

N^4-Benzoyloxycarbonyl-1-BOC-aeg-cytosine (12, 1.5 g, 2.98 mmol) and pentfluorophenol (548 mg, 2.98 mmol) was dissolved in DMF (10 mL). Methylene chloride (10 mL) was added. The reaction mixture was cooled to 0° C in an ice bath, and DCC (676 mg, 3.28 mmol) was added. The ice bath was removed after 3 minutes and the mixture was stirred for 3 h at ambient temperature. The precipitate was isolated by filtration and washed once with methylene chloride. The precipitate was dissolved in boiling dioxane (150 mL) and the solution was cooled to 15° C, whereby DCC precipitated. The precipitated DCC was removed by filtration and the resulting filtrate was poured into water (250 mL) at 0° C. The title compound was isolated by filtration, washed with water, and dried over scapentane in vacuo. Yield 1.30 g (65%). Analysis for C23H22N2O4F5. Found (calc.): C: 52.63(52.02); H: 4.41(4.22); N: 10.55(10.46).

1H-NMR (250 MHz, DMSO-d6): showed essentially the spectrum of the above acid, most probably due to hydrolysis of the ester. FAB-MS: 670 (M+1); 614 (M+1-t-Bu).

EXEMPLARY 12

Synthesis of 4-chloroacarbonyl-9-chloroacridine

4-Chloracarboline (6.25 g, 26.1 mmol), thionyl chloride (25 mL) and 4 drops of DMF were heated gently under a flow of nitrogen until all solid material had dissolved. The solution then was refluxed for 40 minutes. The solution was cooled and excess thionyl chloride was removed in vacuo. The last traces of thionyl chloride were removed by coevaporation with dry benzene (dried over Na-Pb) twice. The remaining yellow powder was used directly in the next reaction.

EXEMPLARY 13

Synthesis of 4-(5-methoxycarbonylpentamidocarbonyl)-9-chloroacridine

Methyl 6-aminoheptane hydrochloride (4.7 g, 25.9 mmol) was dissolved in methylene chloride (90 mL), cooled to 0° C, triethyl amine (15 mL) was added, and the resulting solution was then immediately added to the acid chloride from Example 12. The solution was again stirred vigorously for 30 minutes at 0° C and 1 h at room temperature. The resulting mixture was filtered to remove the remaining solids, which were washed with methylene chloride (20 mL). The reddish-brown methylene chloride filtrate was subsequently washed twice with saturated sodium hydrogen carbonate, once with saturated sodium chloride, dried over magnesium sulfate, and evaporated to dryness, in vacuo. To the resulting oily residue was added dry benzene (35 mL) and ligroin (60°-80° C, dried over Na-Pb). The mixture was heated to reflux. Activated carbon and celite were added and the mixture refluxed for 3 minutes. After filtration, the title compound crystallized upon cooling with magnetic stirring. It was isolated by filtration and washed with petroleum ether. The product was stored over solid potassium hydroxide. Yield 5 g (50%).

EXEMPLARY 14

Synthesis of 4-(5-methoxycarbonylpentamidocarbonyl) amidocarbonyl-9-{(4N-nitrobenzamido)-hexylaminol-aminocaridine

4-(5-Methoxy carbonylpentamidocarbonyl)-9-chloroacridine (1.3 g, 3.38 mmol) and phenol (5 g) were heated to 80° C for 30 minutes under a flow of nitrogen after which 6-(4N-nitrobenzamido)-1-heptylamine (897 mg, 3.38 mmol) was added. The temperature was then increased to 120° C for 2 h. The mixture was cooled and ethynylchloride (80 mL) was added. The resulting solution was washed three times with 2N sodium hydroxide (60 mL, portions) and once with water, dried over magnesium sulfate, and evaporated to dryness, in vacuo. The resulting red oil (1.8 g) was dissolved in methylene chloride (40 mL) and cooled to 0° C. Ether (120 mL) was added and the resulting solution was stirred overnight. This resulted in a mixture of solid material and an oil. The solid was isolated by filtration. The solid and the oil were re-dissolved in methylene chloride (80 mL) and added dropwise to cold ether (150 mL). After 20 minutes of stirring, the title compound was isolated by filtration as orange crystals. The product was washed with ether and dried in vacuo over potassium hydroxide. Yield 1.6 g (77%). M.p. 145°-147° C.

EXEMPLARY 15

Synthesis of 4-(5-carboxypentyl) amidocarbonyl-9-{(4N-nitrobenzamido)-hexylaminol-aminocaridine

4-(5-Methoxy carbonylpentamidocarbonyl)-9-{(4N-nitrobenzamido)hexylaminolaminocaridine (503 mg, 0.82 mmol) was dissolved in DMF (30 mL), and 2N sodium hydroxide (30 mL) was added. After stirring for 15 minutes, 2N hydrochloric acid (35 mL) and water (50 mL) were added at 0° C. After stirring for 30 minutes, the solution was decanted, leaving an oily substance which was dissolved in
boiling methanol (150 mL). Filtered and concentrated to a third of the volume. To the methanol solution were added ether (125 mL) and 5–6 drops of HCl in ethanol. The solution was decanted after 1 h of stirring at 0°C. The oily substance was redissolved in methanol (25 mL) and precipitated with ether (150 mL). The title compound was isolated as yellow crystals after stirring overnight. Yield: 417 mg (80%). M.p. 173°C (decomp.).

**EXAMPLE 16**

(a) Synthesis of 4-((5-pentafluorophenylxoycarbonyl-pentyl)-amidocarbonyl-9-{6'-4-nitrobenzamide}hexyl-amino) aminoacridine(Ac^tOPpp).

The acid from above, Example 15, (300 mg, 0.48 mmol) was dissolved in DMF (2 mL) and methylene chloride (8 mL) was added. Pentafluorophenol (97 mg, 0.53 mmol) was transferred with 2×2 mL of the methylene chloride solution, was added. The resulting solution was cooled to 0°C after which DCC (124 mg, 0.6 mmol) was subsequently added. The ice bath was removed after 5 minutes and the mixture was stirred overnight. The precipitated DCU was removed by centrifugation and the centrifuge was evaporated to dryness, in vacuo, first by a water aspirator and then by an oil pump. The residue was dissolved in methylene chloride (20 mL), filtered, and evaporated to dryness, in vacuo. The residue was again dissolved in methylene chloride and petroleum ether (150 mL). A 1 mL aliquot of 5M HCl in ether was added. The solvent was removed by decanting after 30 minutes of stirring at 0°C. The residual oily substance was dissolved in methylene chloride (100 mL). Petroleum ether (150 mL) was added and the mixture was stirred overnight. The yellow precipitated crystalline material was isolated by filtration and washed with copious amounts of petroleum ether. Yield (after drying): 300 mg (78%). M.p. 97.5°C (decomp.). All samples showed satisfactory elemental analysis. 1H- and 13C-NMR and mass spectra.

(b) Experimental Procedure for the Synthesis of PNAs

**Materials:** BOC-Lys (CIZ), benzhydrylamine-copoly(styrene-1%-divinylbenzene) resin (BHA resin), and p-methylbenzhydrylamine-copoly(styrene-1%-divinylbenzene) resin (MBHA resin) were purchased from Peninsula Laboratories. Other reagents and solvents were: Biograde trifluoroacetic acid from Halocarbon Products; diisopropylethylamine (99%: was not further distilled) and N-acetylimidazole (98%) from Aldrich; H2O was distilled twice; anhydrous HF from Union Carbide; synthesis grade N,N-dimethylformamide and analytical grade methylene chloride (was not further distilled) from Merck; HPLC grade acetonitrile from Lab-Scan; purum grade anisole, N,N-dicyclohexylcarbodiimide, and puriss. grade 2.2.2-trifluoroethanol from Fluka.

**General Methods and Remarks**

Except where otherwise stated, the following applies. The PNA compounds were synthesized by the stepwise solid phase approach (Merrifield, *J. Am. Chem. Soc.*, 1963, 85, 2149) employing conventional peptide chemistry utilizing the TFA-labile tert-butyloxycarbonyl (BOC) group for "temporary" N-protection (Merrifield, *J. Am. Chem. Soc.*, 1964, 86, 304) and the more acid-stable benzylxocarbonyl (Z) and 2-chlorobenzoxycarbonyl (Clz) groups for "permanent" side chain protection. To obtain C-terminal amides, the PNAs were assembled onto the HF-labile BHA or MBHA resin (the MBHA resin has increased susceptibility to the final HF cleavage relative to the unsubstituted BHA resin (Matsueda et al., *Peptides*, 1981, 2, 45). All reactions (except HF reactions) were carried out in manually operated standard solid phase reaction vessels fitted with a coarse glass frit (Merrifield et al., *Biochemistry*, 1982, 21, 5020). The quantitative ninhydrin reaction (Kaiser test), originally developed by Sarin et al. (*Anal. Biochem.*, 1981, 117, 147) for peptides containing "normal" amino acids, was successfully applied (see Table I–III) using the "normally" employed effective extinction coefficient ε=15000 M⁻¹cm⁻¹ for all residues to determine the completeness of the individual couplings as well as to measure the number of growing peptide chains. The theoretical substitution S_n upon coupling of residue number n (assuming both complete deprotection and coupling as well as neither chain termination nor loss of PNA chains during the synthetic cycle) is calculated from the equation:

\[ S_n = \frac{1}{1 + (\Delta MW \times 10^{-2} \text{ mmol/g})} \]

where ΔMW is the gain in molecular weight (\(\Delta MW = g/g\)) and S_n is the theoretical substitution upon coupling of the preceding residue n⁻¹ (\(S_n = \text{mmol/g}\)). The estimated value (%) on the extent of an individual coupling is calculated relative to the measured substitution (unless S was not determined) and include correction for the number of remaining free amino groups following the previous cycle.

**EXAMPLE 17**

**Solid Phase Synthesis of Ac⁻¹-[Taeg]₁₅–NH₃ and Shorter Derivatives**

(a) Stepwise Assembly of BOC-[Taeg]₁₅–BHA Resin.

The synthesis was initiated on 100 mg of preswollen and neutralized BHA resin (determined by the ninhydrin reaction to contain 0.57 mmol NH₃/g) employing single couplings ("Synthetic Protocol 1") using 3.2 equivalents of BOC-Taeg-OPpp in about 33% DMF/CH₂Cl₂. The individual coupling reactions were carried out by shaking for at least 12 h in a manually operated 6 mL standard solid phase reaction vessel and unreacted amino groups were blocked by acetylation at selected stages of the synthesis. The progress of chain elongation was monitored at several stages by the quantitative ninhydrin reaction (see Table I). Portions of protected BOC-[Taeg]₁₅–BHA, BOC-[Taeg]₁₅–BHA, and BOC-[Taeg]₁₅–BHA resins were taken out after assembling 5, 10, and 15 residues, respectively.
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<th>Theor. (mmol/g)</th>
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ND = Not Determined

(b) Synthesis of Ac-1-[Taeg]_{15}-BHA Resin

Following deprotection of the residual BOC-[Taeg]_{15}-BHA resin (estimated dry weight is about 30 mg, ~0.002 mmol growing chains), the H-[Taeg]_{15}-BHA resin was reacted with about 50 equivalents (80 mg, 0.11 mmol) of Ac-1-OPfp in 1 mL of about 66% DMAP/CH_2Cl_2 (i.e., a 0.11M solution of the pentafluorophenylester) in a 3 mL solid phase reaction vessel. As judged by a qualitative ninhydrin reaction, coupling of the acridine moiety was close to quantitative.

(c) Cleavage, Purification, and Identification of H-[Taeg]_{15}-NH_2

A portion of protected BOC-[Taeg]_{15}-BHA resin was treated with 50% trifluoroacetic acid in methylene chloride to remove the N-terminal BOC group (which is a precursor of the potentially harmful tert-butyl cation) prior to the HF cleavage. Following neutralization and washing (performed in a way similar to those of steps 2-4 in "Synthetic Protocol 1"), and drying for 2 h in vacuum, the resulting 67.1 mg (dry weight) of H-[Taeg]_{15}-BHA resin was cleaved with 5 mL of HF/anisole (9:1, v/v) stirring at 0°C for 60 minutes. After removal of HF, the residue was stirred with dry diethyl ether (4×15 mL; 15 minutes each) to remove anisole, filtered under gravity through a fluted glass funnel, and dried. The PNA was then extracted into a 60 mL (4×15 mL, stirring 15 minutes each) 10% aqueous acetic acid solution. Aliquots of this solution were analyzed by analytical reverse-phase HPLC to establish the purity of the crude PNA. The main peak at 13 minutes accounted for about 93% of the total absorbance. The remaining solution was frozen and lyophilized to afford about 22.9 mg of crude material. Finally, 19 mg of the crude product was purified by five batches, each containing 3.8 mg in 1 mL of H_2O. The main peak was collected by use of a semi-preparative reverse-phase column. Acetonitrile was removed on a speed vac and the residual solution was frozen (dry ice) and subsequently lyophilized to give 13.1 mg of >99% pure H-[Taeg]_{15}-NH_2. The PNA molecule readily dissolved in water and had the correct molecular weight based on mass spectral determination. For (M+H)^+ the calculated m/z value was 1349.3 and the measured m/z value was 1347.8.

(d) Cleavage, Purification, and Identification of H-[Taeg]_{10}-NH_2

A portion of protected BOC-[Taeg]_{10}-BHA resin was treated as described in section (c) to yield 11 mg of crude material upon HF cleavage of 18.9 mg dry H-[Taeg]_{10}-BHA resin. The main peak at 15.5 minutes accounted for about 53% of the total absorbance. About 1 mg of the crude product was purified repeatedly (for reasons described below) to give approximately 0.1 mg of at least 80% but presumably >99% pure H-[Taeg]_{10}-NH_2. A rather broad tail eluting after the target peak and accounting for about 20% of the total absorbance could not be removed (only slightly reduced) upon the repeated purification. Judged by the mass spectrum, which only confirms the presence of the correct molecular weight H-[Taeg]_{10}-NH_2, the tail phenomenon is ascribed to more or less well-defined aggregational/conformational states of the target molecule. Therefore, the crude product is likely to contain more than the above-mentioned 53% of the target molecule. H-[Taeg]_{10}-NH_2 is readily dissolved in water. For (M+H)^+ the calculated m/z value was 2679.6 and the measured m/z value was 2681.5.

(e) Cleavage, Purification, and Identification of H-[Taeg]_{15}-NH_2

A portion of protected BOC-[Taeg]_{15}-BHA resin was treated as described in section (c) to yield 3.2 mg of crude material upon HF cleavage of 13.9 mg dry H-[Taeg]_{15}-BHA resin. The main peak at 22.6 minutes was located in a broad bulge accounting for about 60% of the total absorbance (FIG. 12a). Again (see the preceding section), this bulge is ascribed to aggregational/conformational states of the target molecule H-[Taeg]_{15}-NH_2 since mass spectral analysis of the collected "bulge" did not significantly reveal the presence of other molecules. All of the crude product was purified collecting the "bulge" to give approximately 2.8 mg material. For (M+Na)^+ the calculated m/z value was 4033.9 and the measured m/z value was 4032.9.

(f) Cleavage, Purification, and Identification of Ac-1-[Taeg]_{15}-NH_2

A portion of protected Ac-1-[Taeg]_{15}-BHA resin was treated as described in section (b) to yield 14.3 mg of crude material upon HF cleavage of 29.7 mg dry Ac-1-[Taeg]_{15}-
BHA resin. Taken together, the main peak at 23.7 minutes and a “dimer” (see below) at 29.2 minutes accounted for about 40% of the total absorbance (Fig. 12b). The crude product was purified repeatedly by give approximately 1 mg of presumably >99% pure AcTr-[Tae]$_{15}$-NH$_2$ “contaminated” with self-aggregated molecules eluting at 27.4 minutes, 29.2 minutes, and finally as a huge broad bulge eluting with 100% buffer B (Fig. 12c). This interpretation is in agreement with the observation that some peaks grow upon standing (for hours) in aqueous acidic acid solution, and finally precipitate out quantitatively. For (M+H)$^+$ the calculated m/z value was 4593.6 and the measured m/z value was 4588.7.

(g) Synthetic Protocol 1

(1) BOC-deprotection with TFA/CH$_2$Cl$_2$ (1:1, v/v), 3 mL, 3×1 minute and 1×30 minutes; (2) washing with CH$_2$Cl$_2$, 3 mL, 6×1 minute; (3) neutralization with DIEA/CH$_2$Cl$_2$ (1:19, v/v), 3 mL, 3×2 minutes; (4) washing with CH$_2$Cl$_2$, 3 mL, 6×1 minute, and drain for 1 minute; (5) 2–5 mg sample of PNA resin may be removed and dried for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 3.2 equiv. (0.18 mmol, 100 mg) BocTae-GOPp dissolved in 1 mL of CH$_2$Cl$_2$ followed by addition of 0.5 mL of DMF (final concentration of pentfluorophenylether -0.12M); the coupling reaction was allowed to proceed for a total of 12–24 h shaking at room temperature; (7) washing with DMF, 3 L, 1×2 minutes; (8) washing with CH$_2$Cl$_2$, 3 mL, 4×1 minute; (9) neutralization with DIEA/CH$_2$Cl$_2$ (1:19, v/v), 3 mL, 3×2 minutes; (10) washing with CH$_2$Cl$_2$, 3 mL, 6×1 minute; (11) 2–5 mg sample of protected PNA resin is taken out for a rapid qualitative ninhydrin test and further 2–5 mg is dried for a quantitative ninhydrin analysis to determine the extent of coupling (after cycles 7, 10, and 15 reacted amino groups were blocked by acetylation with N-acetylimidazole in methylene chloride).

EXAMPLE 18

Solid Phase Synthesis of AcTr-[Tae]$_{15}$-Lys-NH$_2$ and Shorter Derivatives

(a) Stepwise Assembly of BOC-[Tae]$_{15}$-Lys(CIZ)-BHA Resin

The synthesis was initiated by a quantitative loading (standard DCC in situ coupling in neat CH$_2$Cl$_2$) of BOC-Lys(CIZ) onto 100 mg of preswollen and neutralized BHA resin (0.57 mmol NH$_2$/g). Further extension of the protected PNA chain employed simple couplings (“Synthetic Protocol 2”) for cycles 1 to 5 and 5 to 10 using 3.2 equivalents of BocTae-GOPp in about 33% DMF/CH$_2$Cl$_2$. Cycles 5 to 10 employed an additional DCC (i.e., in situ) coupling of the free acid BocTae-OH in about 33% DMF/CH$_2$Cl$_2$. All coupling reactions were carried out by shaking for at least 12 h in a manually operated 6 mL standard solid phase reaction vessel. Unreacted amino groups were blocked by acetylation at the same stages of the synthesis as was done in Example 17. Portions of protected BOC-[Tae]$_{15}$-Lys(CIZ)-BHA and BOC-[Tae]$_{15}$-Lys(CIZ)-BHA resins were removed after assembling 5 and 10 PNA residues, respectively. As judged by the analytical HPLC chromatogram of the crude cleavage product from the BOC-[Tae]$_{15}$-Lys(CIZ)-BHA resin (see section (e)), an additional “free acid” coupling of PNA residues 5 to 10 gave no significant improvement of the synthetic yield as compared to the throughout single-coupled residues in Example 17.

(b) Synthesis of AcTr-[Tae]$_{15}$-Lys(CIZ)-BHA Resin

Following deprotection of a portion of BOC-[Tae]$_{15}$-Lys(CIZ)-BHA resin (estimated dry weight is about 90 mg, 0.01 mmol growing chains), the H-[Tae]$_{15}$-BHA resin was reacted with about 20 equivalents (141 mg, 0.19 mmol) of AcTr-[Tae]$_{15}$-OPp in 1 mL of about 66% DMF/CH$_2$Cl$_2$ in a 3 mL solid phase reaction vessel. As judged by a qualitative ninhydrin reaction, coupling of the acridine moiety was close to quantitative.

(c) Synthesis of AcTr-[Tae]$_{15}$-Lys(CIZ)-BHA Resin

Following deprotection of the residual BOC-[Tae]$_{15}$-Lys(CIZ)-BHA resin (estimated dry weight about 70 mg, 0.005 mmol growing chains), the H-[Tae]$_{15}$-Lys(CIZ)-BHA resin was reacted with about 25 equivalents (91 mg, 0.12 mmol) of AcTr-[Tae]$_{15}$-OPp in 1 mL of about 66% DMF/CH$_2$Cl$_2$ in a 3 mL solid phase reaction vessel. As judged by a qualitative ninhydrin reaction, coupling of the acridine moiety was close to quantitative.

(d) Cleavage, Purification, and Identification of H-[Tae]$_{15}$-Lys-NH$_2$

A portion of protected BOC-[Tae]$_{15}$-Lys(CIZ)-BHA resin was treated as described in Example 17(c) to yield 8.9 mg of crude material upon UF cleavage of 19 mg dry H-[Tae]$_{15}$-Lys(CIZ)-BHA resin. The main peak at 12.2 minutes (eluted at 14.2 minutes if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) accounted for about 90% of the total absorbance. About 2.2 mg of the crude product was purified to give approximately 1.5 mg of 99% pure H-[Tae]$_{15}$-Lys-NH$_2$.

(e) Cleavage, Purification, and Identification of H-[Tae]$_{15}$-Lys(CIZ)-BHA Resin

A portion of protected BOC-[Tae]$_{15}$-Lys(CIZ)-BHA resin was treated as described in Example 17(c) to yield 1.7 mg of crude material upon UF cleavage of 7 mg dry H-[Tae]$_{15}$-Lys-CIZ)-BHA resin. The main peak at 15.1 minutes (eluted at 17 minutes if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) accounted for about 50% of the total absorbance. About 1.2 mg of the crude product was purified to give approximately 0.2 mg of >95% pure H-[Tae]$_{15}$-Lys-NH$_2$ (Fig. 4). For (M+H)$^+$ the calculated m/z value was 2807.8 and the measured m/z value was 2808.2.

(f) Cleavage, Purification, and Identification of AcTr-[Tae]$_{15}$-Lys-NH$_2$

Protected AcTr-[Tae]$_{15}$-Lys(CIZ)-BHA resin (99.1 mg, dry weight) was cleaved as described in Example 17(c) to yield 42.2 mg of crude material. The main peak at 23.5 minutes (eluted at 23.5 minutes if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) accounted for about 45% of the total absorbance. An 8.87 mg portion of the crude product was purified to give approximately 5.3 mg of >97% pure AcTr-[Tae]$_{15}$-Lys-NH$_2$. For (M+H)$^+$ the calculated m/z value was 2850.8 and the measured m/z value was 2849.8.

(g) Cleavage and Purification of AcTr-[Tae]$_{15}$-Lys-NH$_2$

A 78.7 mg portion of protected AcTr-[Tae]$_{15}$-Lys(CIZ)-BHA resin (dry weight) was cleaved as described in Example 18 to yield 34.8 mg of crude material. The main peak at 23.5 minutes (about the same elution time if injected from an aqueous solution instead of the 10% aqueous acetic acid solution) and a “dimer” at 28.2 minutes accounted for about 35% of the total absorbance. About 4.5 mg of the crude product was purified to give approximately 1.6 mg of presumably >95% pure AcTr-[Tae]$_{15}$-Lys-NH$_2$. This compound could not be free of the “dimer” peak, which grew upon standing in aqueous acetic acid solution.

(h) Synthetic Protocol 2

(1) BOC-deprotection with TFA/CH$_2$Cl$_2$ (1:1, v/v), 3 mL, 3×1 minute and 1×30 minutes; (2) washing with CH$_2$Cl$_2$, 3
mL, 6×1 minute; (3) neutralization with DIAE/CH₂Cl₂ (1: 19, v/v), 3 mL, 3×2 minutes; (4) washing with CH₂Cl₂, 3 mL, 6×1 minute, and drain for 1 minute; (5) 2–5 mg sample of PNA-resin can be removed and dried for a qualitative ninhydrin analysis; (6) for cycles 1 to 5 and cycles 10 to 15 the coupling reaction was carried out by addition of 3.2 equiv. (0.18 mmol, 100 mg) of BOC-Taqg-OPf dissolved in 1 mL of CH₂Cl₂ followed by addition of 0.5 mL of DMF (final concentration of pentafluorophenylester -0.12M). The coupling reaction was allowed to proceed for a total of 12–24 h with shaking; cycles 5 to 10 employed an additional 0.12M DCC coupling of 0.12M BOC-Taqg-OH in 1.5 mL of DMF/CH₂Cl₂ (1:2, v/v); (7) washing with DMF, 3 mL, 1×2 minutes; (8) washing with CH₂Cl₂, 3 mL, 4×1 minute; (9) neutralization with DIAE/CH₂Cl₂ (1:19, v/v), 3 mL, 2×2 minutes; (10) washing with H₂O, 3 mL, 6×1 minute; (11) 2–5 mg sample of protected PNA-resin is removed for a qualitative ninhydrin test (after cycles 7, 10, and 15), and unreacted amino groups were blocked by acetylation with N-acetylglucosamide in methylene chloride).

EXAMPLE 19
Improved Solid Phase Synthesis of H-[Taqg]₁₀-Lys-NH₂

The protected PNA was assembled onto an MBHA resin, using approximately half the loading of the BHA resin used in the previous examples. Furthermore, all cycles except one was followed by acetylation of uncoupled amino groups. The following describes the synthesis in detail:
(a) Preparation of BOC-Lys(CIZ)-NH-CH₉(C₆H₇-C₆H₇-C₆H₇)-C₆H₅ Resin (MBHA Resin) With 0.3 mmol/g

The desired substitution of BOC-Lys(CIZ)-MBHA resin was 0.25–0.30 mmol/g. In order to get this value, 1.5 mmol of BOC-Lys(CIZ) was coupled to 5 g of neutralized and preswollen MBHA resin (determined by quantitative ninhydrin reaction to contain 0.64 mmol NH₂/g) using a single “in situ” coupling (1.5 mmol of DCC) in 60 mL of CH₂Cl₂. The

0.32 mmol/g for the neutralized H-Lys(CIZ)-MBHA resin. This compares well with the maximum value of 0.28 mmol/g for a quantitative coupling of 0.30 mmol BOC-Lys (CIZ)g resin (see Table II).

(b) Stepwise Assembly of BOC-[Taqg]₁₀-Lys(CIZ)-MBHA Resin

The entire batch of H-Lys(CIZ)-MBHA resin prepared in section (a) was used directly (in the same reaction vessel) to assemble BOC-[Taqg]₁₀-Lys(CIZ)-MBHA resin by single couplings ("Synthetic Protocol 3") utilizing 2.5 equivalents of BOC-Taqg-OPf in neat CH₂Cl₂. The quantitative ninhydrin reaction was applied throughout the synthesis (see Table II).

(c) Stepwise Assembly of BOC-[Taqg]₁₀-Lys(CIZ)-MBHA Resin

About 4.5 g of wet BOC-[Taqg]₁₀-Lys(CIZ)-MBHA resin (~0.36 mmol growing chains, taken out of totally ~19 g wet resin prepared in section (b)) was placed in a 55 mL solid phase peptide synthesis (SPPS) reaction vessel. BOC-[Taqg]₁₀-Lys(CIZ)-MBHA resin was assembled by single couplings ("Synthetic Protocol 4") utilizing 2.5 equivalents of BOC-Taqg-OPf in about 30% DMF/CH₂Cl₂. The progress of the synthesis was monitored at all stages by the quantitative ninhydrin reaction (see Table II).

(d) Stepwise Assembly of BOC-[Taqg]₁₀-Lys(CIZ)-MBHA Resin

About 1 g of wet BOC-[Taqg]₁₀-Lys(CIZ)-MBHA resin (~0.09 mmol growing chains, taken out of totally ~4 g wet resin prepared in section (c)) was placed in a 20 mL SPPS reaction vessel. BOC-[Taqg]₁₀-Lys(CIZ)-MBHA resin was assembled by the single-coupling protocol employed in the preceding section utilizing 2.5 equivalents of BOC-Taqg-OPf in about 30% DMF/CH₂Cl₂. The reaction volume was 3 mL (vigorous shaking). The synthesis was monitored by the quantitative ninhydrin reaction (see Table II).

<table>
<thead>
<tr>
<th>Synthetic Residue</th>
<th>Remaining Free Amino Groups After Deprotection (mmol/g)</th>
<th>Single Coupling (μmol/g)</th>
<th>Estimation Extent (%)</th>
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<td>&quot;O&quot;</td>
<td>BOC-Lys(CIZ)</td>
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<tr>
<td>1</td>
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<tr>
<td>10</td>
<td>BOC-Taqg</td>
<td>0.11</td>
<td>0.16</td>
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</table>

(e) Synthesis of Ac-[Taqg]₁₀-Lys(CIZ)-MBHA Resin

Following deprotection of a portion of Boc-[Taqg]₁₀-Lys (CIZ)-MBHA resin (estimated dry weight is about 45 mg), the residue was next acylated quantitatively with a 2 mL mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v/v) for 2 h in a 3 mL solid phase reaction vessel.
29

(f) Cleavage, Purification, and Identification of H-[Tagel]_10-Lys-NH₂

A portion of protected Boc-[Tagel]_10-Lys(CZ)-BHA resin was treated as described in Example 17(c) to yield about 24 mg of crude material upon HF cleavage of 76 mg dry H-[Tagel]_10-Lys(CZ)-BHA resin. The main peak at 15.2 minutes (which includes impurities such as deletion peptides and various byproducts) accounted for about 78% of the total absorbance. The main peak also accounted for about 88% of the "main peak plus deletion peaks" absorbance, which is in good agreement with the overall estimated coupling yield of 90.1% obtained by summarizing the individual coupling yields in Table II. A 7.2 mg portion of the crude product was purified from two batches by use of a semi-preparative reverse-phase column, collecting the main peak in a beaker cooled with dry ice/2-propanol. Each contained 3.6 mg in 1 mL of H₂O. The frozen solution was lyophilized directly (without prior removal of acetonitrile on a speed vac) to give 4.2 mg of 82% pure H-[Tagel]_10-Lys-NH₂.

(g) Cleavage, Purification, and Identification of Ac-[Tagel]_10-Lys-NH₂

A 400.0 mg portion of protected Ac-[Tagel]_10-Lys(CZ)-BHA resin (dry weight) was cleaved as described in Example 17(c), except for the TFA treatment to yield 11.9 mg of crude material. The main peak at 15.8 minutes accounted for about 75% of the total absorbance. A 4.8 mg portion of the crude product was purified to give approximately 3.5 mg of >95% pure Ac-[Tagel]_10-Lys-NH₂. For (M+H)⁺ the calculated m/z value = 2849.8 and the measured m/z value = 2848.8.

(h) Synthetic Protocol 3

(1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 100 mL, 3×1 minute and 1×30 minutes; (2) washing with CH₂Cl₂, 100 mL, 6×1 minute; (3) neutralization with DIEA/CH₂Cl₂ (1:19, v/v), 100 mL, 3×2 minutes; (4) washing with CH₂Cl₂, 100 mL, 6×1 minute, and drain for 1 minute; (5) 2–5 mg sample of PNA-resin is removed and dried for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 2.5 equiv. (3.75 mmol, 2.064 g) Boc-Tg-OPfp dissolved in 35 mL CH₂Cl₂ (final concentration of pentafluorophenylester -0.1M); the coupling reaction was allowed to proceed for a total of 20-24 h with shaking; (7) washing with DMF, 100 mL, 1×2 minutes (to remove precipitate of BOC-Tagel-OH); (8) washing with CH₂Cl₂, 100 mL, 4×1 minute; (9) neutralization with DIEA/CH₂Cl₂ (1:19, v/v), 100 mL, 2×2 minute; (10) washing with CH₂Cl₂, 100 mL, 6×1 minute; (11) 2–5 mg sample of protected PNA-resin was removed for a rapid qualitative ninhydrin test and a further 2–5 mg is dried for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 100 mL mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v/v) for 2 h; (13) washing with CH₂Cl₂, 100 mL, 6×1 minute; (14) 2×2–5 mg samples of protected PNA-resin were removed, neutralized with DIEA/CH₂Cl₂ (1:19, v/v), and washed with CH₂Cl₂ for qualitative and quantitative ninhydrin analyses.

(i) Synthetic Protocol 4

(1) Boc-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 25 mL, 3×1 min and 1×30 minutes; (2) washing with CH₂Cl₂, 25 mL, 6×1 minute; (3) neutralization with DIEA/CH₂Cl₂ (1:19, v/v), 25 mL, 3×2 minutes; (4) washing with CH₂Cl₂, 25 mL, 6×1 minute, and drain for 1 minute; (5) 2–5 mg sample of PNA-resin was removed and dried for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 2.5 equiv. (0.92 mmol, 0.506 g) Boc-Tg-OPfp dissolved in 6 mL CH₂Cl₂ followed by addition of 3 mL DMF (final concentration of pentafluorophenylester -0.1M); the coupling reaction was allowed to proceed for a total of 20–24 hrs with shaking; (7) washing with DMF, 25 mL, 1×2 minutes; (8) washing with CH₂Cl₂, 25 mL, 4×1 minute; (9) neutralization with DIEA/CH₂Cl₂ (1:19, v/v), 25 mL, 2×2 minutes; (10) washing with CH₂Cl₂, 25 mL, 6×1 minute; (11) 2–5 mg sample of protected PNA-resin was removed for a rapid qualitative ninhydrin test and a further 2–5 mg is dried for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 25 mL mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:12, v/v/v) for 2 h (except after the first cycle); (13) washing with CH₂Cl₂, 25 mL, 6×1 minute; (14) 2×2–5 mg samples of protected PNA-resin are taken out, neutralized with DIEA/CH₂Cl₂ (1:19, v/v), and washed with CH₂Cl₂ for qualitative and quantitative ninhydrin analyses.

EXAMPLE 20

Synthesis of N-benzylxycarvoxyl-N'-BOC-aminomethyl glycine

Aminoethyl glycine (52.86 g, 0.447 mol) was dissolved in water (900 mL) and dioxane (900 mL) was added. The pH was adjusted to 11.2 with 2 N NaOH. While the pH was kept at 11.2, tert-butyl-p-nitrophenyl carbonate (128.4 g, 0.537 mol) was dissolved in dioxane (720 mL) and added dropwise over the course of 2 hours. The pH was kept at 11.2 for at least three more hours and then allowed to stand overnight, with stirring. The yellow solution was cooled to 0°C and the pH was adjusted to 3.5 with 2 N HCl. The mixture was washed with chloroform (4×100 mL), and the pH of the aqueous phase was readjusted to 9.5 with 2 N NaOH at 0°C. Benzoylcarboxyl chloride (73.5 mL, 0.515 mol) was added over half an hour, while the pH was kept at 9.5 with 2 N NaOH. The pH was adjusted frequently over the next 4 hours, and the solution was allowed to stand overnight, with stirring. On the following day the solution was washed with ether (3×600 mL), and the pH of the solution was afterwards adjusted to 1.5 with 2 N HCl at 0°C. The title compound was isolated by extraction with ethyl acetate (5×1000 mL). The ethyl acetate solution was dried over magnesium sulfate and evaporated to dryness, in vacuo. This afforded 138 g of the product, which was dissolved in ether (300 mL) and precipitated with the addition of ether (1800 mL). Yield 1247 g (79%), M.p. 64.5°-65°C. Anal. for C₁₀H₁₈N₂O₄ found(calcd.) C: 58.40(57.94); H: 7.02(6.86); N: 7.94(7.95). 1H-NMR (250 MHz, CDCl₃) δ 7.32 & 7.32 (2H, Ph); 5.15 & 5.12 (2H, PhCH); 4.03 & 4.01 (2H, NCH₂CO₂H); 3.46 (b, 2H, BOC-NHCH₂CH₃); 3.28 (b, 2H, BOC-NHCH₂CH₃); 1.43 & 1.40 (9H, t-Bu). HPLC (260 nm) 20.71 min (80.2%) and 21.57 min (19.8%). The UV-spectra (200 nm-300 nm) are identical, indicating that the minor peak consists of Bis-Z-AEP.

EXAMPLE 21

Synthesis of N'-BOC-aminomethylglucose ethyl ester

N-Benzoylcarboxylic acid-N'-BOC-aminomethylglycine (60 g, 0.170 mol) and N,N-dimethyl-4-aminopyridine (6 g) were dissolved in absolute ethanol (500 mL), and cooled to 0°C before the addition of DCC (42.2 g, 0.204 mol). The ice bath was removed after 5 minutes and stirring was continued for 2 more hours. The precipitated DCC (32.5 g, dried) was removed by filtration and washed with ether (3×100 mL). The combined filtrate was washed successively with diluted potassium hydroxide solution (2×400 mL), diluted sodium carbonate (2×1000 mL), and saturated aqueous sodium chloride (2×1000 mL). The ethyl ester was isolated by chromatography on silica gel (50% EtOAC in hexane). Yield 51.8 g (76%), M.p. 59-60°C. Anal. for C₂₄H₃₃NO₂ found(calcd.) C: 74.57(74.44); H: 7.64(7.58). 1H-NMR (250 MHz, CDCl₃) δ 7.52-7.45 & 7.45-7.36 (2H, Ph); 4.33 & 4.31 (2H, NCH₂CO₂H); 3.48 (b, 2H, BOC-NHCH₂CH₃); 3.28 (b, 2H, BOC-NHCH₂CH₃); 1.43 & 1.40 (9H, t-Bu). HPLC (260 nm) 20.21 min (80.7%) and 21.53 min (19.3%). The UV-spectra (200 nm-300 nm) are identical, indicating that the minor peak consists of Bis-Z-AEP.
hydrogen carbonate (2'400 mL) and saturated sodium chloride (1 x 400 mL). The organic phase was filtered, then dried over magnesium sulfate, and evaporated to dryness, in vacuo, which yielded 66.1 g of an oily substance which contained some DCU.

The oil was dissolved in absolute ethanol (600 mL) and was added 10% palladium on carbon (6.6 g) was added. The solution was hydrogenated at atmospheric pressure, where the reservoir was filled with 2N sodium hydroxide. After 4 hours, 3.3 L was consumed out of the theoretical 4.2 L. The reaction mixture was filtered through celite and evaporated to dryness, in vacuo. Afoading 39.5 g (94%) of an oily substance. A 13 g portion of the oily substance was purified by silica gel (SiO2, 600 g) chromatography. After elution with 300 mL of 20% petroleum ether in methylene chloride, the title compound was eluted with 1700 mL of 5% methanol in methylene chloride. The solvent was removed from the fractions with satisfactory purity, in vacuo and the yield was 8.49 g. Alternatively 10 g of the crude material was purified by Kugel Rohr distillation. 

H-NMR (250 MHz, CD3OD): 4.77 (s, 4H, CH2CD3); 3.38 (s, 2H, NCH2CO2Et); 3.16 (t, 2H, NOH-CH2-CH3); 2.08 (t, 2H, NOH-CH2-CH3); 1.43 (s, 9H, t-Bu) and 1.26 (t, 3H, CH3). D3-NMR: 171.4 (COE); 155.6 (CO); 78.3 (CH3); 59.9 (CH3); 49.0 (CH3); 48.1 (CH3); 39.0 (CH3); 26.9 (CH3) and 12.6 (CH3).

**EXAMPLE 22**

Synthesis of N'-BOC-aminooxyglycine methyl ester

The above procedure, Example 21, was used, with methanol being substituted for ethanol. The final product was purified by column purification.

**EXAMPLE 23**

Synthesis of 1-(BOC-aeg)thyme ethyl ester

N'-BOC-aminooxyglycine ethyl ester (13.5 g, 54.8 mmol), DiBOH (9.84 g, 60.3 mmol) and 1-carboxymethyl thymine (11.1 g, 60.3 mmol) were dissolved in DMF (210 mL). Methylene chloride (210 mL) was added. The solution was cooled to 0°C in an ethanol/ice bath and DCC (13.6 g, 65.8 mmol) was added. The ice bath was removed after 1 hour and stirring was continued for another 2 hours at ambient temperature. The precipitated DUC was removed by filtration and washed twice with methylene chloride (2x75 mL). To the combined filtrate was added more methylene chloride (650 mL). The solution was washed successively with diluted sodium hydrogen carbonate (3x500 mL), diluted potassium hydrogen sulfinate (2x500 mL) and saturated sodium chloride (1x500 mL). Some of the precipitate was removed from the organic phase by filtration. The organic phase was dried over magnesium sulfate and evaporated to dryness, in vacuo. The oily residue was dissolved in methylene chloride (150 mL), filtered, and the title compound was precipitated by the addition of petroleum ether (350 mL) at 0°C. The methylene chloride/petroleum ether procedure was repeated once. This afforded 16 g (71%) of a material which was more than 99% pure by HPLC.

**EXAMPLE 24**

Synthesis of 1-(BOC-aeg)thyme

The material from above was suspended in THF (194 mL, gives a 0.2M solution), and 1M aqueous lithium hydroxide (116 mL) was added. The mixture was stirred for 45 minutes at ambient temperature and then filtered to remove residual DUC. Water (40 mL) was added to the solution which was then washed with methylene chloride (300 mL). Additional water (30 mL) was added, and the alkaline solution was washed once more with methylene chloride (150 mL). The aqueous solution was cooled to 0°C and the pH was adjusted to 2 by the dropwise addition of 1N HCl (approx. 110 mL). The title compound was extracted with ethyl acetate (9 x 200 mL). The combined extracts were dried over magnesium sulfate and were evaporated to dryness, in vacuo. The residue was evaporated once from methanol.

Which after drying overnight afforded a colorless glissy solid. Yield: 9.57 g (64%). HPLC: >98% Rf = 14.8 minutes. Anal. for C25H34N8O6: 0.25 H2O Found (calc.) C: 49.29 (49.42); H: 6.52 (6.35); N: 14.11 (14.41). Due to the limited rotation around the secondary amide, several of the signals were doubled in the ratio 2:1 (indicated in the list by mj. for major and mi. for minor). 

H-NMR (250 MHz, DMSO-d6): 8.12 (bs, 1H, CO2H); 11.28 (s, 1H, mide NH); 11.26 (s, 1H, mide NH); 7.30 (s, 1H, mide NH); 7.26 (s, 1H, mide NH); 6.92 (bs, 1H, mide BOC-NH); 6.73 (bt, 1H, mide BOC-NH); 4.64 (s, 2H, mide CH2CON); 4.46 (s, 2H, mide CH2CON); 4.19 (s, 2H, mide CH2CON); 3.97 (s, 2H, mide CH2CON); 2.90-3.01 (unsolved m. includes water, CH2CH2); 1.75 (s, 3H, CH3) and 1.38 (s, 9H, t-Bu).

**EXAMPLE 25**

Synthesis of N'-benzyloxyacarbonyl-1-(BOC-aeg) cytosine

N'-BOC-aminooxy acetyl methyl ester (5.2 g, 20.3 mmol), DiBOH (3.64 g, 22.3 mmol) and N'-BOC-aminooxy acetyl methyl cytosine (6.77 g, 22.3 mmol) were suspended in DMF (100 mL). Methylene chloride (100 mL) was added. The solution was cooled to 0°C and DCC (5.03 g, 24.4 mmol) was added. The ice bath was removed after 2 h and stirring was continued for another hour at ambient temperature. The reaction mixture was then evaporated to dryness, in vacuo. The residue was suspended in ether (100 mL) and stirred vigorously for 30 minutes. The solid material was isolated by filtration and the ether wash procedure was repeated twice. The material was then stirred vigorously for 15 minutes with dilute sodium hydrogen carbonate (approx. 4% solution, 100 mL), filtered, and washed with water. This procedure was then repeated once, which after drying left 17 g of yellowish solid material. The solid was then refluxed with dioxane (200 mL) and filtered while hot. After cooling, water (200 mL) was added. The precipitated material was isolated by filtration, washed with water, and dried. According to HPLC (observing at 260 nm) this material has a purity higher than 99%, besides the DCU. The ester was then suspended in THF (100 mL), cooled to 0°C, and 1N LiOH (61 mL) was added. After stirring for 15 minutes, the mixture was filtered and the filtrate was washed with methylene chloride (2x150 mL). The alkaline solution then was cooled to 0°C and the pH was adjusted to 2.0 with 1N HCl. The title compound was isolated by filtration and was washed once with water, leaving 11.3 g of a white powder after drying. The material was suspended in methylene chloride (300 mL) and petroleum ether (300 mL) was added. Filtration and wash afforded 7.1 g (69%) after drying. HPLC showed a purity of 99% Rf = 19.5 minutes, and a minor impurity at 12.6 minutes (approx. 1%) most likely the Z-deprotected monomer. Anal. for C25H34N8O6 found (calc.) C: 54.16 (54.87); H: 5.76 (5.81) and N: 13.65 (13.91). 

H-NMR (250 MHz, DMSO-d6): 10.78 (bs, 1H, CO2H); 7.88 (2 overlapping doublets, 1H, Cyt H-5); 7.41-7.32 (m, 5H, Ph); 7.01 (2 overlapping doublets, 1H, Cyt H-6); 6.94 & 6.78 (unsolved triplets, 1H, BOC-NH); 5.19 (s, 2H, PhCH2); 4.81 & 4.62 (s, 2H, CH2CON); 4.17 & 3.98 (s, 2H, }
CH₃CO₂H; 3.42-3.03 (m, includes water, CH₃CH₂), and 1.35 & 1.37 (s, 9H, Bu). 13C-NMR: 150.88; 128.52; 128.18; 127.96; 93.90; 66.53; 49.58 and 28.22. IR: Frequency in cm⁻¹ (intensity). 3423 (26.4), 3035 (53.2), 2978 (41.4), 1736 (17.3), 1658 (3.8), 1563 (23.0), 1505 (6.8) and 1456 (26.4).

EXAMPLE 26

Synthesis of 9-carboxymethylenedine ethyl ester

Adenine (10 g, 74 mmol) and potassium carbonate (10.29 g, 74 mmol) were suspended in DMF and ethyl bromoacetate (8.24 mL, 74 mmol) was added. The suspension was stirred for 2.5 h under nitrogen at room temperature and then filtered. The solid residue was washed three times with DMF (10 mL). The combined filtrate was evaporated to dryness, in vacuo. Water (200 mL) was added to the yellowish-orange solid material and the pH adjusted to 6 with 4N HCl. After stirring at 0°C for 10 minutes, the solid was filtered off, washed with water, and recrystallized from 96% ethanol (150 mL). The title compound was isolated by filtration and washed with ether. Yield: 3.4 g (20%), M.p. 215.5-220°C. Anal. for C₉H₈N₂O₂ found (calc.): C: 48.86(48.65); H: 5.01(4.91); N: 31.66(31.42). 1H-NMR (250 MHz; DMSO-d₆): 7.25 (bs, 2H, NH₂), 5.06 (2 H, NCH₃), 4.17 (q, 2H, J=7.11 Hz, OCH₂) and 1.21 (t, 3H, J=7.13 Hz, NCH₃). 13C-NMR: 152.70, 141.30, 61.43, 43.97 and 14.07. FAB-MS: 222 (MH+); IR: Frequency in cm⁻¹ (intensity). 3855 (54.3), 3274(10.4), 3246 (14.0), 3117(5.3), 2989(22.3), 2940 (33.9), 2876(43.4), 2753(49.0), 2346 (56.1), 2106 (57.1), 1899 (55.7), 1762 (14.2), 1742 (14.2), 1742 (1.0), 1671 (18.7), 1644 (10.9), 1600 (6.6), 1582 (7.1), 1522 (43.3), 1477 (7.2), 1445 (35.8) and 1482 (8.6).

The position of alkylation was verified by X-ray crystallography on crystals, which were obtained by recrystallization from 96% ethanol.

Alternatively, 9-carboxymethylenedine ethyl ester can be prepared by the following procedure. To a suspension of adenine (50 g, 0.37 mol) in DMF (1100 mL) in 2 l three-necked flask equipped with a nitrogen inlet, a mechanical stirrer and a dropping funnel, was added 16.4 g (0.407 mol) of hexane-washed sodium hydride-mineral oil dispersion. The mixture was stirred vigorously for 2 hours, after which ethyl bromacetate (75 mL, 0.67 mol) was added dropwise over the course of 3 hours. The mixture was stirred for one additional hour, after which the mixture was indicated conversion of the title compound. The mixture was evaporated to dryness at 1 mm Hg and water (500 mL) was added to the oily residue which caused crystallization of the title compound. The solid was recrystallized from 96% ethanol (600 mL). Yield (after drying): 53.7 g (65.6%). HPLC (215 nm) purity >99.5%.

EXAMPLE 27

Synthesis of N⁶-benzoxycarbonyl-9-carboxymethylenedine ethyl ester

9-Carboxymethylenedine ethyl ester (3.4 g, 15.4 mmol) was dissolved in dry DMF (50 mL) by gentle heating, cooled to 20°C, and added to a solution of N-ethylbenzoxycarbonylmalodiozo tetrafluoroborate (62 mmol) in methylene chloride (50 mL) over a period of 15 minutes in an ice bath. Some precipitation was observed. The ice bath was removed and the solution was stirred overnight. The reaction mixture was treated with saturated sodium hydroxide carbonate (100 mL). After stirring for 10 minutes, the phases were separated and the organic phase was washed successively with one volume of water, dilute potassium hydrogen sulfate (twice), and with saturated sodium chlo-
35 ml.) and activated carbon was added. After stirring for 5 minutes, the solution was filtered. The filtrate was mixed with water (30 ml.) and was allowed to stir overnight. The next day, the white precipitate was removed by filtration, washed with water, and dried, affording 1.16 g (26%) of a material with a purity higher than 98% by HPLC. Addition of water to the mother liquor afforded another 0.53 g of the product with a purity of approx. 95%. Anal. for C₂₅H₂₃N₂O₅H₂O found (calc.): C: 55.01 (54.44); H: 6.85 (6.15) and N: 16.47 (17.09). 1H NMR (250 MHz, CDCl₃) 8.74 (s, 1H, Ade H-2); 8.18 (b, s, 1H, ZNH); 8.10 and 8.04 (s, 1H, H-8); 7.46-7.34 (m, 5H, Ph); 5.63 (unres. t, 1H, BOC-NH); 5.30 (s, 2H, PhCH₃); 5.16 & 5.00 (s, 2H, CH₂CON); 4.29 & 4.06 (s, 2H, CH₂CO₂H); 4.20 (q, 2H, OCH₂CH₃); 3.67-3.29 (m, 4H, CH₂CH₂CH₂); 1.42 (s, 9H, t-Bu) and 1.27 (t, 3H, OCH₃CH₂). The spectrum shows traces of ethanol and DCC.

**EXAMPLE 30**

Synthesis of N²-benzylxocarbonyl-1-(BOC-aeg)adenine

N²-Benzyloxycarbonyl-1-(BOC-aeg)adenine ethyl ester (1.48 g, 2.66 mmol) was suspended in THF (13 ml) and the mixture was cooled to 0°C. Lithium hydroxide (8 ml, 1N) was added. After 15 minutes of stirring, the reaction mixture was filtered, and the filtered solution was washed with methylene chloride (2×25 ml). The pH of the aqueous solution was adjusted to 2 with 1N HCl. The precipitate was isolated by filtration, washed with water, and dried, affording 0.82 g (58%) of the product. The product was additionally precipitated twice with methylene chloride/petroleum ether. Yield (after drying): 0.77 g (55%). M.p. 119°C. (decomp.). Anal. for C₃₅H₂₉N₄O₄·H₂O found (calc.) as C: 53.32 (52.84); H: 5.71 (5.73); N: 17.68 (17.97). PAB-MS, 528.5 (M+H+). 2H NMR (250 MHz, DMSO-d₆), 12.75 (very br. 1H, CO₂H); 10.65 (b, 1H, ZNH); 8.59 (d, 1H, J=2.14 Hz, Ade H-2); 8.31 (s, 1H, Ade H-8); 7.49-7.31 (m, 5H, Ph); 7.03 & 6.75 (unresol. t, 1H, BOC-NH), 5.33 & 5.16 (s, 2H, CH₂CON); 5.22 (s, 2H, PhCH₃); 4.34-3.99 (s, 2H, CH₂CO₂H); 3.54-3.03 (m, ‘m’ includes water, CH₂CH₂ and 1.39 & 1.37 (s, 9H, t-Bu). 13C-NMR, 170.4; 166.6; 152.3; 151.5; 149.5; 145.2; 128.5; 128.0; 127.9; 66.32; 46.77; 46.03; 43.87 and 28.24.

**EXAMPLE 31**

Synthesis of 2-amino-6-chloro-9-carboxymethylpyrine

To a suspension of 2-amino-6-chloropyrurine (1.50 g, 29.6 mmol) and potassium carbonate (12.91 g, 93.5 mmol) in DMF (50 ml) was added bromoacetic acid (4.7 g, 22.8 mmol). The mixture was stirred vigorously for 20 h under nitrogen. Water (150 ml) was added and the solution was filtered through celite to give a clear yellow solution. The solution was acidified to a pH of 3 with 4N hydrochloric acid. The precipitate was filtered and dried in vacuo over silica. Yield: 3.02 g (44.8%). 1H NMR(DMSO-d₆) δ: 8.48 ppm (s, 2H); 6.95 (s, 2H); 8.10 (s, 1H).

**EXAMPLE 32**

Synthesis of 2-amino-6-benzylxocarbonyl-9-carboxyethylpyrine

Sodium (2 g 87 mmol) was dissolved in benzyl alcohol (20 ml) and heated to 130°C and 2 h. After cooling to 0°C, a solution of 2-amino-6-chloro-9-carboxyethylpyrine (4.05 g, 18 mmol) in DMF (85 ml) was slowly added, and the resulting suspension stirred overnight at 20°C. Sodium hydroxide solution (1N, 100 ml) was added and the clear solution was washed with ethyl acetate (3×100 ml). The water phase was then acidified to a pH of 3 with 4N hydrochloric acid. The precipitate was taken up in ethyl acetate (200 ml), and the water phase was extracted with ethyl acetate (2×100 ml). The combined organic phases were washed with saturated sodium chloride solution (2×75 ml), dried with anhydrous sodium sulfate, and evaporated to dryness, in vacuo. The residue was recrystallized from ethanol (300 ml). Yield, after drying in vacuo, was 10.5 g (52%). M.p. 159°C-165°C. Anal. (calc.): C, 56.18; 55.97; H(4.38; 4.32); N(23.4; 23.10). 1H NMR (DMSO-d₆) δ: 84.82 (s, 2H); 5.51 (s, 2H); 6.45 (s, 2H); 7.45 (m, 5H); 7.82 (s, 1H).

**EXAMPLE 33**


2-Amino-benzylxocarbonyl-9-carboxymethylpyrine (0.5 g, 1.67 mmol), methyl-N-[2-tert-butoxycarbonylaminol][ethy]glycinate (0.65 g, 2.8 mmol), diisopropylethylamine (0.54 g, 4.19 mmol) and bromo-tris-pyridiido-phosphoninohexafluoro-phosphate (PyBroP8) (0.798 g, 1.71 mmol) were stirred in DMF (2 ml) for 4 h. The clear solution was poured into an ice-cooled solution of sodium hydrogen carbonate (1N, 40 ml) and extracted with ethyl acetate (3×40 ml). The organic layer was washed with potassium hydrogen sulfate solution (1N, 2×40 ml), sodium hydrogen carbonate (1N, 1×40 ml) and saturated sodium chloride solution (60 ml). After drying with anhydrous sodium sulfate and evaporation in vacuo, the solid residue was recrystallized from 2:1 ethyl acetate/hexane (20 ml) to give the methyl ester in 63% yield. (MS-FA6 514 (M+1)). Hydrolysis was accomplished by dissolving the ester in 1:2 ethanol/water (30 ml) containing concentrated sodium hydroxide (1 ml). After stirring for 2 h, the solution was filtered and acidified to a pH of 3, by the addition of 4N hydrochloric acid. The title compound was obtained by filtration. Yield: 370 mg (72% for the hydrolysis). Purity by HPLC was more than 99%. Due to the limited rotation around the secondary amide several of the signals were doubled in the ratio 2:1 (indicated in the list by m for major and ml for minor). 1H NMR(250 MHz, DMSO-d₆) δ 1:4 (s, 9H); 3.2 (m, 2H); 3.6 (m, 2H); 4.1 (d, 1H, CONRCH₂COOH); 4.4 (s, ml, CONRCH₂COOH); 5.0 (s, ml, Gua-CH₂CO-); 5.2 (s, ml, Gua-CH₂CO); 5.6 (s, 2H); 6.5 (s, 2H); 6.9 (m, ml, BOC-NH); 7.1 (m, ml, BOC-NH); 7.5 (m, 3H); 7.8 (s, 1H); 12.8 (s, 1H). 13C-NMR, 170.95; 170.52; 167.29; 166.85; 160.03; 159.78; 155.84; 154.87; 140.63; 136.76; 128.49; 128.10; 113.04; 78.19; 77.86; 66.95; 49.22; 47.70; 46.94; 45.96; 43.62; 43.31 and 28.25.

**EXAMPLE 34**

Synthesis of 3-BOC-amino-1.2-propanediol

3-Amino-1.2-propanediol (1 equivalent, 40 g, 0.44 mol) was dissolved in water (1000 ml) and cooled to 0°C. Di-tert-butyl dicarbonate (1.2 equivalents, 115 g, 0.526 mol) was added in one portion. The reaction mixture was heated to room temperature on a water bath during stirring. The pH was maintained at 10.5 with a solution of sodium hydroxide (1 equivalent, 17.56 g, 0.44 mol) in water (1200 ml). When the addition of aqueous sodium hydroxide was completed, the reaction mixture was stirred overnight at room temperature. Subsequently, ethyl acetate (750 ml) was added to the reaction mixture. Followed by cooling to 0°C. The pH was
adjusted to 2.5 with 4N sulphuric acid with vigorous stirring. The phases were separated and the water phase was washed with additional ethyl acetate (6×350 mL). The volume of the organic phase was reduced to 900 mL by evaporation under reduced pressure. The organic phase was then washed with a saturated aqueous solution of potassium hydrogen sulfate diluted to twice its volume (1×1000 mL) and with saturated aqueous sodium chloride (1×500 mL). The organic phase was dried (MgSO₄) and evaporated under reduced pressure to yield 50.12 g (60%) of the title compound. The product could be solidified by evaporation from methylene chloride and subsequent freezing. ¹H-NMR (CDCl₃/TMS) δ: 1.43 (t, 9H, Me C), 3.25 (m, 2H, CH₂), 3.57 (m, 2H, CH), 3.73 (m, 1H, CH), 1.3 C-NMR (CDCl₃/TMS) ppm: 28.2 (Me₂C), 42.6 (CH₂), 63.5, 71.1 (CH₂OH, CHOH), 79.5 (Me₂C), 157.0 (C=O).

**EXAMPLE 35**

Synthesis of 2-(BOC-aminomethyl)-L-alanine methyl ester
3-BOC-aminobutyraldehyde (1 equivalent, 20.76 g, 0.109 mol) was suspended in water (150 mL). Potassium periodate (1 equivalent, 24.97 g, 0.109 mol) was added and the reaction mixture was stirred for 2 h at room temperature under nitrogen. The reaction mixture was filtered and the water phase extracted with chloroform (2×250 mL). The organic phase was dried (MgSO₄) and evaporated to afford an almost quantitative yield of BOC-aminoacetaldehyde as a colorless oil, which was used without further purification in the following procedure.

Palladium on carbon (10%, 0.8 g) was added to MeOH (250 mL) under nitrogen with cooling (0° C.) and vigorous stirring. Anhydrous sodium acetate (2 equivalents, 4.49 g, 54.7 mmol) and L-alanine methyl ester, hydrochloride (1 equivalent, 3.82 g, 27.4 mmol) were added. BOC-aminoacetaldehyde (4.79 g, 30.1 mmol, 1.1 eqv) was dissolved in MeOH (150 mL) and added to the reaction mixture. The reaction mixture was hydrogenated at atmospheric pressure and room temperature until hydrogen uptake had ceased. The reaction mixture was filtered through celite, which was washed with additional MeOH. The MeOH was removed under reduced pressure. The residue was suspended in water (150 mL) and the pH adjusted to 8 by dropwise addition of 0.5N NaOH with vigorous stirring. The water phase was extracted with methylene chloride (4×250 mL). The organic phase was dried (MgSO₄), filtered through celite, and evaporated under reduced pressure to yield 63.6 g (94%) of the title compound as a clear, pale yellow oil. MS (FAB-MS): m/z (%)=247 (100), M+1, 191 (90), 147 (18). ¹H-NMR (250 MHz, CDCl₃) δ: 1.18 (d, J=7.0 Hz, 3H, Me), 1.36 (s, 9H, Me₂C), 1.89 (b, 1H, NH), 2.51 (m, 1H, CH), 2.66 (m, 1H, CH), 3.10 (m, 2H, CH₂), 3.27 (q, J=7.0 Hz, 1H, CH), 3.64 (s, 3H, OMe), 5.06 (b, 1H, carbamate NH), ¹³C-NMR (ppm): 18.8 (Me₂C), 28.2 (Me₂C), 40.1, 47.0 (CH₂), 51.6 (OMe), 56.0 (CH), 155.8 (carbamate C=O), 175.8 (ester C=O).

**EXAMPLE 36**

Synthesis of N-(BOC-aminomethyl)-L-(1-thiinylacetyl)-L-alanine methyl ester
To a solution of BOC-aminomethyl-L-alanine methyl ester (1.23 g, 5 mmol) in DMF (10 mL) was added Dibb-OH (0.9 g, 5.52 mmol) and 1-thiinylacetic acid (1.01 g, 5.48 mmol). When 1-thiinylacetic acid dissolved, dialchomethane (10 mL) was added and the solution was cooled in an ice bath. After the reaction mixture had reached 0° C., DCC (1.24 g, 6.01 mmol) was added. Within 5 minutes after the addition, a precipitate of DCU was seen. After a further 5 minutes, the ice bath was removed. Two hours later, tlc analysis showed the reaction to be complete. The mixture was filtered and the precipitate washed with dichloromethane (100 mL). The resulting solution was extracted twice with 5% sodium hydrogencarbonate (150 mL) and twice with saturated potassium hydrogen sulfate (25 mL) in water (100 mL). After a final extraction with saturated sodium chloride (150 mL), the solution was dried with magnesium sulfate and evaporated to give a white foam. The foam was purified by column chromatography on silica gel using dichloromethane with a methanol gradient as eluent. This yielded a pure compound (>99% by HPLC) (1.08 g, 52.4%). FAB-MS: 413 (M+1) and 431 (M+1water). ¹H-NMR (CDCl₃) δ: 4.52 (s, 2H, CH₂); 3.73 (s, 3H, OMe); 3.2-3.6 (m, 4H, ethyl CH₃); 1.90 (s, 3H, Me in T); 1.49 (d, 3H, Me in Ala; J=7.3 Hz); 1.44 (s, 9H, BOC).

**EXAMPLE 37**

Synthesis of N-(BOC-aminomethyl)-N-(1-thiinylacetyl)-L-alanine
The methyl ester of the title compound (2.07 g, 5.02 mmol) was dissolved in methanol (100 mL) and cooled in an ice bath. 2M Sodium hydroxide (100 mL) was added. After stirring for 10 minutes, the pH of the mixture was adjusted to 3 with 4M hydrogen chloride. The solution was subsequently extracted with ethyl acetate (3×100 mL). The combined organic extracts were dried over magnesium sulfate. After evaporation, the resulting foam was dissolved in ethyl acetate (400 mL) and a 5 mL of methanol to dissolve the solid material. Petroleum ether was then added until precipitation started. After allowing the mixture to stand overnight at −20° C., the precipitate was removed by filtration. This yielded 1.01 g (50.5%) of pure compound (>99% by HPLC). The compound was recrystallized from 2-propanol. FAB-MS: 399 (M+1). ¹H-NMR (DMSO-d₆) δ: 11.35 (s, 1H, COO); 7.42 (s, 1H, H₂); 4.69 (s, 2H, CH₂); 1.83 (s, 3H, Me in T); 1.50–1.40 (m, 12H, Me in Ala+BOC).

**EXAMPLE 38**

(a) Synthesis of N-(BOC-aminomethyl)-N-(1-thiinylacetyl)-D-alanine methyl ester
To a solution of BOC-aminomethyl alanine methyl ester (2.48 g, 10.1 mmol) in DMF (20 mL) was added Dibb-OH (1.8 g, 11 mmol) and thyminecadic acid (2.14 g, 11.6 mmol). After dissolution of 1-thiinylacetic acid, methylene chloride (20 mL) was added and the solution cooled in an ice bath. When the reaction mixture had reached a temperature of 0° C., DCC (2.88 g, 14 mmol) was added. Within 5 minutes of the addition, a precipitate of DCU was seen. After 35 minutes the ice bath was removed. The reaction mixture was filtered 3.5 h later and the precipitate washed with methylene chloride (200 mL). The resulting solution was extracted twice with 5% sodium hydrogen carbonate (200 mL) and twice with saturated potassium hydrogen sulfate in water (100 mL). After final extraction with saturated sodium chloride (250 mL), the solution was dried with magnesium sulfate and evaporated to give an oil. The oil was purified by short column silica gel chromatography using methylene chloride with a methanol gradient as eluent. This yielded a compound which was 96% pure according to HPLC (1.05 g, 25.3%) after precipitation with petroleum ether. FAB-MS: 413 (M+1). ¹H-NMR (CDCl₃) δ: 5.64 (t, 1H, BOCNH; J=5.89 Hz); 4.56 (d, 2H, CH₂); 4.35 (q, 1H, CH in Ala; J=7.25 Hz); 3.74 (s, 3H, OMe); 3.64–3.27 (m, 4H, ethyl H's); 1.90 (s, 3H, Me in T); 1.52–1.44 (t, 12H, BOC +Me in Ala).
(h) **Synthesis of N-((BOC-aminoethyl)-N-((1-thyminyl)acetyl)-D-alanine**

The methyl ester of the title compound (1.57 g, 3.81 mmol) was dissolved in methanol (100 mL) and cooled in an ice bath. Sodium hydroxide (2M, 100 mL) was added. After stirring for 10 minutes, the pH of the mixture was adjusted to 3 with 4M hydrogen chloride. The solution was then extracted with ethyl acetate (3x100 mL). The combined organic extracts were dried over magnesium sulfate. After evaporation, the oil was dissolved in ethyl acetate (200 mL). Petroleum ether was added (to a total volume of 600 mL) until precipitation started. After standing overnight at -20°C, the precipitate was removed by filtration. This afforded 1.02 g (67.3%) of the title compound, which was 94% pure according to HPLC: FAB-MS: 399 (M+1), 'H-NMR, δ: 11.34 (s, 1H, COOH); 7.42 (s, 1H, H2); 4.69 (s, 2H, CH2); 4.40 (q, 1H, CH in Ala, J=7.20 Hz); 1.83 (s, 3H, Me in T); 1.52-1.40 (m, 12H, BOC+Me in Ala).

**EXAMPLE 39**

Synthesis of N-(N'-BOC-3'-aminopropyl)-N-N-(1-thyminyl)-acetyl)glycine methyl ester

N-(N'-BOC-3'-aminopropyl)-glycine methyl ester (2.84 g, 0.0115 mol) was dissolved in DMF (35 mL), followed by addition of DibOH (2.07 g, 0.0127 mol) and 1-thyminylacetic acid (2.34 g, 0.0127 mol). Methylene chloride (35 mL) was added and the mixture cooled to 0°C in an ice bath. After addition of DCC (2.85 g, 0.0138 mol), the mixture was stirred at 0°C for 2 h. followed by 1 h at room temperature. The precipitated DCU was removed by filtration, washed with methylene chloride (25 mL), and a further amount of methylene chloride (150 mL) was added to the filtrate. The organic phase was extracted with sodium hydrogen carbonate (1 volume saturated diluted with 1 volume water, 6x250 mL), potassium sulfate (1 volume saturated diluted with 4 volumes water, 3x250 mL), and saturated aqueous sodium chloride (1x250 mL), dried over magnesium sulfate, and evaporated to dryness in vacuo. The solid residue was suspended in methylene chloride (35 mL) and stirred for 1 h. The precipitated DCU was removed by filtration and washed with methylene chloride (25 mL). The filtrate was evaporated to dryness in vacuo, and the residue purified by column chromatography on silica gel, eluting with a mixture of methanol and methylene chloride (1 volume saturated diluted with 1 volume water, 6x100 mL), potassium sulfate (1 volume saturated diluted with 4 volumes water, 3x100 mL), and saturated aqueous sodium chloride (1x100 mL), dried over magnesium sulfate, and evaporated to dryness in vacuo. The solid residue was suspended in methylene chloride (15 mL), and stirred for 1 h. The precipitated DCU was removed by filtration and washed with methylene chloride. The filtrate was evaporated to dryness in vacuo, and the residue purified by column chromatography on silica gel, eluting with a mixture of methanol and methylene chloride (1 volume saturated diluted with 1 to 6% methanol in methylene chloride). This afforded the title compound as a white solid (3.05 g, 64%). M.p. 76°-79°C (decomp.). Anal. for C16H28N4O5, found (calc.) C: 52.03 (52.42) H: 6.90 (6.84) N: 13.21 (13.58). The compound showed satisfactory 'H and 13C-NMR spectra.

**EXAMPLE 40**

Synthesis of N-(N'-BOC-3'-aminopropyl)-N-((1-thyminyl)-acetyl)glycine

N-(N'-BOC-3'-aminopropyl)-N-((1-thyminyl)acetyl) glycine methyl ester (3.02 g, 0.00732 mol) was dissolved in methanol (25 mL) and stirred for 1.5 h with 2M sodium hydroxide (25 mL). Methylene chloride was removed by evaporation in vacuo, and the pH adjusted to 2 with 4M hydrochloric acid at 0°C. The product was isolated as white crystals by filtration, washed with water (3x10 mL), and dried over silica gel in vacuo. Yield: 2.19 g (75%). Anal. for C17H26N4O5, H2O, found (calc.) C: 45.95 (49.03) H: 6.47 (6.29) N: 13.43 (13.45). The compound showed satisfactory 'H and 13C-NMR spectra.

**EXAMPLE 41**

Synthesis of 3-(1-thyminyl)propanoic acid methyl ester

Thymine (14 g, 0.11 mol) was suspended in methanol. Methyl acrylate (39.6 mL, 0.44 mol) was added, along with catalytic amounts of sodium hydroxide. The solution was refluxed in the dark for 45 h, evaporated to dryness in vacuo, and the residue dissolved in methanol (8 mL) with heating. After cooling in an ice bath, the product was precipitated by addition of ether (20 mL), isolated by filtration, washed with ether (3x15 mL), and dried over silica gel, in vacuo. Yield: 11.23 g (48%). M.p. 112°-119°C. Anal. for C16H26N4O4, found (calc.) C: 51.14 (50.94) H: 5.78 (5.70) N: 11.52 (13.20). The compound showed satisfactory 'H and 13C-NMR spectra.

**EXAMPLE 42**

Synthesis of 3-(1-thyminyl)propanoic acid methyl ester

3-(1-Thyminyl)propanoic acid methyl ester (1 g, 0.0047 mol) was suspended in 2M sodium hydroxide (15 mL), refluxed for 10 minutes. The pH was adjusted to 0.3 with conc. hydrochloric acid. The solution was extracted with ethyl acetate (10x25 mL). The organic phase was extracted with saturated aqueous sodium chloride, dried over magnesium sulfate, and evaporated to dryness in vacuo, to give the title compound as a white solid (0.66 g, 71%). M.p. 118°-121°C. Anal. for C18H27N4O4, found (calc.) C: 48.38 (48.49) H: 5.09 (5.09) N: 13.93 (14.14). The compound showed satisfactory 'H and 13C-NMR spectra.

**EXAMPLE 43**

Synthesis of N-(N'-BOC-aminoethyl)-N-N-(1-thyminyl)-propionyl)glycine ethyl ester

N-(N'-BOC-aminoethyl)glycine ethyl ester (1 g, 0.0041 mol) was dissolved in DMF (12 mL). DibOH (0.73 g, 0.0045 mol) and 3-(1-thyminyl)propanoic acid (0.89 g, 0.0045 mol) were added. Methylene chloride (12 mL) was then added and the mixture was cooled to 0°C in an ice bath. After addition of DCC (1.01 g, 0.0049 mol), the mixture was stirred at 0°C for 2 h, followed by 1 h at room temperature. The precipitated DCU was removed by filtration, washed with methylene chloride (25 mL), and a further amount of methylene chloride (50 mL) was added to the filtrate. The organic phase was extracted with sodium hydrogen carbonate (1 volume saturated diluted with 1 volume water, 6x100 mL), potassium sulfate (1 volume saturated diluted with 4 volumes water, 3x100 mL), and saturated aqueous sodium chloride (1x100 mL), dried over magnesium sulfate, and evaporated to dryness in vacuo. The solid residue was suspended in methylene chloride (15 mL), and stirred for 1 h. The precipitated DCU was removed by filtration and washed with methylene chloride. The filtrate was evaporated to dryness in vacuo, and the residue purified by column chromatography on silica gel, eluting with a mixture of methanol and methylene chloride (gradient from 1 to 6% methanol in methylene chloride). This afforded the title compound as a white solid (1.02 g, 59%). Anal. for C16H26N4O4, found (calc.) C: 53.15 (53.51) H: 6.90 (7.09) N: 12.76 (13.13). The compound showed satisfactory 'H and 13C-NMR spectra.

**EXAMPLE 44**

Synthesis of N-(N'-BOC-aminoethyl)-N-(1-thyminyl)-propanoyl)glycine

N-(N'-BOC-aminoethyl)-N-(1-thyminyl)-propanoyl) glycine ethyl ester (0.83 g, 0.00195 mol) was dissolved in...
methanol (25 mL). Sodium hydroxide (2M, 25 mL) was added. The solution was stirred for 1 h. The methanol was removed by evaporation in vacuo, and the pH adjusted to 2 with 4M hydrochloric acid at 0°C. The product was isolated by filtration, washed with ether (3×15 mL), and dried over anhydrous calcium sulfate. Yield: 0.769 g (99%). M.p. 213°C (decromp.).

**EXAMPLE 45**

Synthesis of Mono-BOC-ethylenediamine (2)

tert-Butyl-4-nitrophenyl carbonate (1, 10.9 g, 0.0418 mol) dissolved in DMF (50 mL) was added dropwise over a period of 30 minutes to a solution of ethylenediamine (27.9 mL, 0.418 mol) and DMF (50 mL) and stirred overnight. The mixture was evaporated to dryness in vacuo, and the resulting oil dissolved in water (250 mL). After cooling to 0°C, the pH was adjusted to 3.5 with 4M hydrochloric acid. The solution was then filtered and exchanged with chloroform (3×250 mL). The pH was adjusted to 12 (at 0°C) with 2M sodium hydroxide, and the aqueous solution extracted with methylene chloride (3×300 mL). After treatment with a solution of saturated aqueous sodium chloride (250 mL), the methylene chloride solution was dried over magnesium sulfate. After filtration, the solution was evaporated to dryness in vacuo, resulting in 4.22 g (63%) of the product as an oil. 1H-NMR (90 MHz, CDCl3) δ: 1.44 (s, 9H); 2.87 (t, 2H); 3.1 (q, 2H); 5.62 (sb).

**EXAMPLE 46**

Synthesis of (N-BOC-aminoethyl)-p-alanine methyl ester.HCl

Mono-BOC-ethylenediamine (2) (16.28 g, 0.102 mol) was dissolved in acetonitrile (400 mL) and methyl acrylate (91.5 mL, 1.02 mol) was transferred to the mixture with acetonitrile (200 mL). The solution was refluxed overnight under nitrogen in the dark to avoid polymerization of methyl acrylate. After evaporation to dryness in vacuo, a mixture of water and ether (200 mL+200 mL) was added, and the solution was filtered and vigorously stirred. The aqueous phase was extracted one additional time with ether and then freeze dried to yield a yellow solid. Recrystallization from ethyl acetate yielded 13.09 g (46%) of the title compound. M.p. 138°-140°C. Anal. for C13H17NO3Cl, found (calc.) C: 46.49 (46.72) H: 8.38 (8.20) N: 9.83 (9.91) Cl: 12.45 (12.54). 1H-NMR (90 MHz; DMSO-d6) δ: 1.39 (s, 9H); 2.9 (m, 8H); 3.64 (s, 3H).

**EXAMPLE 47**

Synthesis of N-[(1-Thyminyl)acetyl]-N'-BOC-aminoethyl-β-alanine methyl ester

(N-BOC-aminoethyl)-β-alanine methyl ester.HCl (3) (2 g, 0.0071 mol) and (1-N-4-Z)-cytosinylacetic acid pentafluorophenyl ester (5) (2.828 g, 0.00812 mol) were dissolved in DMF (50 mL). Triethyl amine (1.12 mL, 0.00812 mol) was added and the mixture stirred overnight. After addition of methylene chloride (200 mL), the organic phase was extracted with aqueous sodium hydroxide (3×250 mL), half-saturated solution of aqueous potassium hydrogen sulfate (3×250 mL), and saturated solution of aqueous sodium chloride (250 mL), and dried over magnesium sulfate. Filtration and evaporation to dryness in vacuo, resulted in a yield of 2.9 g (99%) of product (oil). 1H-NMR (250 MHz; CDCl3; due to limited rotation around the secondary amide several of the signals were doubled) δ: 1.43 (s, 9H); 1.88 (s, 3H); 2.63 (t, 1H); 2.74 (t, 1H); 3.25-3.35 (4xt, 8H); 3.65 (2xt, 2H); 3.66 (s, 1.5); 3.72 (s, 1.5); 4.61 (s, 1H); 4.72 (s, 2H); 5.59 (s, 0.5H); 5.96 (s, 0.5H); 7.11 (s, 1H); 10.33 (s, 1H).

**EXAMPLE 48**

Synthesis of N-[(1-Thyminyl)acetyl]-N'-BOC-aminoethyl-β-alanine methyl ester (3 g, 0.0073 mol) was dissolved in 2M sodium hydroxide (30 mL), the pH adjusted to 2 at 0°C with 4M hydrochloric acid, and the solution stirred for 2 h. The precipitate was isolated by filtration, washed three times with cold water, and dried over anhydrous calcium sulfate. Yield: 2.23 g (77%). M.p. 170°-176°C. Anal. for C12H19NO4, H2O, found (calc.) C: 49.49 (49.03) H: 6.31 (6.78) N: 13.84 (13.45). 1H-NMR (90 MHz; DMSO-d6) δ: 1.38 (s, 9H); 1.76 (s, 3H); 2.44 and 3.29 (m, 8H); 4.55 (s, 2H); 7.3 (s, 1H); 11.23 (s, 1H). FAB-MS: 399 (M+1).

**EXAMPLE 49**

Synthesis of N-[(1-N4-Z)-cytosinylacetyl]-N'-BOC-aminoethyl-β-alanine methyl ester

(N-BOC-aminoethyl)-β-alanine methyl ester.HCl (3) (2 g, 0.0071 mol) and (1-N-4-Z)-cytosinylacetic acid pentafluorophenyl ester (5) (3.319 g, 0.0071 mol) were dissolved in DMF (50 mL). Triethyl amine (0.99 mL, 0.0071 mol) was added and the mixture stirred overnight. After addition of methylene chloride (200 mL), the organic phase was extracted with aqueous sodium hydroxide carbonate (3×250 mL), half-saturated solution of aqueous potassium hydrogen sulfate (3×250 mL), and saturated solution of aqueous sodium chloride (250 mL), and dried over magnesium sulfate. Filtration and evaporation to dryness in vacuo, resulted in 3.36 g of solid compound which was recrystallized from methanol. Yield: 2.42 g (64%). M.p. 158°-161°C. Anal. for C12H16N2O4, found (calc.) C: 55.19 (56.49) H: 6.19 (6.26) N: 12.86 (13.18). 1H-NMR (250 MHz; CDCl3; due to limited rotation around the secondary amide several of the signals were doubled) δ: 1.43 (s, 9H); 2.57 (t, 1H); 3.60–3.23 (m’s, 6H); 3.60 (s, 3H); 3.66 (s, 1.5H); 4.80 (s, 1H); 4.88 (s, 1H); 5.20 (s, 2H); 7.80-7.25 (m’s, 7H). FAB-MS: 532 (M+1).

**EXAMPLE 50**

Synthesis of N-[(1-N4-Z)-cytosinylacetyl]-N'-BOC-aminoethyl-β-alanine methyl ester (0.621 g, 0.0012 mol) was dissolved in 2 M sodium hydroxide (8.5 mL) and stirred for 2 h. Subsequently, the pH was adjusted to 2 at 0°C with 4M hydrochloric acid and the solution stirred for 2 h. The precipitate was isolated by filtration, washed three times with cold water, and dried over anhydrous calcium sulfate. Yield: 0.326 g (54%). The white solid was recrystallized from 2-propanol and washed with petroleum ether. M.p. 163°-165°C (decom). Anal. for C12H16N2O4, found (calc.) C: 49.49 (49.03) H: 6.31 (6.78) N: 13.84 (13.45). 1H-NMR (250 MHz; CDCl3; due to limited rotation around the secondary amide several of the signals were doubled) δ: 1.40 (s, 9H); 2.57 (t, 1H); 2.65 (t, 1H); 3.60-3.32 (m’s, 6H); 4.85 (s, 1H); 4.98 (s, 1H); 5.21 (s, 2H); 5.71 (s, 1H); 7.79-7.25 (m’s, 7H). FAB-MS: 518 (M+1).
EXAMPLE 51

Solid Phase Synthesis of H-[Taeg]₂-[Aega]-[Taeg]₁-Lys-NH₂

The protected PNA was assembled onto a BOC-Lys(CIZ) modified MBHA resin with a substitution of approximately 0.15 mmol/g (determined by quantitative Ninhydrin reaction). Capping of only uncoupled amino groups was carried out before the incorporation of the BOC-Goeh-OH monomer.

Stepwise Assembly of H-[Taeg]₂-[Aega]-[Taeg]₁-Lys-NH₂ (synthetic protocol)

Synthesis was initiated on 102 mg (dry weight) of pre-swollen (overnight in DCM) and neutralized BOC-Lys(CIZ)-MBHA resin. The steps performed were as follows: (1) BOC-deprotection with TFA/DCM (dichloromethane) (1:1, v/v), 1×2 minutes and 1×0.5 h, 3 mL; (2) washing with DCM, 4×20 seconds, 3 mL; washing with DMF, 2×20 seconds, 3 mL; washing with DCM, 2×20 seconds, 3 mL; and drain for 30 seconds; (3) neutralization with DIEA/DCM (1:19 v/v), 2×3 minutes, 3 mL; (4) washing with DCM, 4×20 seconds, 3 mL; and drain for 1 minute; (5) addition of 4 equivalents of disopropyl carbodiimide (0.06 mmol, 9.7 μL) and 4 equivalents of BOC-Lys-OH (0.06 mmol, 24 mg) or Boc-Lys-OPfp (0.06 mmol, 30 mg) dissolved in 0.6 mL of 1:1 (v/v) DCM/DMF (final concentration of monomer 0.1M), the coupling reaction was allowed to proceed for 0.5 h while shaking at room temperature; (6) drain for 20 seconds; (7) washing with DMF, 2×20 seconds and 1×2 minutes, 3 mL; washing with DCM 4×20 seconds, 3 mL; (8) neutralization with DIEA/DCM (1:19 v/v), 2×3 minutes, 3 mL; (9) washing with DCM 4×20 seconds, 3 mL, and drain for 1 minute; (10) Qualitative Kaiser test; (11) blocking of unreacted amino groups by acetylation with Ac₂O/pyridine/DCM (1:1:2, v/v), 1×2 h, 0.5 h, 3 mL; and (12) washing with DCM, 4×20 seconds, 2×2 minutes and 2×20 seconds, 3 mL. Steps 1–12 were repeated until the desired sequence was obtained. All qualitative Kaiser tests were negative (straw-yellow colour with no coloration of the beads) indicating near 100% coupling yield. The PNA oligomer was cleaved and purified by the normal procedure. FAB-MS: 2832.11 [M⁺]+1 (calc. 2832.15)

EXAMPLE 52

Solid Phase Synthesis of H-Taeg-Aeg-[Taeg]₁-Lys-NH₂

(a) Stepwise Assembly of BOC-Taeg-A(Z)aqe-[Taeg]₁-Lys(CIZ)-MBHA Resin.

About 0.3 g of wet BOC-[Taeg]₁-Lys(CIZ)-MBHA resin was placed in a 3 mL SPS reaction vessel. BOC-Taeg-A(Z)aqe-[Taeg]₁-Lys(CIZ)-MBHA resin was assembled by in situ DCC coupling (single) of the A(Z)aqe residue utilizing 0.19M of BOC-A(Z)aqe-OBz together with 0.15 M DCC in 2.5 mL of 50% DMF/CH₂Cl₂ and a single coupling with 0.15 M BOC-Taeg-OPfp in neat CH₂Cl₂ ("Synthetic Protocol 5"). The synthesis was monitored by the quantitative ninhydrin reaction, which showed about 50% incorporation of A(Z)aqe and about 96% incorporation of Taeg.

(b) Cleavage, Purification, and Identification of H-Taeg-Aeg-[Taeg]₁-Lys-NH₂

The protected BOC-Taeg-A(Z)aqe-[Taeg]₁-Lys(CIZ)-BHA resin was treated as described in Example 17(c) to yield about 15.6 mg of crude material upon HF cleavage of 53.1 mg dry H-Taeg-A(Z)aqe-[Taeg]₁-Lys(CIZ)-BHA resin. The main peak at 14.4 minutes accounted for less than 50% of the total absorbance. A 0.5 mg portion of the crude product was purified to give approximately 0.1 mg of H-Taeg-Aeg-Taeg-Lys-NH₂. For (MH⁺), the calculated m/z value was 2816.16 and the measured m/z value was 2816.28.

(c) Synthetic Protocol 5

(1) BOC-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 2.5 mL, 3×1 minute and 1×30 minutes; (2) washing with CH₂Cl₂, 2.5 mL, 6×1 minute; (3) neutralization with DIEA/CH₂Cl₂ (1:19 v/v), 2.5 mL, 3×2 minutes; (4) washing with CH₂Cl₂, 2.5 mL, 6×1 minute; and drain for 1 minute; (5) 2–5 mg sample of PNA-resin was removed and dried for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 0.47 mmol (0.25 g) BOC-A(Z)aqe-OBz dissolved in 1.25 mL of DMF followed by addition of 0.47 mmol (0.1 g) DCC in 1.25 mL of CH₂Cl₂ or 0.36 mmol (0.2 g) BOC-Taeg-OPfp in 2.5 mL of CH₂Cl₂; the coupling reaction was allowed to proceed for a total of 20–24 h while shaking; (7) washing with DMF, 2.5 mL, 1×2 minutes; (8) washing with CH₂Cl₂, 2.5 mL, 4×1 minute; (9) neutralization with DIEA/CH₂Cl₂ (1:19 v/v), 2.5 mL, 2×2 minutes; (10) washing with CH₂Cl₂, 2.5 mL, 6×1 minute; (11) 2–5 mg sample of protected PNA-resin was removed and dried for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 25 mL mixture of acetic anhydride/pyridine/CH₂Cl₂ (1:1:2, v/v), 2 h (except after the last cycle); and (13) washing with CH₂Cl₂, 2.5 mL, 6×1 minute; (14) 2×2–5 mg samples of protected PNA-resin are removed, neutralized with DIEA/CH₂Cl₂ (1:19 v/v) and washed with CH₂Cl₂ for ninhydrin analyses.

EXAMPLE 53

Solid Phase Synthesis of H-[Taeg]₁₂-[Aega]-[Taeg]₁-Lys-NH₂

(a) Stepwise Assembly of BOC-[Taeg]₁₂-[Aega]-[Taeg]₁-Lys(CIZ)-MBHA Resin.

About 0.5 g of wet BOC-[Taeg]₁₂-Lys(CIZ)-MBHA resin was placed in a 5 mL SPS reaction vessel. BOC-[Taeg]₁₂-[Aega]-[Taeg]₁-Lys(CIZ)-MBHA resin was assembled by in situ DCC coupling of both the A(Z)aqe and the Taeg residues utilizing 0.15M to 0.16 of protected PNA monomer (free acid) together with an equivalent amount of DCC in 2 mL neat CH₂Cl₂ ("Synthetic Protocol 6"). The synthesis was monitored by the quantitative ninhydrin reaction which showed a total of about 82% incorporation of A(Z)aqe after coupling three times (the first coupling gave about 50% incorporation; a fourth HOBr-mediated coupling in 50% DMF/CH₂Cl₂ did not increase the total coupling yield significantly) and quantitative incorporation (single couplings) of the Taeg residues.

(b) Cleavage, Purification, and Identification of H-[Taeg]₁₂-Aeg-[Taeg]₁-Lys-NH₂

The protected BOC-[Taeg]₁₂-[Aega]-[Taeg]₁-Lys(CIZ)-BHA resin was treated as described in Example 17(c) to yield about 16.2 mg of crude material upon HF cleavage of 102.5 mg dry H-[Taeg]₁₂-[Aega]-[Taeg]₁-Lys(CIZ)-BHA resin. A small portion of the crude product was purified. For (MH⁺), the calculated m/z value was 2050.85 and the measured m/z value was 2050.90.

(c) Synthetic Protocol 6

(1) BOC-deprotection with TFA/CH₂Cl₂ (1:1, v/v), 2 mL, 3×1 minute and 1×30 minutes; (2) washing with CH₂Cl₂, 2 mL, 6×1 minute; (3) neutralization with DIEA/CH₂Cl₂ (1:19 v/v), 2 mL, 3×2 minutes; (4) washing with CH₂Cl₂, 2 mL, 6×1 minute; and drain for 1 minute; (5) 2–5 mg sample
of PNA-resin was removed and dried for a quantitative ninhydrin analysis to determine the substitution; (6) addition of 0.44 mmol (0.23 g) BOC-A(Z)2-Cl dissolved in 1.5 mL of CH$_2$Cl$_2$ followed by addition of 0.44 mmol (0.09 g) DCC in 0.5 mL of CH$_2$Cl$_2$ or 0.33 mmol (0.13 g) BOC-Taeg-Cl in 1.5 mL of CH$_2$Cl$_2$ followed by addition of 0.33 mmol (0.07 g) DCC in 0.5 mL of CH$_2$Cl$_2$', the coupling reaction was allowed to proceed for a total of 20–24 h with shaking; (7) washing with DMSF, 2 mL, 1×2 minutes; (8) washing with CH$_2$Cl$_2$, 2 mL, 4×1 minute; (9) neutralization with DIEA/CH$_2$Cl$_2$ (1:19, v/v), 2 mL, 2×2 minutes; (10) washing with CH$_2$Cl$_2$, 2 mL, 6×1 minute; (11) 2–5 mg sample of protected PNA-resin was removed and dried for a quantitative ninhydrin analysis to determine the extent of coupling; (12) blocking of unreacted amino groups by acetylation with a 25 mL mixture of acetic anhydride/pyridine/CH$_2$Cl$_2$ (1:12:1, v/v/v) for 2 h (except after the last cycle); (13) washing with CH$_2$Cl$_2$, 2 mL, 6×1 minute; and (14) 2×2–5 mg samples of protected PNA-resin were removed, neutralized with DIEA/CH$_2$Cl$_2$ (1:19, v/v) and washed with CH$_2$Cl$_2$ for ninhydrin analyses.

**EXAMPLE 54**

**Hybridization Experiments**

The PNA oligomer H-T$_{45}$C$_1$C$_7$TCTC-LysNH$_2$ (SEQ ID NO:48) was prepared according to the procedure described in Example 51. Hybridization experiments with this sequence should resolve the issue of orientation, since it is truly asymmetrical. Such experiments should also resolve the issues of pH-dependency of the T$_{m}$ and the stoichiometry of complexes formed.

Hybridization experiments with the PNA oligomer H-T$_{45}$C$_1$C$_7$TCTC-LysNH$_2$ were performed as follows:

<table>
<thead>
<tr>
<th>Row</th>
<th>Hybridized With</th>
<th>pH</th>
<th>T$_{m}$ (°C)</th>
<th>l</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>5'-(dA)$<em>{45}$- (dC)$<em>1$- (dG)$</em>{7}$- (dA)$</em>{5}$ (SEQ ID NO:49)</td>
<td>7.2</td>
<td>55.5</td>
<td>2:1</td>
</tr>
<tr>
<td>2</td>
<td>5'-(dA)$<em>{45}$- (dC)$<em>1$- (dG)$</em>{7}$- (dA)$</em>{5}$ (SEQ ID NO:49)</td>
<td>9.0</td>
<td>26.0</td>
<td>2:1</td>
</tr>
<tr>
<td>3</td>
<td>5'-(dA)$<em>{45}$- (dC)$<em>1$- (dG)$</em>{7}$- (dA)$</em>{5}$ (SEQ ID NO:49)</td>
<td>5.0</td>
<td>88.5</td>
<td>2:1</td>
</tr>
<tr>
<td>4</td>
<td>5'-(dA)$<em>{45}$- (dC)$<em>1$- (dG)$</em>{7}$- (dA)$</em>{5}$ (SEQ ID NO:49)</td>
<td>7.2</td>
<td>38.0</td>
<td>2:1</td>
</tr>
<tr>
<td>5</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (dA)$_{1}$</td>
<td>9.0</td>
<td>31.5</td>
<td>—</td>
</tr>
<tr>
<td>6</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (dA)$_{1}$</td>
<td>5.0</td>
<td>52.5</td>
<td>2:1</td>
</tr>
<tr>
<td>7</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:51)</td>
<td>9.0</td>
<td>19.0</td>
<td>2:1</td>
</tr>
<tr>
<td>8</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:51)</td>
<td>5.0</td>
<td>21.5</td>
<td>—</td>
</tr>
<tr>
<td>9</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:51)</td>
<td>10</td>
<td>29.0</td>
<td>2:1</td>
</tr>
<tr>
<td>10</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:52)</td>
<td>7.2</td>
<td>31.5</td>
<td>—</td>
</tr>
<tr>
<td>11</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:53)</td>
<td>7.2</td>
<td>50.5</td>
<td>—</td>
</tr>
<tr>
<td>12</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:53)</td>
<td>7.2</td>
<td>24.5</td>
<td>—</td>
</tr>
<tr>
<td>13</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:54)</td>
<td>9.0</td>
<td>20.5</td>
<td>—</td>
</tr>
<tr>
<td>14</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:54)</td>
<td>5.0</td>
<td>57.0</td>
<td>—</td>
</tr>
<tr>
<td>15</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:54)</td>
<td>16</td>
<td>25.0</td>
<td>—</td>
</tr>
<tr>
<td>16</td>
<td>5'-(dG)$<em>{45}$- (dA)$</em>{1}$- (dC)$<em>{7}$- (dG)$</em>{5}$ (SEQ ID NO:54)</td>
<td>4.0</td>
<td>39.5</td>
<td>—</td>
</tr>
</tbody>
</table>

l = stoichiometry determined by UV-mixing curves
— = not determined

These results show that a truly mixed sequence gave rise to well-defined melting curves. The PNA oligomers can actually bind in both orientations (compare row 1 and 4), although there is a preference for the N-terminal/3' orientation. Introducing a single mismatch opposite a T or C caused a lowering of T$_{m}$ by more than 16° C. at pH 7.2; at pH 5.0 the T$_{m}$ was lowered more than 27° C. This shows that there is a very high degree a sequence-selectivity which should be a general feature for all PNA CIT sequences.

As indicated above, there is a very strong pH-dependency for the T$_{m}$, indicating that Hoogsteen basepairing is important for the formation of hybrids. Therefore, it is not surprising that the stoichiometry was found to be 2:1.

The lack of symmetry in the sequence and the very large decrease in T$_{m}$ when mismatches are present show that the Watson-Crick strand and the Hoogsteen strand are parallel when bound to complementary DNA. This is true for both the orientations, i.e., 5'N-terminal and 3'N-terminal.

**EXAMPLE 55**

T$_{m}$ of PNA Oligomers

The results of hybridization experiments with H-T$_{46}$G7LysNH$_2$ were as follows:

<table>
<thead>
<tr>
<th>Row</th>
<th>Deoxynucleoside</th>
<th>T$_{m}$ (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>5'-(dA)$<em>{45}$- (dA)$</em>{1}$- (dA)$_{5}$ (SEQ ID NO:55)</td>
<td>55.0</td>
</tr>
<tr>
<td>2</td>
<td>5'-(dA)$<em>{45}$- (dG)$</em>{1}$- (dA)$_{5}$ (SEQ ID NO:56)</td>
<td>47.0</td>
</tr>
<tr>
<td>3</td>
<td>5'-(dA)$<em>{45}$- (dG)$</em>{1}$- (dG)$_{5}$ (SEQ ID NO:57)</td>
<td>46.5</td>
</tr>
<tr>
<td>4</td>
<td>5'-(dA)$<em>{45}$-(dT)$</em>{1}$- (dA)$_{5}$ (SEQ ID NO:58)</td>
<td>48.5</td>
</tr>
<tr>
<td>5</td>
<td>5'-(dA)$<em>{45}$-(dT)$</em>{1}$- (dG)$_{5}$ (SEQ ID NO:59)</td>
<td>55.5</td>
</tr>
<tr>
<td>6</td>
<td>5'-(dA)$<em>{45}$-(dT)$</em>{1}$- (dG)$_{5}$ (SEQ ID NO:60)</td>
<td>47.0</td>
</tr>
</tbody>
</table>

As observed by comparing rows 1, 3, and 6 with rows 2, 4, 5, and 7, G can, in this mode, discriminate between C/A and G/T in the DNA strand, i.e., sequence discrimination is observed. The complex in row 3 was furthermore determined to be 2 PNA: 1 DNA RNA by UV-mixing curves.

**EXAMPLE 56**


The protected PNA was assembled onto a BOC-Lys(CIZ) modified MBHA resin with a substitution of approximately 0.145 mmol/g. Capping of only uncoupled amino groups was carried out before the incorporation of the BOC-Aaqg-OH monomer. Synthesis was initiated on 100 mg (dry weight) of neutralised BOC-Lys(CIZ)-MBHA resin that had been preswollen overnight in DCM. The incorporation of the monomers followed the protocol of Example 28, except at step 5 for the incorporation of the BOC-Aaqg-OH monomer. Step 5 for the present synthesis involved addition of 4 equivalents of
diisopropyl carbodiimide (0.06 mM, 9.7 µL) and 4 equivalents of BOC-Aeeg-OH (0.06 mmol, 32 mg) dissolved in 0.6 mL of DCM/DMF (1:1, v/v) (final concentration of monomer 0.1M). The coupling reaction was allowed to proceed for 1×15 minutes and 1×60 minutes (recurring).

All qualitative Kaiser tests were negative (straw-yellow color with no coloration of the beads). The PNA oligomer was cleaved and purified by the standard procedure. FAB-MS average mass found (calc.) (M+H) 4145.1 (4146.1).

EXAMPLE 57

(a) Stepwise Assembly of BOC-[Taeg]2-A(Z)aeeg-Taeg-C(Z)aeeg-A(Z)aeeg-Taeg-C(Z)aeeg-(Lys(CIZ)-MBHA Resin. About 1 g of wet BOC-Lys(CIZ)-MBHA (0.28 mmol Lys/g) resin was placed in a 5 mL SPSS reaction vessel. BOC-[Taeg]2-A(Z)aeeg-Taeg-C(Z)aeeg-A(Z)aeeg-Taeg-C(Z)aeeg-(Lys(CIZ)-MBHA resin was assembled by in situ DCC coupling of the five first residues utilizing 0.16M of BOC-C(Z)OH, BOC-Taeg-OH or BOC-A(Z)aeeg-OH, together with 0.16M DCC in 2 mL of 50% DME/CH2Cl2 and by analogous in situ DCC coupling of the five last residues. Each coupling reaction was allowed to proceed for a total of 20–24 h with shaking. The synthesis was monitored by the ninhydrin reaction, which showed nearly quantitative incorporation of all residues except of the first A(Z)aeeg residue. which had to be coupled twice. The total coupling yield was about 96% (first coupling, about 89% efficiency).


The protected BOC-[Taeg]2-A(Z)aeeg-Taeg-C(Z)aeeg-A(Z)aeeg-Taeg-C(Z)aeeg-Lys(CIZ)-MBHA resin was treated as described in Example 17(c) to yield about 53.4 mg of crude material upon HF cleavage of 166.1 mg of dry BOC-[Taeg]2-A(Z)aeeg-Taeg-C(Z)aeeg-A(Z)aeeg-Taeg-C(Z)aeeg-Lys(CIZ)-MBHA resin. The crude product (53.4 mg) was purified to give 18.3 mg of H-[Taeg]2-Aeeg-Taeg-Caeg-Aeeg-Taeg-Caeg-Lys-NH2. For (M+H)=calculated m/z=2780.17, measured m/z=2780.07.

EXAMPLE 58
Solid Phase Synthesis of H-[Taeg]2-Lys(CIZ)-MBHA Resin

The PNA oligomer was assembled on 500 mg (dry weight) of MBHA resin that had been preswollen overnight in DCM. The resin was initially substituted with approximately 0.15 mmol/g BOC-Lys(CIZ) as determined by quantitative ninhydrin reaction. The stepwise synthesis of the oligomer followed the synthetic protocol described in Example 28 employing 0.077 g (0.2 mmol) BOC-Taeg-OH and 31.3 µL (0.2 mmol) of diisopropylcarbodiimide in 2 mL of 50% DME/CH2Cl2 in each coupling. Capping of uncoupled amino groups was carried out before deprotection in each step. All qualitative Kaiser tests were negative indicating near quantitative coupling yield.

EXAMPLE 59
Synthesis of the Backbone Moiety for Scale-up By Reductive Amination

(a) Preparation of BOC-aminoacetaldehyde. 3-Amino-1,2-propanediol (80 g, 0.88 mol) was dissolved in water (1500 mL) and the solution was cooled to 4°C, after which BOC-anhydride (230 g, 1.05 mol) was added in one portion. The solution was gently heated to room temperature in a water bath. The pH was maintained at 10.5 by the dropwise addition of sodium hydroxide. Over the course of the reaction, a total of 70.2 g of NaOH dissolved in 480 mL of water, was added. After stirring overnight, ethyl acetate (1000 mL) was added, the mixture cooled to 0°C and the pH adjusted to 2.5 by the addition of 4M hydrochloric acid.

The ethyl acetate layer was removed and the acidic aqueous solution was extracted with more ethyl acetate (6–500 mL). The combined ethyl acetate solution was reduced to a volume of 1500 mL using a rotary evaporator. The resulting solution was washed with half saturated potassium hydrogen sulphate (1500 mL) and then with saturated sodium chloride. It then was dried over magnesium sulphate and evaporated to dryness. in vacuo. Yield: 145.3 g (86%).

(b) Preparation of (N'-BOC-aminomethyl)glycine methyl ester

Paladium on carbon (10%, 2.00 g) was added to a solution of BOC-aminoacetalddehyde (10 g, 68.9 mmol) in methanol (150 mL) at 0°C. Sodium acetate (11.3 g, 138 mmol) in methanol (150 mL), and glycine methyl ester hydrochloride (8.65 g; 68.9 mmol) in methanol (75 mL) were added. The mixture was hydrogenated at atmospheric pressure for 2.5 h. The mixture was filtered through celite and evaporated to dryness. in vacuo. The material was redissolved in water (150 mL) and the pH adjusted to 8 with 0.5N NaOH. The aqueous solution was extracted with methylene chloride (5×150 mL). The combined extracts were dried over sodium sulphate and evaporated to dryness, in vacuo. This resulted in 14.1 g (88%) yield of (N'-BOC-aminoethyl)glycine methyl ester. The crude material was purified by kugelrohr distillation at 120° C. and 0.5 mm Hg to give 11.3 g (70%) of a colorless oil. The product had a purity that was higher than the material produced in example 26 according to tlc analysis (10% methanol in methylene chloride).

Alternatively, sodium cyanoborohydride can be used as reducing agent instead of hydrogen (with Pd[C] as catalyst), although the yield (42%) was lower.

(c) Preparation of (N'-BOC-aminoethyl)glycine ethyl ester

The title compound was prepared by the above procedure with glycine ethyl ester hydrochloride substituted for glycine methyl ester hydrochloride. Also, the solvent used was ethanol. The yield was 78%.

EXAMPLE 60
Solid Phase Synthesis of H-Tyr-[Taeg]10-Lys-NH2

(a) Stepwise Assembly of BOC-Tyr(Bz)-[Taeg]10-Lys(CIZ)-MBHA Resin.

About 0.2 g of wet BOC-[Taeg]10-Lys(CIZ)-MBHA resin was placed in a 5 mL SPSS reaction vessel. BOC-Tyr(Bz)-[Taeg]10-Lys(CIZ)-MBHA resin was assembled by standard
in situ DCC coupling utilizing 0.32M of BOC-CTyr(BrZ)-OH together with 0.32M DCC in 3 mL neat CH₂Cl₂, overnight. The ninhydrin reaction showed about 97% incorporation of BOC-Tyr(BrZ).

(b) Cleavage, Purification, and Identification of H-Tyr-[Taeg]₁₀-Lys-NH₂

The protected BOC-Tyr(BrZ)-[Taeg]₁₀-Lys(CIZ)-MBHA resin was treated as described in Example 17(c) to yield about 5.5 mg of crude material upon HF cleavage of 20.7 mg of dry H-Tyr(BrZ)-[Taeg]₁₀-Lys(CIZ)-MBHA resin. The crude product was purified to give 2.5 mg of H-Tyr-[Taeg]₁₀-Lys-NH₂.

EXAMPLE 61

Solid Phase Synthesis of Dansyl-[Taeg]₁₀-Lys-NH₂

(a) Stepwise Assembly of Dansyl-[Taeg]₁₀-Lys(CIZ)-MBHA Resin.

About 0.3 g of wet BOC-[Taeg]₁₀-Lys(CIZ)-MBHA resin was placed in a 5 mL SPPS reaction vessel. Dansyl-[Taeg]₁₀-Lys(CIZ)-MBHA resin was assembled by coupling of 0.5M dansyl-Cl in 2 mL of pyridine, overnight. The ninhydrin reaction showed about 95% incorporation of the dansyl group.

(b) Cleavage, Purification, and Identification of Dansyl-[Taeg]₁₀-Lys-NH₂

The protected dansyl-[Taeg]₁₀-Lys(CIZ)-MBHA resin was treated as described in Example 17(c) to yield about 12 mg of crude material upon HF cleavage of 71.3 mg of dry dansyl-[Taeg]₁₀-Lys(CIZ)-MBHA resin. The crude product was purified to give 5.4 mg of dansyl-[Taeg]₁₀-Lys-NH₂.

EXAMPLE 62

Solid Phase Synthesis of H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-NH₂


About 0.2 g of the above-mentioned MBHA resin was placed in a 5 mL SPPS reaction vessel and neutralized. BOC-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-CIZ)-MBHA resin was assembled by single in situ DCC coupling of the Caeg residue utilizing 0.15M of BOC-Caeg-OH together with 0.15M DCC in 2.5 mL of 50% DMF/CH₂Cl₂ and by coupling the Taeg residues with 0.15M BOC-Caeg-OPfp in 2.5 mL of CH₂Cl₂. Each coupling reaction was allowed to proceed with shaking overnight. The synthesis was monitored by the ninhydrin reaction, which showed close to quantitative incorporation of all the residues.

(b) Cleavage, Purification, and Identification of H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-NH₂

The protected BOC-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-CIZ)-MBHA resin was treated as described in Example 17(c) to yield about 44.4 mg of crude material upon HF cleavage of about 123 mg of dry H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-CIZ)-MBHA resin. Crude product (11 mg) was purified to give 3.6 mg of H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-NH₂.

EXAMPLE 63


(a) Stepwise Assembly of BOC-[Taeg]₁₀-Caeg-[Taeg]₁₀-(CIZ)-MBHA Resin.

About 0.3 g of wet H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys(CIZ)-MBHA resin from the earlier synthesis of BOC-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys(CIZ)-MBHA resin was placed in a 5 mL SPPS reaction vessel. After coupling of the next residue five times, a total incorporation of BOC-C(Z)aeg of 87% was obtained. The five repeated couplings were carried out with 0.18M BOC-C(Z)aeg-OPfp in 2 mL of TFE/CH₂Cl₂ (1.2 v/v), 2 mL of TFE/CH₂Cl₂ (1.2 v/v), 2 mL of TFE/CH₂Cl₂ (1.2 v/v) with two drops of dioxane and two drops of DIEA (this condition gave only a few per cent coupling yield). 2 mL of TFE/CH₂Cl₂ (1.2 v/v) plus 0.5 g phenol, and 1 mL of CH₂Cl₂ plus 0.4 g of phenol, respectively. The two final Taeg residues were incorporated close to quantitatively by double couplings with 0.25M BOC-Taeg-OPfp in 25% phenol/CH₂Cl₂. All couplings were allowed to proceed overnight.


The protected BOC-[Taeg]₁₀-Caeg-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys(CIZ)-MBHA resin was treated as described in Example 17(c) to yield about 7 mg of crude material upon HF cleavage of 80.7 mg of dry H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys(CIZ)-MBHA resin. The crude product was purified to give 1.2 mg of H-[Taeg]₁₀-Caeg-[Taeg]₁₀-Caeg-[Taeg]₁₀-Lys-NH₂ (>99.9% purity).

EXAMPLE 64

Alternative protecting group strategy for PNA synthesis

(a) Synthesis of test compounds: 2-Amino-6-0-benzylpurine: To a solution of 2.5 g (0.109 mol) of sodium in 100 mL of benzyl alcohol was added 10.75 g (0.063 mol) of 2-amino-6-chloropurine. The mixture was stirred for 12 h at 120°C. The solution was cooled to room temperature and neutralized with acetic acid and extracted with 10 portions of 50 mL of 0.2N sodium hydroxide. The collected sodium hydroxide phases were washed with 100 mL of diethyl ether and neutralized with acetic acid, whereby precipitation starts. The solution was cooled to 0°C and the yellow precipitate was collected by filtration. Recrystallization from ethanol gave 14.2 g (92%) of pure white crystals of the target compound. ¹H-NMR (250 MHz, DMSO-d₆): δ: 7.92 (8H); 7.60-7.40 (benzyl aromatic); 6.36 (2-NH₂); 5.57 (benzyl CH₃).

(2-Amino-6-0-benzylpurinyl)methyl ethanoate: A mixture of 5 g (0.0207 mol) of 2-amino-6-o-benzylpurine, 30 mL of DMF and 2.9 g (0.021 mol) of potassium carbonate was stirred at room temperature. Methyl bromoacetate (3.2 g, 1.9 mL 0.0209 mol) was added dropwise. The solution was filtered after 4 h and the solvent was removed under reduced pressure (4 mm Hg, 40°C). The residue was recrystallized two times from ethyl acetate to give 3.7 g (57%) of the target compound. ¹H-NMR (250 MHz, DMSO-d₆): δ: 7.93 (8H); 7.4-7.7 (benzyl aromatic); 6.61 (2-NH₂); 5.03 (benzyl CH₂); 5.59 (CH₃); 3.78 (OCH₃).

(2-N-p-Toluenesulfonylaminido-6-o-benzylpurinyl)methyl ethanoate: To a solution of 0.5 g (1.66 mmol) of 2-amino-6-o-benzylpurinyl)methyl ethanoate in 25 mL of methylene chloride was added 0.53 g (1.62 mmol) of p-toluenesulfonic anhydride and 0.22 g (1.62 mmol) of potassium carbonate. The mixture was stirred at room temperature. The mixture was then filtered and the solvent removed at reduced pressure (15 mm Hg, 40°C). Diethyl ether was added to the oily residue. The resulting solution was stirred overnight, whereby the target compound (0.415 mg, 55%) precipitated and was collected by filtration. ¹H-NMR (250 MHz, DMSO-d₆): δ: 8.97 (8H); 7.2-7.8 (aromatic); 5.01 (benzyl CH₂); 4.24 (CH₃); 3.73 (OCH₃); 2.43 (CH₃).

(b) Stability of the tosyl protected base residue in TFA and HF.

The material was subjected to the standard deprotection conditions (TFA-deprotection) and the final cleavage con-
51

ditions with HF. The products were then subjected to HPLC-
analysis using a 4p RCM 8×100 Nova pack column and
solvents A (0.1% TFA in water) and B (0.1% TFA in
acetonitrile) according to the following time gradient with a
flow of 2 mL/minute.

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>% A</th>
<th>% B</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>100</td>
<td>0</td>
</tr>
<tr>
<td>5</td>
<td>100</td>
<td>0</td>
</tr>
<tr>
<td>35</td>
<td>0</td>
<td>100</td>
</tr>
<tr>
<td>37</td>
<td>0</td>
<td>100</td>
</tr>
<tr>
<td>39</td>
<td>100</td>
<td>0</td>
</tr>
</tbody>
</table>

The following retention times were observed: (a) Compound
1: 30.77 minutes; (b) compound 2: 24.22 minutes; and (c)
compound 3: 11.75 minutes. The analysis showed that the
O2- benzyl group was removed both by TFA and HF, whereas
there was no cleavage of the tosyl group in TFA, but
quantitative removal in HF under the standard cleavage
conditions.

EXAMPLE 65

Synthesis of 5-bromouracil-N2-methyl acetate

5-Bromouracil (5 g, 26.2 mmol) and potassium carbonate
(7.23 g, 52.3 mmol) were suspended in DMF (75 mL). Methyl
bromacetate (2.48 mL, 26.1 mmol) was added over a period
of 5 minutes. The suspension was stirred for 2 h at
room temperature and then filtered. The solid residue was
washed twice with DMF, and the combined filtrates were
evaporated to dryness, in vacuo. The residue was an oil
containing the title compound, DMF, and some unidentified
impurities. It was not necessary to pure the title compound
before hydrolysis. 1H-NMR (DMSO-d6, 250 MHz) δ: 8.55
(improvisy); 8.27 (CBr=N=CH); 8.02 (improvisy); 4.76
(improvisy); 4.70 (improvisy); 4.62 (NCH2OCOCH3); 3.78
(COOC3H3); 2.96 (DMF); 2.80 (DMF). 13C-NMR (DMSO-
d6, 250 MHz) ppm: 168.8 (OOC3H3); 172.5
(CH=C=BrCON); 161.6 (DMF); 151.9 (NCON); 145.0 (CO-
CBr=CH=CH); 95.6 (COOC=CH=CH); 52.6 (improvisy);
52.5 (OCH3); 49.7 (improvisy); 48.8 (NCH2OCOCH3); 43.0
(improvisy); 36.0 (DMF), UV (Meedanol; nm m): 226; 278.
IR (KBr/cm-1): 3158s (N=CH); 1743s (+C=O, COOC3H3); 1701s
(+C=O, CONH); 1418s (CH2, CO2H); 1223s
(+C-O, COOCH3); 864 m (+CH, Br=CH=C-H). FAB-MS m/z
assignment: 265/263 (M+H).

EXAMPLE 66

Synthesis of (5-bromouracil)acetic acid

Water (30 mL) was added to the oil of the crude product
from Example 65 and the mixture was dissolved by adding
sodium hydroxide (2M, 60 mL). After stirring at 0 C. for 10
minutes, hydrochloric acid (4M, 45 mL) was added to adjust
the pH of the solution to 2, and the title compound precipi-
tated. After 50 minutes, the solid residue was isolated by
filtration, washed once with cold water, and dried in vacuo
over silica gel. Yield: 2.46 g (38%). Mp: 250°-251° C. Anal.
for C2H2BrN2O4: Found (calc.): C: 28.78 (28.94); H: 2.00
(2.02); Br: 32.18 (32.09); N: 11.29 (11.25). 1H-NMR
(DMSO-d6, 250 MHz) δ: 12.55 (1H, s, COOH); 11.97 (1H,
s, NH): 8.30 (1H, s, C=C-H); 4.49 (2H, s, NCH2COOH).
13C-NMR (DMSO-d6, 250 MHz) ppm: 169.4 (COOH);
159.8 (NHOCOC3H3); 150.04 (NCON); 145.8 (COOC3-
CH3); 94.6 (COOC3H3); 48.8 (NCH2COOH). V
(Meethanol; mm m): 226; 278. IR (KBr, cm-1): 3207s (+N=CH);
1708s (+C=O, COOCH3); 1687s; 1654s (+C=O, COOCH3).
CONH): 1192s (+C-O, COOH); 842 m (δ CH, Br=C=C-H).
FAB-MS m/z: 251/249 (M+H5).

EXAMPLE 67

Synthesis of N-(BOC-aminoethyl)-N-(5-bromouracil)
methylene-carbononyglycine ethyl ester

BOC-aminoethylglycine ethyl ester (1.8 g, 7.30 mmol)
was dissolved in DMF (10 mL). Dibut-OH (1.31 g, 8.03
mmol) was added, whereby a precipitate was formed. DMF
(2×10 mL) was added until the precipitate was dissolved.
The product of Example 66 (2 g, 8.03 mmol) was added
slowly to avoid precipitation. Methylene chloride (30 mL)
was added, and the mixture was cooled to 0° C. and then
filtered. The precipitate (DCU) was washed twice with
methylene chloride. To the combined filtrate was added
methylene chloride (100 mL). The mixture was washed with
3×100 mL of half-saturated NaHCO3-solution
(H2O:saturated NaHCO3 solution. 1:1. v/v), then with
2×100 mL of dilute KHSO4 solution (H2O: saturated
KHSO4 solution. 1:1, v/v), and finally with saturated NaCl
solution (1×100 mL). The organic phase was dried over
magnesium sulphate, filtered, and evaporated to dryness in
vacuo (about 15 mm Hg and then about 1 mm Hg). The
residue was suspended in methylene chloride (35 mL),
stirred for 45 minutes at room temperature, and the DCU
filtered. Petroleum ether (2 volumes) was added dropwise
to the filtrate at 0° C, whereby an oil precipitated. The liquor
was decanted and the remaining oil dissolved in methylene
chloride (20–50 mL). Precipitated was effected by the addi-
tion of petroleum ether (2 volumes). This procedure was
repeated 5 times until an impurity was removed. The
impurity can be observed by tlc with 10% MeOH/CH2Cl2 as
the developing solvent. The resulting oil was dissolved in
methylene chloride (25 mL) and evaporated to dryness in
vacuo, which caused solidification of the title compound.
Yield: 2.03 g (58.6%). Mp: 87°-89° C. Anal. for
C21H27Br2N2O4. Found (calc.): C: 42.33 (42.78); H: 5.15
(5.28); Br: 17.20 (16.74); N: 1.69 (11.74). 1H-NMR
(DMSO-d6, 250 MHz, J in Hz) δ: 1.93 & 11.92 (1H, s,
C=ONH=CH=O); 8.09 & 8.07 (1H, s, C=CH=H); 7.00 & 6.80
(1H, t, BOC-CH=CO); 4.80 & 4.02 (2H, s, NCH2CONH);
4.35 & 4.24 (2H, s, NCONH=CH=O); 4.27-4.15 (2H, m,
COOHCS=CH2); 3.47-3.43 (3H, m, BOC-NHCH2CH2N=CH3);
3.28-3.25 3.12-3.09 (2H, m, BOC-NHCH2CH2N=CH3);
1.46 & 1.45 (9H, s, t-Bu); 1.26 & 1.32 (3H, t, J=7.1,
COOCH3CH3). 13C-NMR (DMSO-d6, 250 MHz) ppm:
169.3 & 169.0 (t-BuOC=O); 167.4 & 167.1 (COOEOB);
159.8 (C=C=CONH); 155.9 (NCH2CONH); 150.4 (NCON);
145.9 (COOC3H3); 94.5 (COOC3H3); 78.2 (MeC2); 61.3 & 60.7
(COOC3H3); 49.1 & 48.0 (NCH2COOH); 48.0 & 47.0
(NCHOCH); 38.6 (NCONHCH2N); 38.2 (NCONHCH2N); 26.3
(CH2COOCH3); 14.1 (COCH3). UV (Meedanol; NM): 226; 280, IR
(KBr, cm-1): 3207s (+N=CH); 1708s (+C=O, COOH). CONH): 1520s
(+C-O, COOCH3); 1170s (+C-O, COOEt-Bu); 859m
(CH3, Br=C=CH=O). FAB-MS m/z (assignment, relative
intensity): 479/477 (M+H, 5); 432/421 (M+2H-t-Bu, 8);
379/377 (M+2H-Boc, 100); 233/231 (M-2H=bone, 20).

EXAMPLE 68

Synthesis of N-(BOC-aminoethyl)-N-(5-bromouracil-
N2-methylene-carbonylglycine)

The product of Example 67 (1.96 g, 4.11 mmol) was
dissolved in methanol (30 mL) by heating, and then cooled
to 0° C. Sodium hydroxide (2M, 30 mL) was added, and
the mixture stirred for 30 minutes. HCl (1M, 70 mL) was added
53

ph 2). The water phase was extracted with ethyl acetate (3.65 mL) and evaporated to dryness. Yield: 1.77 g (96%).

M.p. 92°-97°C. Anal. for C₆H₅Br₂N₄O₂. Found (calcld.): C: 42.10 (42.36); H: 3.43 (3.55); N: 16.25 (16.47); ¹H-NMR (DMSO-d₆). 250 MHz. J. in Hz: δ: 13.19 (1H, s, COO); 11.41 (1H, s, N); 7.69 (1H, d. J₅,H,C-C=CH₂=7.8, J₅,H,C=CH₂=7.8). COOH = CH₃): 4.49 (2H. s. N-CH₂CO₂H). ¹C-NMR (DMSO-d₆. 250 MHz ppm): 169.9 (COOH); 163.9 (CH=CHCON); 151.1 (NCON). 146.1 (COOH=CH₂); 100.9 (COOH=CH₂). 48.7 (N=CH₂CO₂H). UV (Methanol: nm max: 246; 263. IR (KBr); cm⁻¹: 3122s (_NH); 1706w (s. N=CH₂). 1608w, 1692vs (_C=O). 1325s (_C=O-CH₂). 767 (2 CH₃, C=O). FAB-MS m/z (assignment): 171 (M+H).

EXAMPLE 71

Synthesis of N-(BOC-aminooxy)-N-(uracil-N₁-methylene-carbonyl) glycine ethyl ester

(BOC-aminooxy)glycine ethyl ester (2 g, 8.12 mmol) was dissolved in DMF (10 mL). Dibut-OH (1.46 g, 8.93 mmol) was added and a precipitate was formed. DCM (20 mL) was added until all was dissolved. The product of Example 70 (1.52 g, 8.93 mmol) was added slowly to avoid precipitation. Methylene chloride (30 mL) was added and the mixture was cooled to 0°C, after which DCC (2.01 g, 9.74 mmol) was added. The mixture was stirred for 1 h at 0°C. 2 h at room temperature, and then filtered. The precipitated DCU was washed twice with methylene chloride. To combined filtrates was added methylene chloride (100 mL), and the solution was washed with 3×100 mL of half-saturated NaHCO₃ solution (H₂O: saturated NaHCO₃ solution. 1:1. v/v), then with 2×100 mL of dilute KH₂SO₄ solution (H₂O: saturated KH₂SO₄ solution. 4:1. v/v) and finally with saturated NaCl solution (1×100 mL). The organic phase was dried over magnesium sulphate, filtered and evaporated to dryness in vacuo (about 15 mm Hg and then about 1 mm Hg). The residue was suspended in methylene chloride (32 mL), and stirred for 35 minutes at room temperature, and 30 minutes at 0°C and then filtered. The precipitate (DCU) was washed with methylene chloride. Petroleum ether (2 volumes) was added dropwise to the combined filtrate at 0°C, which caused separation of an oil. The mixture was decanted, the remaining oil was then dissolved in methylene chloride (20 mL), and then again precipitated by addition of petroleum ether (2 volumes). This procedure was repeated 5 times until an impurity was removed. The impurity can be seen by TLC with 10% MeOH/CH₂Cl₂ as the developing solvent. The resulting oil was dissolved in methylene chloride (20 mL) and evaporated to dryness in vacuo, which caused solidification of the title compound. Yield: 1.71 g (53%). M.p. 68.5-75.7°C. Anal. for C₁₂H₁₄N₂O₄. Found (calcld.): C: 50.61 (51.25); H: 6.48 (6.58); N: 13.33 (14.06). ¹H-NMR (DMSO-d₆. 250 MHz. J. in Hz): δ: 11.36 (1H, s, C=ON=O), 7.51 (4H, d. J₅,H,C-C=CH₂=7.4, J₅,H,C-C=CH₂=7.4). COOH = CH₂): 4.13 (2H, s. N=CH₂CO₂H). ¹C-NMR (DMSO-d₆. 250 MHz ppm): 168.8 (COOMe); 164.0 (C=O=CH₂); 151.1 (NCON). 146.1 (CO=CH₂); 101.3 (CO=CH₂). 52.5 (COOH). 48.7 (N=CH₂CO₂H). UV (Methanol: nm max: 226; 226. IR (KBr; cm⁻¹): 3164s (_NH); 1748vs (_C=O). COOMe); 1733vs (_C=O, CONH); 1455vs (a CH₃, CH₂O); 1243VS (_C=O,COOMe); 701m (d CH, H=C=CH₂). FAB-MS m/z: 185 (M+H).

EXAMPLE 70

Synthesis of uracilcarboxylic acid

Water (90 mL) was added to the product of Example 69 (8.76 g, 47.5 mmol), followed by 30 mL of 95% hydroxide (2M. 40 mL). The mixture was heated for 40 minutes, until all the methyl ester has reacted. After stirring at 0°C for 15 minutes, hydrochloric acid (4M. 25 mL) was added (pH 2). The title compound precipitated and the mixture was filtered after 2-3 h. The precipitate was washed once with the mother liquor and twice with cold water and dried in vacuo over isopentyl. Yield: 6.65 g (82%). M.p. 288°-289°C. Anal.
55

(5-C-O, COOEt); 1172s (5-C-O, COOEt-Bu); 718w (s CH-
5-C-C-H). FAB-MS m/z (assignment, relative intensity): 399 (M+H+ 35); 343 (M+2H+ → Bu, 100); 299 (M+2H+ →
BOC, 103); 153 (M−backbone, 30).

EXAMPLE 72

Synthesis of N-(BOC-aminomethyl)-N-(uracilimethylene-
carbonyl)glycine

The product of Example 71 (1.56 g, 3.91 mmol) was
56
dissolved in methanol (20 mL) and then cooled to 0°C.
Hydrochloric acid (4M, 46 mL) was added (pH 2).

The water phase was extracted with ethyl acetate (3×50
mL, 47×30 mL). The combined ethyl acetate extracts
60
were washed with saturated NaCl solution (360 mL).
The organic phase was dried over magnesium sulfate,
filtered, and evaporated to dryness, in vacuo. The residue
was dissolved in methanol and evaporated to dryness, in vacuo. Yield: 0.55
65
g (38%). Mp 164°-170°C. Anal. for C14H22N2O4: Found:
68.6 68.5 (C); 53.9 (H); N: 1461 (15.13).

H-NMR (DMSO-d6, 250 MHz, J in Hz): δ: 12.83 (1H, s, 
NCOOH); 11.36 (1H, s, C=ONH=O); 7.52-7.45 (1H, m, 
COOH→CHN); 7.00 & 6.82 (1H, t, BOC-NH); 5.67-5.62
69
(1H, m, COOH→C=O); 4.76 & 4.58 (2H, s, NCH2CON); 
4.26 & 4.05 (2H, s, NCH2COOH); 3.46-3.39 (2H, m, 
BOCNCCH2CH2N); 3.25-3.23 & 3.15-3.09 (2H, m, 
BOCNCCH2CH2N); 1.46 (9H, s, t-Bu).

Example 73

Synthesis of H-U10-LysNH2

Synthesis of the title compound was accomplished by
75
using the following protocol: (1) BOC-deprotection with
TFA/CH2Cl2 (1:1, v/v): 3×1 minute and 1×30 minutes; (2)
washing with CH2Cl2, 6×1 minute; (3) neutralization with
DIEA/CH2Cl2 (1:19, v/v); 3×2 minutes; (4) washing with 
CH2Cl2, 6×1 minute, and drain for 1 minute; (5) at some
70
stages of the synthesis, 2-5 mg sample of PNA-resin was
removed and dried for a ninhydrin analysis to determine the
substitution; (6) addition of BOC-protected PNA monomer
(free acid) in DMF followed by addition of DCC in CH2Cl2;
the coupling reaction was allowed to proceed for a total of
24 h with shaking; (7) washing with DMF, 1×2 minutes; (8)
washing with CH2Cl2, 4×1 minute; (9) neutralization with
DIEA/CH2Cl2 (1:19, v/v), 2×2 minutes; (10) washing with
80
CH2Cl2, 6×1 minute; (11) occasionally, 2-5 mg sample of
protected PNA-resin was removed and dried for a ninhydrin
analysis to determine the extent of coupling; (12) at some
stages of the synthesis, unreacted amino groups were
blocked by acetylation with a mixture of acetic anhydride/
pyridine/CH2Cl2 (1:1:2, v/v/v) for 2 h followed by washing
with CH2Cl2, 6×1 minute, and, occasionally, ninhydrin
85
analysis.

The synthesis was initiated on approximately 100 mg of
Lys(CIZ)-MBHA-resin. The crude product (12 mg) was

pure enough for hybridization studies. The hybrid between
80
5'-3'-MU10 and H-U10 had Tm of 67.5°C.

EXAMPLE 74

Enhanced Stability of Duplex Between PNA Containing
2.6-Diaminopurine and DNA

Four PNA decamers (H-GTATAGCTACT-LysNH2,
H-GTATGCTACT-LysNH2, H-GTATGCTACT-LysNH2
and H-GTATGCTACT-LysNH2, where D is 2.6-
diaminopurine) were synthesized according to procedures
described above. The PNA decamers were allowed to
hybridize with complementary DNA and the thermal
melt profiles of the PNA:DNA duplexes were measured and
compared to control DNA:DNA duplexes and with
PNA:DNA duplexes wherein the DNA contains a single
85
T=C mismatch opposite a D residue in the PNA. The results
are shown in the table below.

<table>
<thead>
<tr>
<th>PNA</th>
<th>DNA</th>
<th>Tm (°C) (DNA)</th>
<th>Tm (°C) (PNA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DNA-1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DNA-2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DNA-3</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*DNA-1 is 5'-JATAGCTACT-3' (SEQ ID NO:7)
*DNA-2 is 5'-JATAGCTACT-3' (SEQ ID NO:8)
*DNA-3 is H-GTATGCTACT-LysNH2 (SEQ ID NO:9)
**ND = Not Determined

The presence of 2.6-diaminopurine increased the Tm by
4°-5°C in PNA:DNA duplexes whereas the increase in
DNA:DNA duplexes was only 3°-4°C. This difference may
reflect increased stacking in the former duplex. This
tendency is also observed when unmodified homoduplexes of
PNA:DNA and DNA:DNA are compared to duplexes having
different D:T base pairs, resulting in Tm increases of 12°
and 10.5°C, respectively. Upon comparison of the binding of
PNA-3 to DNA-1 and DNA-2, a decrease in Tm of 23.5°C
is observed when a C mismatch is introduced into the
PNA:DNA duplex containing only one 2.6-diaminopurine
nucleoside, whereas a decrease in Tm of only 18°C is
observed in case of PNA-2 which does not contain any
2.6-diaminopurine nucleobases. The increased
specificity in terms of a decrease in the melting temperature (Tm) of
the duplex was not observed when additional D:T basepairs were
introduced. This finding reflects the increased stability of the
duplex containing more than a single 2.6-
diaminopurine nucleobase indicating that the more stable
duplex is better able to accommodate the mismatch-induced
structural change in the duplex.

EXAMPLE 75

Binding of PNA Containing 2.6-Diaminopurine to DNA

A PNA decamer containing six 2.6-diaminopurine nucleo-
bases was allowed to hybrdize to complementary DNA. The
thermal stability of this duplex was determined and compared to thermal stability of a duplex formed between DNA and a PNA decamer devoid of 2,6-diaminopurine nucleobases. The Tm results are shown in the table below.

<table>
<thead>
<tr>
<th></th>
<th>*DNA-6 (Tm in °C)</th>
<th>*DNA-7 (Tm in °C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PNA-5</td>
<td>71</td>
<td>50</td>
</tr>
<tr>
<td>H-AAAAGCGAGGAG-3' (SEQ ID NO:10)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PNA-6</td>
<td>≥85</td>
<td>71</td>
</tr>
<tr>
<td>H-DDDDGGDGDDG-3' (SEQ ID NO:11)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*DNA-6 is 5'-CTCTCCCTTTT-3' (SEQ ID NO:12)
DNA-7 is 5'-TTTCTCCCTTC-3' (SEQ ID NO:13)

The incorporation of 2,6-diaminopurine nucleobases into PNA increases thermal stability (represented by the Tm) of the duplex between DNA and the PNA containing a 2,6-diaminopurine nucleobase to 85°C when compared with the thermal stability (Tm = 71°C) of the duplex between DNA and a PNA devoid of 2,6-diaminopurine nucleobases.

**EXAMPLE 76**

Synthesis of ethyl-N6-(benzoyloxy-carbonyl)-2,6-diaminopurin-9-yl-acetate (14, FIG. 7a)

To a suspension of 2.6-diaminopurine (3 g, 19.46 mmol) in dry DMF (90 mL) was added NaH (60% in oil, 0.87 g, 21.75 mmol). After 1 hour ethyl bromoacetate (4.23 g, 25.34 mmol) was added. The reaction mixture became homogeneous in 30 minutes and was allowed to stir for an additional 90 minutes. The DMF was removed in vacuo resulting in a tan powder. The tan powder was then reflushed with 1.4-dioxane (200 mL) for 10 minutes and filtered through celite.

The solution was concentrated to give a light yellow powder. To the light yellow powder (5.52 g) in 1.4-dioxane (150 mL) was added freshly prepared N-benzoyloxy-carbonyl-N'-methylimidazolium triflate (10.7 g, 29.2 mmol). The reaction mixture was stirred at room temperature for 16 h resulting in a reddish solution. The dioxane was removed in vacuo and the crude material was recrystallized from MeOH-H2O ether to give 4.56 g (63%) of the title compound as a cream-colored solid.

H NMR (DMSO-d6) δ: 10.12 (bs, 1H), 7.43 (m, 5H), 6.40 (bs, 2H), 5.17 (s, 2H), 4.94 (s, 2H), 4.18 (q, J = 7.2, 2H), 1.21 (t, J = 7.2, 2H), 1.33 (s, 3H), 0.15 (s, 3H), 0.14 (s, 3H).

**EXAMPLE 77**

Synthesis of N6-(benzoyloxy-carbonyl)-2,6-diaminopurin-9-yl-acetic acid (15, FIG. 7a)

Ethyl-N6-(benzoyloxy-carbonyl)-2,6-diaminopurin-9-yl-acetate (14, 3 g, 8.11 mmol) was dissolved in NaOH (2N, 30 mL). After 1 h the solution was acidified to pH 2.5 with 2M HCl. The precipitate was filtered, washed with water, and dried to give 2.82 g (98%) of the title compound as a white solid.

IR (KBr): 3300, 3095, 1750, 1630, 1590, 1410. H NMR (DMSO-d6) δ: 10.11 (s, 1H), 7.91 (s, 1H), 7.45-7.33 (m, 5H), 6.40 (s, 2H), 5.17 (s, 2H), 4.83 (s, 2H).

**EXAMPLE 78**

Synthesis of BOC-aaminocetadlyde (16, FIG. 7b)

The title compound was prepared according to a published literature procedure (Daeholm et al., Organic Preparations and Procedures Intl., 1993, 25, 457).
saturated potassium hydrogen sulfate (that was half-diluted with water), separated and concentrated. The crude material was purified by silica gel flash column chromatography using chloroform:methanol (9:1, v/v), as the eluent. Concentration and drying of the appropriate fractions gave 1.25 g (42%) of the title compound.

EXAMPLE 83

Standard Protocol For PNA Synthesis and Characterization

Instrument: PerSeptive Biosystems 8909 Expedite.
Synthesis Scale: 2 μmole.
Reagents:
Wash A: 20% DMSO in NMP
Wash B: 2M Collidine in 20% DMSO in NMP
Deblock: 5% m-Cresol, 95% TFA
Neutralizer: 1M DIA in 20% DMSO in NMP
Cap: 0.5M Acetic Anhydride. 1.5M Collidine in 20% DMSO in NMP
Activator: 0.2M HATU in DMP
Monomers: 0.22M in 2M Collidine (50% Pyridine in DMP)

Synthesis: The solid support (BOC-BHA-PEG-resin) is washed with 708 μL of Wash A. Deblock (177 μL) is passed through the column 3 times over 6.3 minutes. The resin is then washed with 1416 μL of Wash A. The free amine is neutralized with 1063 μL of Neutralizer. The resin is washed with 1062 L of Wash B. Monomer and Activator (141 μL each) are slowly added over the column over 14 minutes. The resin is washed with 708 μL of Wash B and 708 μL of Wash A. Uncapped amine is capped with slow addition of 708 μL of Cap solution over 5 minutes. The resin is then washed 2124 μL of Wash A. The cycle is repeated until synthesis of the desired PNA sequence is completed.

Cleavage: The PNA-resin is washed with 5 mL of MeOH and 5 mL of Acetic acid under vacuum. The dried resin is emptied into a 1.5 mL Durapore ultrafiltration unit. Thioanisole (25 μL), 50 μL of m-Cresol, 100 μL of TFA and 150 μL of TFMSA is added to the resin, vortexed for about 30 seconds and allowed to stand for 2 h. The mixture is then centrifuged for 5 minutes at 10K and the inner tube with resin is removed. Approximately 1.5 mL of ether is added to the TFMSA solution to precipitate the product. The TFA solution is vortexed, followed by centrifugation at 10K for 2 minutes. The ether is removed in vacuo. Ether precipitation and centrifugation are repeated an additional 2 times. The dry pellet is heated in a heat block (55°C) for 15 to 30 minutes to remove excess ether and redissolved in 200 μL of H2O. Solvent is added to 100 mg of Dowex Acetate Resin in a 1.5 mL Durapore ultrafiltration unit, vortexed, allowed to stand for 30 minutes and centrifuged at 10K for 2 minutes.

Characterization: The absorbance of a 1 μL sample in 1 mL of H2O is measured at 260 nm. Isopropanol (50%) in H2O with 1% Acetic acid (100 μL) is added to 4 μL of the sample. This sample is characterized by electrospray mass spectrometry.

Common Abbreviations
NMP: N-methyl pyrrolidinone
TFA: Trifluoroacetic acid
DIEA: N,N-Diisopropylethylamine
HATU: O-(7-azabenzo[d]azol-1-yl)-1-1,3,3-trimethyluronium hexafluorophosphate
TFMSA: Trifluormethanesulfonic Acid

EXAMPLE 84

PNA Oligomers Containing 2,6-Diaminopurine Attached to Aminoethyl-Lysine Backbone

Using the title compound of Example 82, the aminoethyl-glycine PNA monomers of examples 24 through 34, and the standard protocol for PNA synthesis illustrated in Example 83, the following PNA oligomers were prepared:

<table>
<thead>
<tr>
<th>SEQ ID NO:</th>
<th>14</th>
<th>15</th>
</tr>
</thead>
<tbody>
<tr>
<td>TTTCG-GGAC-CCGC</td>
<td>GCDK-DMAE-GC</td>
<td></td>
</tr>
</tbody>
</table>

C. G. and T are nucleobases cytosine, guanine, and thymine respectively, attached to an aminoethyl-glycine PNA backbone. Dk is a 2,6-diaminopurine attached to an aminoethyl-lysine backbone as illustrated in the previous examples. Aminoethyl-lysine backbone is an aminoethyl-glycine backbone with butylamine substituent at α-position, i.e., lysine side-chain.

EXAMPLE 85

Synthesis of PNA Oligomers Having At Least One A. G. C, or T Attached To A Lysine-containing Backbone

Using the procedures of Example 83, the aminoethylglycine PNA monomers of examples 24 through 34, and monomers of Examples 76–82, the following PNA oligomers were synthesized (Tk is thymine attached to an aminoethyl-lysine backbone; Gk is guanine attached to an aminoethyl-lysine backbone; Ck is cytosine attached to an aminoethyl-lysine backbone):

| SEQ ID NO: | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 | 34 | 35 | 36 | 37 | 38 | 39 | 40 | 41 | 42 | 43 | 44 |
|------------|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|

EXAMPLE 86

Synthesis of methyl α-formylsuccinate

This procedure is a modification of published method (Fisseler et al., Biochemistry, 1970, 9, 3136). Sodium methoxide (40.5 g, 0.75 mol) was suspended in dry ether (500 mL) and stirred under nitrogen at 0°C. A mixture of dimethylsuccinate (65.4 mL, 0.5 mol) and methylformate (123 mL, 2 mol) was added dropwise over 30 minutes. The
reaction mixture was stirred at 0°C for 2 h and then at room temperature overnight. Subsequently, the reaction mixture was evaporated to a viscous brown residue which was washed once with petroleum ether and then dissolved in 3M hydrochloric acid (160 mL). This solution was made weakly acidic with concentrated hydrochloric acid and then extracted with dichloromethane (4×250 mL). The organic phase was dried (MgSO₄), filtered and evaporated under reduced pressure. The resulting residue was distilled in a kugelrohr apparatus at 60°C and 0.6 mBar yielding 52.3 g of a mixture of the title compound and dimethyl succinate in the molar ratio 80:20 (determined by NMR) as a colorless oil. The product is purified of the dimethyl succinate by continuous extraction with diethyl ether. Alternatively the mixture can be used directly for the next step.

1H NMR (DMSO-d₆, TMS): δ: 3.2 (s, 2H, CH₂), 3.59 (s, 3H, OMe), 3.61 (s, 3H, OMe), 7.73 (s, 1H, CHO) 10.86 (br s, 1H, CHO). 13C NMR (DMSO-d₆, TMS) ppm: 28.9 (CH₂), 51.0 (OMe), 51.6 (OMe), 102.1 (C=CH-OH), 136.6 (CHO), 168.3 (COO), 171.7 (COO).

EXAMPLE 87

Synthesis of pseudoisocytosine-5-ylacetic acid

This procedure is a modification of a published method (Beran et al., Collect. Czech. Chem. Commun., 1983, 48, 292). Sodium methoxide (41.9 g, 0.78 mol) was dissolved in dry methanol (200 mL) and guanidine hydrochloride (49.4 g, 0.52 mol) was added. The mixture was stirred for 10 minutes under nitrogen at room temperature. A solution of methyl triformylsuccinate (30 g, 0.17 mol) in dry methanol (100 mL) was added to the mixture. The reaction mixture was refluxed under nitrogen for 3 h and then stirred at room temperature overnight. The reaction mixture was filtered, and the filtered residue washed once with methanol. The collected filtrate and washing were evaporated under reduced pressure. The resulting residue was dissolved in water (80 mL) and the solution was acidified with concentrated hydrochloric acid to pH 4.2. After having been stirred at 0°C the mixture was filtered, the precipitate washed once with water and then freeze-dried leaving 28.29 g (97%) of the title compound as a white solid.

Anal. Calcd for C₂₂H₂₃N₅O₅: C, 40.45; H, 4.53; N, 23.59. Found: C, 40.13; H, 4.22; N, 23.26. Due to the poor solubility properties of the product it was further characterized as its sodium salt. The title compound (0.42 g, 2.5 mmol) and sodium bicarbonate were dissolved in boiling water (35 mL). The solution was cooled and evaporated. The residue was dissolved in water (6 mL) and ethanol (4 mL) and isopropanol (8 mL) were added. The sodium salt was collected by filtration, washed with absolute ethanol and petroleum ether and dried to yield 0.31 g of the product (65%) as white crystals.

1H NMR (D₂O, TMS): δ: 3.10 (s, 2H, CH₂COO), 7.40 (s, 1H, H5). 13C NMR (DMSO-d₆, TMS) ppm: 34.8 (CHOOC), 112.0 (C-5), 145.6-146.5 (m, C-2), 155.1 (C-6), 169.4 (C-4), 179.3 (COOH). MS (FAB) m/z (%): 192 (100, M+H).
chlorobenzyloxy) allyl ester (18) as per the procedure of Example 81. The resulting monomeric compound is treated as per the procedure of Example 82 to give the deprotected compound ready for use in oligomer synthesis.

EXAMPLE 92

Synthesis of PNA Oligomer Having a Pseudoisocytosine Attached To an Aminoethyl-lysine Backbone

Aminoethyl-lysine pseudoisocytosine monomer was incorporated into PNAs using the procedure of Example 83.

EXAMPLE 93

Preparation of PNA Monomers Having Adenine, Guanine, Cytosine, and Thymine Attached To an Aminoethyl-lysine Backbone.

a) Preparation of the guanine monomer: To N6-benzyl-9-carboxymethylene-guanine (2.63 g, 8.78 mmol) was added DIEA (2.6 mL, 20 mmol), DMF (30 mL), dichloromethane (70 mL), and N-(BOC-aminoethyl)-lysine-(2-chlorobenzyloxy) allyl ester (18, 3.7 g, 8.04 mmol). The reaction mixture was stirred under nitrogen for 20 minutes. PyBrop (4 g, 8.58 mmol) was added and the reaction mixture stirred for an additional 16 h. The reaction mixture was concentrated and the residue was purified by silica gel flash column chromatography using chloroform/hexanes/methanol (12:7:1, v/v/v) to give 4 g (60%) of the title compound as the allyl ester.

To the allyl ester (4 g, 5.37 mmol) was added THF (100 mL), tetrakis palladium (0.18 g, 0.15 mmol), and morpholine (6.1 g, 70 mmol). The reaction mixture was stirred under nitrogen for 2.5 h and concentrated. The residue was purified by silica gel flash column chromatography using chloroform/hexanes/methanol (11:8:1, v/v/v) to give 2.67 g (60%) of the title compound.

b) Preparation of the adenine monomer: The procedure used for the guanine monomer in Example 93(a) above was followed for the synthesis of the adenine monomer using N6-benzyl-9-carboxymethylene-adenine.

c) Preparation of the cytosine monomer: To N-(BOC-aminoethyl)-lysine-(2-chlorobenzyloxy) allyl ester (18, 8.21 g, 17.7 mmol), added triethylamine (10 mL, 98 mmol) and dichloromethane (200 mL). The solution was cooled to about 0°C in an ice bath under nitrogen. To the cooled solution was added chloroacetyl chloride (2.2 mL, 27.6 mmol) over 10 minutes and the reaction mixture stirred at room temperature for 16 h. The reaction mixture was concentrated and the residue was purified by silica gel flash column chromatography using ethyl acetate/hexanes (1:1, v/v) to give 6.54 g (68%) of the N-acetylated lysine backbone.

Cytosine is protected at the N-4 position by treatment with benzyl chlorofomate in pyridine at 0°C. To give N4-benzyl-cytosine.

To N4-benzyl-cytosine (1.31 g, 5.34 mmol) was added DMF (200 mL), and 60% NaOH in mineral oil (0.22 g, 5.4 mmol) and the resulting mixture was stirred under nitrogen for 30 minutes. To the resulting mixture was added the N-acetylated lysine backbone (2.9 g, 5.34 mmol) in DMF (25 mL) and the mixture stirred for 16 h. The reaction mixture was concentrated and the residue dissolved in dichloromethane (250 mL). The dichloromethane phase was washed with water (200 mL) and concentrated. The resulting residue was purified by silica gel flash column chromatography using dichloromethane/hexanes: methanol (8:2:1) to give 2.4 g (85%) of the cytosine attached to the aminomethyl-lysine backbone as the allyl ester.

The allyl ester is converted to the active monomer by deprotection using pivaloyl following the procedure used in Example 93(a) above to give 1.05 g (46%) of the title compound.

Also see Examples 86-91.

d) Preparation of the thymine monomer: The thymine monomer was prepared following the procedure of Example 93(c) above.

EXAMPLE 94

In Vitro Evaluation of PNAs Targeted to HCV

HCV replication in cell culture has not yet been achieved. Consequently, in vitro translation assays are used as standard assays to evaluate test compounds for their anti-HCV activity. One such standard in vitro translation assay was used to evaluate PNAs of the present invention for their ability to inhibit synthesis of HCV protein in a rabbit reticulocyte assay.

Plasmids containing full-length cDNA sequence for the desired portion of the HCV mRNA was prepared. A T7 promoter was introduced into the plasmid immediately adjacent to the 5' cap site. A similar strategy was used for maintaining a control in which a cDNA plasmid containing coding sequences for a truncated intercellular adhesion molecule type I was modified. As a result of a deletion at base 554 relative to the ICAM-1 AUG, a frameshift occurs with a stop codon generated at base 679. The resulting open reading frame encodes a truncated ICAM-1 polypeptide with a lower molecular weight.

Uncapped transcripts for in vitro translation were prepared by T7 transcription of the plasmid using the Megascript transcription kit (Ambion, Inc.) according to the instructions provided by the manufacturer. The plasmid was linearized by restriction endonuclease digestion at a site in the linker region of the plasmid immediately downstream of the 3'-untranslated sequences of the cDNA insert in order to generate a transcript nearly identical in sequence to authentic mRNA. Following transcription, free nucleotides were removed using G-50 Quickspin columns (Boehringer-Mannheim) and the amount of transcript present was quantitated by optical density.

In vitro translation reactions contained 300 ng of the HCV transcript (final concentration of 10 nM), 7 μL of rabbit reticulocyte lysate (RRL, Promega), 8.8 μCi of [35S]-methionine (1175 Ci/mmol, Amersham), 13 μM IVT amino acids mix devoid of methionine (Promega), 8 units of RNasin (Promega) and PNAs in a total volume of 15 μL. A similar control reaction contained 100 ng (30 nM) of the truncated ICAM-1 transcript instead of the HCV transcript. The target and control RNA were heated at 65°C for 5 minutes, incubated at 37°C for 15 minutes and then mixed with lysate components. The translation mix was incubated at 37°C for 60 minutes and the reaction was terminated by the addition of 2x Laemmli gel loading buffer. After boiling, proteins were fractionated on precast 14% acrylamide gels (Novex, San Diego), fixed in 10% propanol, 5% acetic acid, 3% glycerol, dried and analyzed with a PhosphorImager.

PNAs effectively blocked in vitro translation of HCV protein. PNAs that were evaluated in an in vitro translation assay are shown in the table below (Dk and Tk are 2.6-diaminopurine and thymine, respectively, attached to an aminomethyl-lysine backbone).
Results of the in vitro translation assay are shown in FIG. 8. It is observed that 13642 (which is a 12-mer PNA containing two 2,6-diaminopurine nucleobases bearing lysine side chains) with an $E_C^30$ of approximately 29 nM is more effective at blocking in vitro translation of HCV protein than 265-12 (which is devoid of lysine side chains) with an $E_C^30$ of approximately 57 nM. Further, upon comparing 11908 and 13639, at a concentration as low as 30 nM, it is evident that the PNA with lysine side chains (i.e. 13639) is more effective at blocking in vitro translation of HCV protein than 11908, which does not contain any lysine side chains. This clearly indicates that the presence of a side chain in PNA enhances its ability to block in vitro translation of HCV protein.

EXAMPLE 95

Thermal Stability of PNA Duplexes

Duplex-forming PNAS were synthesized as described in Example 85. PNA having the sequence H-GTAX-GATx-CAC-Tx-R (SEQ ID NO:1, wherein Tx represents a thymine monomer bearing an amino acid side chain) was allowed to hybridize with complementary DNA having the sequence 5'-AGT-GAT-CTA-C-3' (SEQ ID NO:7) and complementary PNA having the sequence H-AGT-GAT-GTA-CTA-C-1-LysNH$_2$ (SEQ ID NO:9), and the thermal stabilities ($T_m$) of the duplexes were determined in 10 mM phosphate, 100 mM NaCl and 1 mM EDTA at a pH of 7. The results are shown in the table below.

<table>
<thead>
<tr>
<th>SEQ IS #</th>
<th>ID No.</th>
<th>SEQUENCE</th>
</tr>
</thead>
<tbody>
<tr>
<td>13642</td>
<td>14</td>
<td>TTT-GGC-CDC-CDCk</td>
</tr>
<tr>
<td>13414</td>
<td>19</td>
<td>TKTY-TGG-AGG-AATK-CAT-C-3'</td>
</tr>
<tr>
<td>13639</td>
<td>20</td>
<td>TGG-TGG-TAC-TCA-TGG</td>
</tr>
<tr>
<td>265-12</td>
<td>45</td>
<td>TTT-GGC-GAC-CCA</td>
</tr>
<tr>
<td>11908</td>
<td>46</td>
<td>TCG-TGG-CGG</td>
</tr>
<tr>
<td>8215</td>
<td>47</td>
<td>Gly-TTT-AGG-ATT-COT-GCT-CAT-GT-LysNH$_2$</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>X*</th>
<th>C-Terminal</th>
<th>$T_m$ (°C)</th>
<th>$T_m$ (°C)</th>
<th>$T_m$ (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glycine</td>
<td>a*</td>
<td>52</td>
<td>68</td>
<td>38</td>
</tr>
<tr>
<td>Glycine</td>
<td>b'</td>
<td>49</td>
<td>67</td>
<td>38</td>
</tr>
<tr>
<td>L-Lysine</td>
<td>a</td>
<td>52</td>
<td>64</td>
<td>41</td>
</tr>
<tr>
<td>D-Lysine</td>
<td>a</td>
<td>55</td>
<td>N/D</td>
<td>40</td>
</tr>
<tr>
<td>L-Serine</td>
<td>a</td>
<td>45</td>
<td>62</td>
<td>37</td>
</tr>
<tr>
<td>D-Serine</td>
<td>a</td>
<td>50</td>
<td>64</td>
<td>38</td>
</tr>
<tr>
<td>L-Glutamic</td>
<td>b</td>
<td>NC</td>
<td>N/D</td>
<td>NC</td>
</tr>
<tr>
<td>Acid</td>
<td>D-Glutamic</td>
<td>b</td>
<td>42</td>
<td>60</td>
</tr>
<tr>
<td>Acid</td>
<td>L-Aspartic</td>
<td>b</td>
<td>39</td>
<td>N/D</td>
</tr>
<tr>
<td>Acid</td>
<td>L-Lysine</td>
<td>a</td>
<td>40</td>
<td>53</td>
</tr>
</tbody>
</table>

NC = non-cooperative (no duplex formation occurred)
N/D = not determined

The results show that glycine in the backbone can be replaced by other amino acids for a moderate loss in hybridization potency. Upon comparing D-lysine versus L-lysine and D-serine versus L-serine, it is evident that D-amino acids are better accommodated in the backbone of PNAS. Furthermore, the introduction of a negatively-charged side chain in the PNA backbone (e.g. glutamic acid and aspartic acid) decreases hybridization potency as indicated by a decreased $T_m$, whereas a positively-charged side chain (e.g. lysine) increases the hybridization potency as indicated by a higher $T_m$. Also, the PNA sequences bound better to antiparallel DNA than to parallel DNA.

Those skilled in the art will appreciate that numerous changes and modifications may be made to the preferred embodiments of the present invention and that such changes and modifications may be made without departing from the spirit of the invention. It is therefore intended that the appended claims cover all such equivalent variations as fall within the true spirit and scope of the invention.
(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 11 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: 2,6- dimiospurine

GTAONTCACT N

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 11 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 3
   (D) OTHER INFORMATION: 2,6- dimiospurine

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: 2,6- dimiospurine

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 8
   (D) OTHER INFORMATION: 2,6- dimiospurine

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 11
   (D) OTHER INFORMATION: lysine residue

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GTNONTCACT N

(2) INFORMATION FOR SEQ ID NO:4:

(ix) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GTAGATCACT

(2) INFORMATION FOR SEQ ID NO:5:

(ix) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: 2,6- dimiospurine
(x i) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GTA GNT CAC T

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 3
(D) OTHER INFORMATION: 2,6-diaminopurine

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 5
(D) OTHER INFORMATION: 2,6-diaminopurine

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 8
(D) OTHER INFORMATION: 2,6-diaminopurine

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:6:

G T N G T C N C T

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:7:

A G T G A C T C A C

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:8:

A G T O A C C T A C

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:9:

A G T G A T C T A C

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 11 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 11
  ( D ) OTHER INFORMATION: lysine residue

( 1 ) SEQUENCE DESCRIPTION: SEQ ID NO:10:
AAAAOGAGAO N 11

( 2 ) INFORMATION FOR SEQ ID NO:11:

( 1 ) SEQUENCE CHARACTERISTICS:
  ( A ) LENGTH: 11 bases
  ( B ) TYPE: nucleic acid
  ( C ) STRANDEDNESS: single
  ( D ) TOPOLOGY: linear

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 1
  ( D ) OTHER INFORMATION: 2,6- diminoopurine

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 2
  ( D ) OTHER INFORMATION: 2,6- diminoopurine

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 3
  ( D ) OTHER INFORMATION: 2,6- diminoopurine

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 4
  ( D ) OTHER INFORMATION: 2,6- diminoopurine

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 7
  ( D ) OTHER INFORMATION: 2,6- diminoopurine

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 9
  ( D ) OTHER INFORMATION: 2,6- diminoopurine

( 1 ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 11
  ( D ) OTHER INFORMATION: lysine residue

( 1 ) SEQUENCE DESCRIPTION: SEQ ID NO:11:
NNNNNGONONG N 11

( 2 ) INFORMATION FOR SEQ ID NO:12:

( 1 ) SEQUENCE CHARACTERISTICS:
  ( A ) LENGTH: 10 bases
  ( B ) TYPE: nucleic acid
  ( C ) STRANDEDNESS: single
  ( D ) TOPOLOGY: linear

( 1 ) SEQUENCE DESCRIPTION: SEQ ID NO:12:
CTCTCCTTTT 10

( 2 ) INFORMATION FOR SEQ ID NO:13:

( 1 ) SEQUENCE CHARACTERISTICS:
  ( A ) LENGTH: 10 bases
  ( B ) TYPE: nucleic acid
  ( C ) STRANDEDNESS: single
  ( D ) TOPOLOGY: linear
(x1) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TTTCTCTC

(2) INFORMATION FOR SEQ ID NO:14:

(1) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 12 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 8
   (D) OTHER INFORMATION: 2,6-diaminopurine attached to
   aminoethyl-lysine backbone

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TTTCGCNGCC CN

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 8 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: 2,6-diaminopurine attached to
   aminoethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: 2,6-diaminopurine attached to
   aminoethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: 2,6-diaminopurine attached to
   aminoethyl-lysine backbone

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GCNNNCGC

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 14 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine
   backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine
   backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 11
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine
   backbone
(x) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CCTTGGCAAGTCTC

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 14 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 2
   (D) OTHER INFORMATION: guanine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: thymine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: thymine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 6
   (D) OTHER INFORMATION: guanine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 7
   (D) OTHER INFORMATION: guanine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 8
   (D) OTHER INFORMATION: guanine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 9
   (D) OTHER INFORMATION: guanine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 10
   (D) OTHER INFORMATION: guanine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 11
   (D) OTHER INFORMATION: thymine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 12
   (D) OTHER INFORMATION: thymine attached to aminomethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 13
   (D) OTHER INFORMATION: cytosine attached to aminomethyl-lysine backbone

(x) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CCTTGGCAAGTCTC

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 14 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: cytosine attached to aminomethyl-lysine backbone
FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 2
D) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 3
D) OTHER INFORMATION: cytosine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 4
D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 5
D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 7
D) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 8
D) OTHER INFORMATION: cytosine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 10
D) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 11
D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 12
D) OTHER INFORMATION: cytosine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 13
D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

FEATURE:

A) NAME/KEY: Modified-site
B) LOCATION: 14
D) OTHER INFORMATION: cytosine attached to aminoethyl-lysine backbone

SEQUENCE DESCRIPTION: SEQ ID NO:18:

CGCTTGGCAQ TCTC

INFORMATION FOR SEQ ID NO:19:

SEQUENCE CHARACTERISTICS:

A) LENGTH: 16 bases
B) TYPE: nucleic acid
C) STRANDEDNESS: single
D) TOPOLOGY: linear
(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 2
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 3
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 8
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 9
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 12
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 15
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:19:

TTAGGATC GTC T

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 12 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 7
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 10
   (D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:20:
(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 8 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 6
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

GCCTTGC

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 5
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 10
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 15
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 16
   (D) OTHER INFORMATION: thymine attached to aminooethyl-lysine
      backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

CCTGACTGGCTTT

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 16 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear
(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 9
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 13
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 15
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 16
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CCGGCGGC ACTGGT

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 16 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 3
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 6
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 12
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 13
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CATGTOGCO OTTAGG

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 15 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ix) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: thymine attached to aminoisethyl-lysine backbone
(i) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 6
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(ii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 10
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(iii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 13
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(iv) SEQUENCE DESCRIPTION: SEQ ID NO:25:

TCGGTGAAG TGTAG

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 16 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 4
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(iii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 8
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(iv) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 14
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(v) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 16
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(vi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

CAGTCACTC AACTCT

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 16 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 3
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(iii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 8
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone
(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 15
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
CCTCCTCCTC

(2) INFORMATION FOR SEQ ID NO:28:

(xi) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: cytosine attached to aminocetyl-lysine backbone

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 4
(D) OTHER INFORMATION: cytosine attached to aminocetyl-lysine backbone

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 3
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 12
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 9
(D) OTHER INFORMATION: cytosine attached to aminocetyl-lysine backbone

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 13
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
CATCGTCCGCTAC

(2) INFORMATION FOR SEQ ID NO:29:

(xi) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 16 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 4
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone
(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 8
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 14
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 16
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(x i ) SEQUENCE DESCRIPTION: SEQ ID NO:29:

CACTCAGTC AAECTT

(2 ) INFORMATION FOR SEQ ID NO:30:

(i ) SEQUENCE CHARACTERISTICS:
   (A ) LENGTH: 16 bases
   (B ) TYPE: nucleic acid
   (C ) STRANDEDNESS: single
   (D ) TOPOLOGY: linear

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 5
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 8
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 15
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(x i ) SEQUENCE DESCRIPTION: SEQ ID NO:30:

CCTCCACTCC C0CCTC

(2 ) INFORMATION FOR SEQ ID NO:31:

(i ) SEQUENCE CHARACTERISTICS:
   (A ) LENGTH: 20 bases
   (B ) TYPE: nucleic acid
   (C ) STRANDEDNESS: single
   (D ) TOPOLOGY: linear

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 3
   (D ) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 7
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i x) FEATURE:
   (A ) NAME/KEY: Modified-site
   (B ) LOCATION: 11
   (D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone
(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 16
(D) OTHER INFORMATION: cytosine attached to aminocetyl-lysine backbone

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:31:
CAACATGGT TCCCCCAAAC

(2) INFORMATION FOR SEQ ID NO:32:
(ix) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 bases
(B) TYPE: nucleic acid
(C) STRAND/DIRECTION: single
(D) TOPOLOGY: linear

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: fluorescence conjugated

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 2
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 8
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 10
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 12
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:32:
OCTCGCTGCT CTC

(2) INFORMATION FOR SEQ ID NO:33:
(ix) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 bases
(B) TYPE: nucleic acid
(C) STRAND/DIRECTION: single
(D) TOPOLOGY: linear

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: conjugated with fluorescent dye

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 2
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 8
(D) OTHER INFORMATION: thymine attached to aminocetyl-lysine backbone

(ix) FEATURE:
(A) NAME/KEY: Modified-site
(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 10
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 12
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) SEQUENCE DESCRIPTION: SEQ ID NO:33:
GTAAGGCTCTCTCTC

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: fluorescein conjugated

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 5
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 8
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 9
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 11
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) SEQUENCE DESCRIPTION: SEQ ID NO:34:
CAAAATGTTCTCGAAA

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: conjugated to fluorescent dye

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 5
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(i) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 8
(D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone
( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 9
  ( D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 11
  ( D ) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:35:
CAATGGTCTCGAA

( 2 ) INFORMATION FOR SEQ ID NO:36:

( i ) SEQUENCE CHARACTERISTICS:
  ( A ) LENGTH: 15 bases
  ( B ) TYPE: nucleic acid
  ( C ) STRANDEDNESS: single
  ( D ) TOPOLOGY: linear

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 1
  ( D ) OTHER INFORMATION: fluorescein conjugated

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 5
  ( D ) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 7
  ( D ) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 9
  ( D ) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 11
  ( D ) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:36:
ACCTGAGGAGCCAG

( 2 ) INFORMATION FOR SEQ ID NO:37:

( i ) SEQUENCE CHARACTERISTICS:
  ( A ) LENGTH: 15 bases
  ( B ) TYPE: nucleic acid
  ( C ) STRANDEDNESS: single
  ( D ) TOPOLOGY: linear

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 1
  ( D ) OTHER INFORMATION: conjugated to fluorescent dye

( i x ) FEATURE:
  ( A ) NAME/KEY: Modified-site
  ( B ) LOCATION: 5
  ( D ) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone
(i) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 7
   (D) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

(ii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 9
   (D) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

(iii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 11
   (D) OTHER INFORMATION: guanine attached to aminoethyl-lysine backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

ACCTGAGGACCCAG 15

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 15 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: fluorescein conjugated

(iii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(iv) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 2
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(v) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 10
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(vi) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 13
   (D) OTHER INFORMATION: thymine attached to aminoethyl-lysine backbone

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

TTGACCACCTCTGA 15

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 15 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(ii) FEATURE:
   (A) NAME/KEY: Modified-site
   (B) LOCATION: 1
   (D) OTHER INFORMATION: conjugated to fluorescent dye

(iii) FEATURE:
   (A) NAME/KEY: Modified-site
( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 1
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 10
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 13
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:39:
TTGCCACGT CCTGA

( 2 ) INFORMATION FOR SEQ ID NO:40:

( i ) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 15 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 1
( D ) OTHER INFORMATION: fluorescein conjugated

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 2
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 7
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 8
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 14
( D ) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:40:
TGCCCCGCA AACGT

( 2 ) INFORMATION FOR SEQ ID NO:41:

( i ) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 15 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

( i x ) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 1
OTHER INFORMATION: conjugated to fluorescent dye

A) NAME/KEY: Modified-site
B) LOCATION: 2
D) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 7
D) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 8
D) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 14
D) OTHER INFORMATION: guanine attached to aminooethyl-lysine backbone

SEQUENCE DESCRIPTION: SEQ ID NO:41:

TGCCGCGGAA AACGT

INFORMATION FOR SEQ ID NO:42:

A) SEQUENCE CHARACTERISTICS:
   A) LENGTH: 15 bases
   B) TYPE: nucleic acid
   C) STRANDEDNESS: single
   D) TOPOLOGY: linear

A) NAME/KEY: Modified-site
B) LOCATION: 1
D) OTHER INFORMATION: fluorescent conjugated

A) NAME/KEY: Modified-site
B) LOCATION: 3
D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 6
D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 12
D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 13
D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

A) NAME/KEY: Modified-site
B) LOCATION: 15
D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

SEQUENCE DESCRIPTION: SEQ ID NO:42:

CTCCTTGCAC GTTCT

INFORMATION FOR SEQ ID NO:43:
( i ) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: conjugated to fluorescein dye

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 3
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 6
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 12
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 13
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 15
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:43:

CCTCOTOCAC GTTCT

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 13 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: fluorescein conjugated

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 1
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 5
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(B) LOCATION: 7
(D) OTHER INFORMATION: thymine attached to aminooethyl-lysine backbone

(i x) FEATURE:
(A) NAME/KEY: Modified-site
(1) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 12 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

(ii) SEQUENCE DESCRIPTION: SEQ ID NO:44:
TGATGCGA CCTCT

(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 12 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

(ii) SEQUENCE DESCRIPTION: SEQ ID NO:45:
TTTCGCGACC CA

(2) INFORMATION FOR SEQ ID NO:46:

(i) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 22 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

(ii) SEQUENCE DESCRIPTION: SEQ ID NO:46:
TCGTGCTCAT GG

(2) INFORMATION FOR SEQ ID NO:47:

(i) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 11 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

(ii) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 1
( D ) OTHER INFORMATION: glycine residue

(iii) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 22
( D ) OTHER INFORMATION: tyrosine residue

(iii) SEQUENCE DESCRIPTION: SEQ ID NO:47:
NTTAGGATT COTGCTCAT GGN

(2) INFORMATION FOR SEQ ID NO:48:

(i) SEQUENCE CHARACTERISTICS:
( A ) LENGTH: 11 bases
( B ) TYPE: nucleic acid
( C ) STRANDEDNESS: single
( D ) TOPOLOGY: linear

(ii) FEATURE:
( A ) NAME/KEY: Modified-site
( B ) LOCATION: 11
( D ) OTHER INFORMATION: tyrosine residue

(iii) SEQUENCE DESCRIPTION: SEQ ID NO:48:
TTTCTCTCTC N
(2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

AAAAGGAAG

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 50:

GAAGGAAGAAA

(2) INFORMATION FOR SEQ ID NO: 51:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 51:

AAAAGTAAG

(2) INFORMATION FOR SEQ ID NO: 52:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 52:

AAAAGTGAA

(2) INFORMATION FOR SEQ ID NO: 53:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

GAGATGAAA

(2) INFORMATION FOR SEQ ID NO: 54:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

GATGGAAAA
(2) INFORMATION FOR SEQ ID NO:55:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

AAAAAAAAAAAA

(2) INFORMATION FOR SEQ ID NO:56:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

AAAAAAAAAAAA

(2) INFORMATION FOR SEQ ID NO:57:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

AAAAAAAAAAAA

(2) INFORMATION FOR SEQ ID NO:58:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

AAAAAAAAAAAA

(2) INFORMATION FOR SEQ ID NO:59:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

AAAAAAAAAAAA

(2) INFORMATION FOR SEQ ID NO:60:

(i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 10 bases
   (B) TYPE: nucleic acid
   (C) STRANDEDNESS: single
   (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

AAAAAAAAAAAA
What is claimed is:

1. A method for enhancing the DNA or RNA sequence specificity of a peptide nucleic acid by incorporating a 2,6-diaminopurine nucleobase in said peptide nucleic acid having formula:

   \[
   \begin{align*}
   \text{R}^1 & \quad \text{O} & \quad \text{L} & \quad \text{N} & \quad \text{O} & \quad \text{R}^7 \\
   \text{R}^7 & \quad \text{N} & \quad \text{H} & \quad \text{N} \quad \text{H} & \quad \text{NH}^+ \quad \text{R}^1
   \end{align*}
   \]

   wherein:
   
   each L is independently selected from a group consisting of naturally-occurring nucleobases and non-naturally-occurring nucleobases, provided that at least one L is 2,6-diaminopurine nucleobase;
   
   each \( R^7 \) is independently hydrogen or \( \text{C}_1-\text{C}_8 \) alkylamine;
   
   \( R^8 \) is \( \text{OH}, \text{NH}_2 \) or \( \text{NH}^\text{Lys}NH_2 \);
   
   \( R' \) is \( \text{H}, \text{COCH}_3 \) or \( \text{t-butoxycarbonyl} \); and
   
   \( n \) is an integer from 1 to 30.

2. The method of claim 1 wherein said each \( R^7 \) is \( \text{C}_3-\text{C}_6 \) alkylamine.

3. The method of claim 2 wherein said each \( R^7 \) is \( \text{C}_4-\text{C}_5 \) alkylamine.

4. The method of claim 3 wherein said each \( R^7 \) is butylamine.

5. The method of claim 1 wherein at least one of said \( R^7 \) is \( \text{C}_3-\text{C}_6 \) alkylamine.

6. The method of claim 5 wherein at least one of said \( R^7 \) is \( \text{C}_4-\text{C}_5 \) alkylamine.

7. The method of claim 6 wherein at least one of said \( R^7 \) is butylamine.

8. The method of claim 7 wherein substantially all of said \( R^7 \) is butylamine.

9. The method of claim 1 wherein the carbon atom to which the substituent \( R^7 \) is attached is stereochemically enriched.

10. The method of claim 9 wherein said stereochemical enrichment is of the R configuration.

11. The method of claim 1 wherein said peptide nucleic acid is derived from an amino acid.

12. The method of claim 11 wherein said peptide nucleic acid is derived from D-lysine.

13. A method for enhancing the DNA or RNA binding affinity of a peptide nucleic acid by incorporating a 2,6-

   diaminopurine nucleobase in said peptide nucleic acid having formula:

   \[
   \begin{align*}
   \text{R}^1 & \quad \text{O} & \quad \text{N} & \quad \text{O} & \quad \text{R}^7 \\
   \text{R}^7 & \quad \text{N} & \quad \text{H} & \quad \text{N} \quad \text{H} & \quad \text{NH}^+ \quad \text{R}^1
   \end{align*}
   \]

   wherein:
   
   each L is independently selected from a group consisting of naturally-occurring nucleobases and non-naturally-occurring nucleobases, provided that at least one L is 2,6-diaminopurine nucleobase;
   
   each \( R^7 \) is independently hydrogen or \( \text{C}_1-\text{C}_8 \) alkylamine;
   
   \( R^8 \) is \( \text{OH}, \text{NH}_2 \) or \( \text{NH}^\text{Lys}NH_2 \);
   
   \( R' \) is \( \text{H}, \text{COCH}_3 \) or \( \text{t-butoxycarbonyl} \); and
   
   \( n \) is an integer from 1 to 30.

14. The method of claim 13 wherein said each \( R^7 \) is \( \text{C}_3-\text{C}_6 \) alkylamine.

15. The method of claim 14 wherein said each \( R^7 \) is \( \text{C}_4-\text{C}_5 \) alkylamine.

16. The method of claim 15 wherein said each \( R^7 \) is butylamine.

17. The method of claim 13 wherein at least one of said \( R^7 \) is \( \text{C}_3-\text{C}_6 \) alkylamine.

18. The method of claim 17 wherein at least one of said \( R^7 \) is \( \text{C}_4-\text{C}_5 \) alkylamine.

19. The method of claim 18 wherein at least one of said \( R^7 \) is butylamine.

20. The method of claim 19 wherein substantially all of said \( R^7 \) is butylamine.

21. The method of claim 13 wherein the carbon atom to which the substituent \( R^7 \) is attached is stereochemically enriched.

22. The method of claim 21 wherein said stereochemical enrichment is of the R configuration.

23. The method of claim 13 wherein said peptide nucleic acid is derived from an amino acid.

24. The method of claim 23 wherein said peptide nucleic acid is derived from D-lysine.

* * * * *
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Col. 5, line 51, please delete “(I)” and insert therefor --(II)--.
Col. 15, line 14, please delete “A” and insert therefor --Å--.
Col. 17, line 16, please delete “CONRCH₂CO₂H” and insert therefor --CONRCH₂CO₂H--.
Col. 17, line 51, “(s, mi., T-CH-CO-)” and insert therefor --(s, mi., T-CH₂-CO-)--.
Col. 22, line 26, Equation, please delete “(Sₙ₋₁)” and insert therefor --(Sₙ₋₁)--.
Col. 27, line 16, please delete “H₂Cl₂” and insert therefor --CH₂Cl₂--.
Col. 30, line 50, please delete “(2H. PhCH)” and insert therefor --(2H. PhCH₂)--.
Col. 33, line 2, please delete “tBu” and insert therefor --t-Bu--.
Col. 37, line 52, please delete “(m,1H,CH)” and insert therefor --(m,1H,CH₂)--.
Col. 41, line 66, please delete “67” and insert therefor --δ--.
Col. 45, line 65, please delete “CIT” and insert therefor --C/T--.

Col. 50, line 25, please delete “2-Amino-60-benyl” and insert therefor --2-Amino-6-O-benzyl--.
Col. 50, line 51, please delete “(2-N-p-Toluenesulfonamido-60-benzylpurinyl)” and insert therefor --(2-N-p- Toluenesulfonamido-6-O-benzylpurinyl)--.
Col. 51, line 45, please delete “(a CH, CH₃O)” and insert therefor --(3 CH, CH₂O)--.
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Col. 51, line 65, please delete “V” and insert therefor --UV--.
Col. 53, line 6, please delete “(caic.)” and insert therefor --(calc.)--.
Col. 53, line 22, please delete “a” and insert therefor --α--.
Col. 53, line 43, please delete “C₆H₅N₂O₄" and insert therefor --C₆H₅N₂O₄--.
Col. 53, line 52, please delete “a” and insert therefor --α--.
Col. 54, line 29, please delete “NaHCO₃," and insert therefor --NaHCO₃--.
Col. 54, line 64, please delete “(NCC₂CON)” and insert therefor --(NCH₂CON)--.
Col. 54, line 65, please delete “(BOC-NHCHN)” and insert therefor --(BOC-NHCH₂N)--.
Col. 54, line 65, please delete “(C(CH))” and insert therefor --(C(CH₃))₂--.
Col. 55, line 31, please delete “(NCH₂CON)” and insert therefor --(NCH₂CON)--.
Col. 57, Table, please delete “≥85" and insert therefor --≥85--.
Col. 58, line 16, please delete “Na₂SO₄" and insert therefor --Na₂SO₄--.
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Col. 62, line 13, please delete "C₂H₅N₃O₃" and insert therefor --C₇H₅N₃O₃--.
Col. 62, line 36, please delete "(DMSO-₆, TMS)" and insert therefor --(DMSO-d₆, TMS)--.

Signed and Sealed this
Twelfth Day of October, 1999

Attest:

Q. TODD DICKINSON
Attesting Officer  Acting Commissioner of Patents and Trademarks