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Published in:
Journal of Geophysical Research: Biogeosciences

Link to article, DOI:
10.1002/2017JG003999

Publication date:
2018

Document Version
Peer reviewed version

Link back to DTU Orbit

Citation (APA):

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Shifts in the Source and Composition of Dissolved Organic Matter in Southwest Greenland Lakes Along a Regional Hydro-climatic Gradient

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Abstract
Dissolved organic matter (DOM) concentration and quality were examined from Arctic lakes located in three clusters across south-west (SW) Greenland, covering the regional climatic gradient: cool, wet coastal zone; dry inland interior; and cool, dry ice-marginal areas. We hypothesized that differences in mean annual precipitation between sites would result in a reduced hydrological connectivity between lakes and their catchments and that this concentrates degraded DOM. The DOM in the inland lake group was characterized by a lower aromaticity and molecular weight, a low soil-like fluorescence, and carbon stable isotope ($\delta^{13}$C-DOC) values enriched by ~2‰ relative to the coastal group. DOC-specific absorbance (SUVA$_{254}$) and DOC-specific soil-like fluorescence (SUVF$_{C_1}$) revealed seasonal and climatic gradients across which DOM exhibited a dynamic we term “pulse-process”: Pulses of DOM exported from soils to lakes during snow and ice melt were followed by pulses of autochthonous DOM inputs (possibly from macrophytes), and their subsequent photochemical and microbial processing. These effects regulated the dynamics of DOM in the inland lakes and suggested that if circumpolar lakes currently situated in cool wetter climatic regimes with strong hydrological connectivity have reduced connectivity under a drier future climate, they may evolve toward an end-point of large stocks of highly degraded DOC, equivalent to the inland lakes in the present study. The regional climatic gradient across SW Greenland and its influence on DOM properties in these lakes provide a model of possible future changes to lake C cycling in high-latitude systems where climatic changes are most pronounced.

Plain Language Summary
Arctic regions are currently experiencing change due to warming climate, and Arctic lakes are natural recorders of this change. We conducted a study of 20 Arctic lakes situated across a climatic gradient in southwest Greenland ranging from cool and wet coastal region to a warm and dry inland region. We found that Arctic carbon cycles are responding to this change by concentrating dissolved organic carbon (DOC), a collection of molecules that form the basis of lake food webs, which may be very old and represent long-term accumulation of carbon in these lake waters. We suspect that the force driving this change is a loss of the connectivity between these lakes and their watersheds. Under this scenario, lakes receive less organic carbon from soils in their surrounding watersheds and perhaps more from fringing aquatic plants. Moreover, earlier loss of lake ice may lead to longer open water periods and the chance for sunlight to convert DOC to carbon dioxide, representing a feedback to regional climate. As global change influences the Arctic broadly, modifications to the hydrologic connectivity of lakes to their watersheds may have important implications for Arctic inland C cycling.

1. Introduction
Arctic terrestrial organic carbon (C) stores are substantial (~1,400 Pg; Tarnocai et al., 2009) but are becoming increasingly reactive and mobile as the biogeochemically active layer deepens with regional warming and as soil microbial activity is enhanced (Cory et al., 2013). Coupled with altered hydrology, these processes driven by changes in global climate result in a growing fraction of the C pool being transferred laterally as particulate and dissolved organic matter (DOM) to streams and lakes (Tank et al., 2012). Moreover, transfer of terrestrial C is complemented by an increase in primary production in lakes due to lengthening of the growing season and elevated nutrient availability (derived from melting permafrost and atmospheric deposition; e.g.,

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Anderson et al., 2017), which potentially provides a further autochthonous source of DOM. These changing C sources suggest that the optical and chemical properties of the DOM in Arctic lakes will change as Arctic regions undergo warming.

Lakes are an important component of many Arctic landscapes and represent hot spots of biogeochemical processes. Arctic lakes are located in environments undergoing climate change at unprecedented rapidity. Roughly 80% of Arctic lakes are situated in tundra landscapes with continuous permafrost, which is changing rapidly (Vincent et al., 2008). The abundance of lakes at high latitudes is exemplified in the Kangerlussuaq area of SW Greenland where ~15% of the land surface consists of lakes (Anderson et al., 2001). This lake district spans well-defined climate, permafrost, environmental, and limnological gradients from the edge of the Greenland ice sheet to the coast. There are clear gradients in effective precipitation and air temperature that influence vegetation and landscape hydrological connectivity (Anderson et al., 2017). These climatic controls result in distinct differences in limnology, for example, thermal stratification, conductivity, nutrients, and hypolimnetic oxygen availability, but most notably of lake dissolved organic carbon (DOC) concentrations from ice sheet to coast (Anderson & Stedmon, 2007).

Sobek et al. (2007) suggested a hierarchical regulation of DOC in lakes in which climate and topographic characteristics set a range of DOC concentrations for a region of lakes. Catchment and lake properties, such as the proportion of wetland and water residence time, influence DOC within a particular lake. Specifically, lakes with high hydrological connectivity are expected to have comparatively high DOC concentrations as a result of allochthonous C inputs. In this study, we define hydrological connectivity with respect to lake DOM cycling, as water-mediated transfer of organic matter to a lake from its landscape (Tetzlaff et al., 2007). Currently, the presence of continuous permafrost means that the hydrological connectivity of SW Greenland lakes to their watersheds via subsurface linkages between tundra soils and lakes is limited. Therefore, we expect that surface runoff dominates and is closely linked to mean annual precipitation (m.a.p.) (Anderson et al., 2017). We anticipate that hydrological connectivity exhibits seasonal variability across the precipitation gradient in which the SW Greenland lakes are situated, strengthening as spring snowmelt runs into the lakes, but weakening as summer progresses, where evapoconcentration in the inland lakes becomes important as indicated by their higher conductivity (Anderson et al., 2001, 2017; Anderson & Stedmon, 2007).

DOC concentrations of Greenland lakes are well described and appear to fit the hierarchical regulation suggested by Sobek et al. (2007) (Anderson & Stedmon, 2007). However, DOM characteristics are not well described, and yet they are important to understand with respect to DOM processing and reactivity as the lakes in the Kangerlussuaq area respond to rapid regional warming (Anderson et al., 2017). Moreover, recent work has shown coherent declines in mean lake DOC concentrations across SW Greenland over the past 10 years, which suggests an alteration, or perhaps an acceleration, of Arctic lake C cycling in this area of rapid global environmental change (Anderson et al., 2017; Saros et al., 2015).

The aim of this paper is to examine the potential sources of DOM in the Arctic lakes of Greenland using chromophoric DOM (CDOM) absorption and fluorescence across well-defined limnological and climate gradients. We summarize observations made over 10 years and place our findings in context of hydrological connectivity of these lakes to their watersheds. We extend the Sobek paradigm to DOM quality by hypothesizing that lakes with higher m.a.p., and thus stronger hydrological connectivity, have DOM characteristics consistent with that of soil-derived DOM. We further hypothesized that lakes with lower m.a.p., and thus weaker hydrological connectivity, accumulate heavily degraded DOM resulting from microbial and/or photochemical processing of both soil-derived and internally produced organic matter. Thus, we expect these lakes to have higher concentrations of DOM but also DOM that is less reactive than that present in lakes with lower concentrations of fresher DOM. Our results are viewed in the context of a rapidly changing Arctic climate and suggest alterations to lake C cycling at northern latitudes that may occur under drier climatic scenarios.

2. Methods

2.1. Study Area

The lakes in this study are located across an approximately 130 km transect ranging from the coast of SW Greenland inland through the Kellyville (Central) region toward the edge of the ice sheet. This transect represented a longitudinal gradient of mean summer temperatures and m.a.p. across SW Greenland that we define...
as a cool-wet Coastal region near 53°W (8°C, 410 mm), containing low Arctic maritime vegetation. Evaporation is an important climatic variable in a warm-dry Central region near 51°W (11°C, 183 mm) and a cool-dry Ice margin region near 50°W (8°C, 269 mm) (Nielsen, 2010). Two lakes of the Ice margin region are fed by glacial meltwater. The ages of the lakes range from 10,000 years (Coastal group) to 8,000 years (Central group) to ~4–5,000 years (Ice margin group) (Bennike & Björck, 2002). Locations and physical properties of the lakes grouped by these regions can be found in Tables S1 and S2. The primary sampling strategy was determined by prior knowledge of the regional limnology (Anderson et al., 2001; Whiteford et al., 2016): we sampled lake clusters with distinctive limnology to provide regional replication when determining lake DOM characteristics, across the longitudinal climatic gradient. The data set contained 20 lakes sampled intensively during 2009, 2013, and 2014 (Table S1), along with data from 29 lakes sampled in 2002 and 2003 (Anderson & Stedmon, 2007) (Table S2).

2.2. Sampling Methods

Water sampling for DOM absorbance and fluorescence, and DOC concentrations was conducted between June and August for all years, with additional under-ice water sampling conducted in April 2014. Sampling was not consistent between years; e.g., Coastal lakes were not sampled in 2003, 2009, or 2014 while in 2013, no under-ice samples were taken. Therefore, we were not able to critically address seasonality effects (see Table S1 for sampling dates). Sampling details for lakes sampled in 2002 and 2003 can be found in Anderson and Stedmon (2007) but were consistent with subsequent sampling in 2009, 2013, and 2014. Sampling was conducted via a small rubber boat at approximately the deepest part of each lake. Surface water samples in this study were collected from the epilimnion of each lake via Van Dorn samplers and filtered through precombusted (450°C, 6 h) Whatman GFF glass filters (0.7 μm nominal mesh size) and stored frozen until analysis, generally from 3 to 6 months after collection. For the 2002 and 2003 lake samples, filtration was achieved through 0.45 μm porosity filters. Anderson and Stedmon (2007) compared results using both filter sizes and note <5% difference. Surface soils (5–10 cm depth) were collected in August 2014 near several lakes each in the Central (n = 5) and Ice margin (n = 4) regions and a known mass (12–25 g) of wet soil added to 50 mL ultrapure laboratory water and extracted in the dark for 24 h. Then, the soil extract solutions were filtered through GFF filters and treated as any other DOM sample.

2.3. Optical Properties

Prior to measurement for absorbance and fluorescence, samples were thawed to room temperature. Absorbance was measured between 240 and 750 nm on a Varian 300 UV spectrophotometer using 1, 5, or 2.3. Optical Properties

Prior to measurement for absorbance and fluorescence, samples were thawed to room temperature. Absorbance was measured between 240 and 750 nm on a Varian 300 UV spectrophotometer using 1, 5, or 10 cm quartz cells depending on the amount of CDOM present in the sample. Strongly absorbing samples were diluted. Ultrapure Milli-Q laboratory water (18.2 MΩ resistivity) was used as a blank. Absorbance was converted to Napierian absorption coefficients (α, in m⁻¹) using the following equation:

\[
a_\lambda = \frac{2.303A_\lambda}{L},
\]

where A is the blank-corrected absorbance from the spectrophotometer, λ is the wavelength in nm, and L is the path length of the quartz cell in millimeters. CDOM was quantified with the absorption coefficient at 254 nm, α₂⁵₄. Several derived parameters from absorption were computed to assess CDOM quality. Specific ultraviolet absorption at 254 nm (SUVA₂⁵₄) was computed as the decadal absorption at 254 nm divided by DOC concentration as an index of aromatic carbon content (m² g⁻¹ C) (Weishaar et al., 2003). Likewise α₂⁷₅ was computed as the Napierian absorption at 375 nm divided by DOC concentration. The latter index was computed to compare our results with prior work (Anderson & Stedmon, 2007).

CDOM absorption decreases with increasing wavelength in an approximately exponential fashion. The rate of change in absorption with wavelength is quantified as the slope (S) of an exponential function fit to an absorption spectrum and is indicative of DOM quality. Absorption spectral slopes, S, were measured from 300 to 650 nm with the following equation:

\[
a_\lambda = a_\lambda e^{-S(\lambda-\lambda_0)} + K,
\]

where \(\lambda_0\) is 375 nm and K is a background constant. Spectral slopes also were measured from 275 to 295 nm (\(\lambda_{275-295}\)) and from 350 to 400 nm (\(\lambda_{350-400}\)) with the spectral slope ratio (\(S_{350-400}/S_{275-295}\)) being the ratio of the slopes computed over these two narrower wavebands (Helms et al., 2008).
CDOM fluorescence was measured on separate Varian Eclipse spectrofluorometers for the 2002–2003 data and the 2009–2014 data. For each instrument, standard corrections for lamp excitation and detector response were applied as well as corrections for inner filter effects. Fluorescence intensity was calibrated into Raman units (Cory et al., 2010; Lawaetz & Stedmon, 2009). Excitation wavelengths were sampled from 240 to 450 nm at 5 nm intervals. Emission wavelengths were sampled from 300 to 600 nm at 2 nm intervals. Milli-Q water was used as a blank. Emission spectra were concatenated into excitation-emission matrices (EEMs) and visualized as contour plots. Two parameters derived from EEM fluorescence, the biological index (BIX) and the humification index (HIX), were extracted from EEMs measured on each sample. BIX indicates recent autochthonous biological production, while HIX indicates the degree of complex, high molecular weight organic matter rich in aromatic moieties, resulting from the formation of humic substances (Birdwell & Engel, 2010).

2.4. Dissolved Organic Carbon (DOC)

After optical analyses, samples were acidified with 85% H₃PO₄ to pH 2 and sparged for 10 min with ultra-high purity argon to remove inorganic carbon (as CO₂). Sparged samples were then measured for DOC and δ¹³C-DOC on an OI Analytical 1030D Aurora analyzer in wet oxidation mode coupled to a Thermo Delta V Plus isotope ratio mass spectrometer (Osburn & St-Jean, 2007). DOC concentrations were calibrated each analytical day with prepared solutions of caffeine, and δ¹³C values were normalized to the Vienna Peedee belemnite (‰versus PDB) scale with prepared solutions from IAEA sucrose (δ¹³C = −10.8‰) and caffeine (δ¹³C = −27.77‰). Error on DOC concentrations was <3%, and reproducibility on δ¹³C-DOC values was ±0.19‰ (N = 23).

2.5. Literature Data

Absorbance and DOC concentration data from 2002 and 2003 (exclusive of high conductivity oligosaline lakes) were acquired from Anderson and Stedmon (2007). Details of sampling and analysis protocols are contained therein. Briefly, CDOM absorbance was measured on a Shimadzu UV-2401PC spectrophotometer, while DOC concentrations were measured on a Shimadzu 5000 TOC analyzer. EEM fluorescence also was measured on those samples on a Varian Eclipse spectrofluorometer, but not included in that study; these data are included in the present study.

2.6. Parallel factor analysis Modeling

Parallel factor analysis (PARAFAC) was conducted on fluorescence EEMs from SW Greenland lakes sampled during 2002–2003 and during 2009–2014 (Stedmon et al., 2003). Each EEM was normalized to its integrated fluorescence, which allows highly fluorescent and weakly fluorescent samples to be modeled simultaneously without highly fluorescent samples exerting significant leverages on the PARAFAC model (Murphy, Bro, et al., 2014). A series of three to eight component models were fitted to the data, and a six-component model was validated. Model validation was achieved through splitting the data in half and modeling each half separately, fitting 10 models with random starts, and by inspection of residuals (Murphy, Bro, et al., 2014). The loadings in the excitation and emission mode for each component were matched to the OpenFluor database, a collection of PARAFAC models from a variety of natural and artificial systems (Murphy, Stedmon, et al., 2014). Matching at >95% similarity was used to determine if a component was identical to components from other models. MATLAB software and the toolbox DOMFluor were used for all PARAFAC models (Stedmon & Bro, 2008).

2.7. Statistical Tests

Most DOM concentration variables (e.g., a₂₅₄ and DOC) had significantly different standard deviations as determined by Bartlett’s test, so comparisons of DOM optical and chemical median values among lake regions were made using a Kruskal-Wallis nonparametric analysis of variance (ANOVA) with a Mann-Whitney posttest, or modeled with permutational multivariate analysis of variance (PERMANOVA) based on Euclidean distances (permutation n = 9,999 using a bootstrap approach) and conducted in PAST v.3.0. For correlation analysis, Pearson’s coefficient (r) was computed for normally distributed data and Spearman’s rho (ρ) was computed on ranks for nonnormally distributed data. Principal component analysis (PCA) was conducted on a subset of results using the PLS_Toolbox for MATLAB to explore DOM optical and chemical variables in relation to any separation of sample scores between the lakes, grouped by region. Data for PCA were mean-centered prior to modeling. A linear mixed effects regression model (lme) was fitted to
seasonal SUVA$_{254}$ and DOC-specific fluorescence (SUVF) data from 2014 using the Statistics and Machine Learning Toolbox for MATLAB. These variables were included as fixed effects, which influenced the slope of the regression and random effects grouped by month, which influenced the intercept.

3. Results

3.1. DOC and CDOM Concentrations in Lakes Along Kangerlussuaq

DOC and CDOM concentrations varied both seasonally and spatially in lakes across SW Greenland (Figure 1), but in general were highly related (log DOC = $-0.415 \pm 0.061 + 0.955 \pm 0.037 \times \log$ a$_{254}$; adjusted $R^2 = 0.88; p < 0.0001; n = 96$). Summary statistics were based on ranks because most variables did not pass Bartlett’s test of equal standard deviations. In general, the central lake group had much higher median concentrations of DOC (34.92 mg L$^{-1}$) than the Coastal lake group (2.14 mg L$^{-1}$) or the Ice margin lake group (7.33 mg L$^{-1}$) (Table 1). The Central lake group had the lake with the overall highest DOC concentration (61.30 mg L$^{-1}$), while the Ice margin lake group had the lowest DOC concentration (0.98 mg L$^{-1}$), which was very similar to the minimum DOC concentration of the Coastal lake group (1.02 mg L$^{-1}$). The Central lake group also had the largest amount of CDOM quantified as the absorption coefficient at 254 nm ($a_{254}$; Table 1) and exemplified in absorption spectra from 250 to 550 nm (Figure S1 in the supporting information). Median $a_{254}$ value for the Central lake group (96.10 m$^{-1}$) was over fourfold higher than the median for the Ice margin lake group (22.47 m$^{-1}$) and nearly tenfold higher than the Coastal lake group (9.36 m$^{-1}$). When considered together, these differences in median DOC concentrations and median $a_{254}$ values were significantly different among the lake groups (PERMANOVA: F = 59.43; $P = 0.0001$). Further, both DOC and $a_{254}$ were negatively correlated with m.a.p. (Pearson’s $r = -0.891$ and $-0.770$, respectively; $P < 0.0001$ for both variates).

3.2. CDOM Absorption and Fluorescence

CDOM quality, as determined by spectral slope values ($S$ and $S_R$), also varied among the lake groups. Median $S$ value for the Coastal lake group was 17.07 m$^{-1}$ and increased for the Ice margin lake group (20.71 m$^{-1}$) to the highest median value for the Central lake group (23.33 m$^{-1}$). Significant differences were found between medians of all lake groups, indicating that the Central lakes had higher CDOM absorption than the Ice margin and Coastal lakes ($P < 0.0001$; Figure S1). A similar result was found for differences among $S_R$ values for each lake group. Median $S_R$ values were lowest for the Coastal lakes (0.99) and higher for the Central (1.29) and Ice margin (1.28) lakes. The Central lake $S_R$ values were significantly different from both the Central lakes and the Ice margin lakes ($P < 0.001$ in each case), though the Central and Ice margin lakes were not significantly different from one another ($P = 0.562$).

Despite the Central lake group being markedly different from the Coastal and Ice margin lakes with respect to DOC concentration and CDOM absorbance, excitation-emission matrix (EEM) spectra of examples of lakes from each lake group exhibited both common and distinct patterns (Figure S2). The DOM fluorescence intensity varied widely as across lake groups and was greatest for the Central lakes. For example, Coastal lake AT10 had fluorescence values approaching four QSU and centered on excitation/emission (Ex/Em) of 240/430 (nm/ nm), squarely in the region of typical freshwater CDOM fluorescence (Figure S2a). Notable secondary Ex/Em occurred in the protein-like fluorescence region (275/340). Central lake SS2 had fluorescence values approaching nearly 60 QSU (Figure S2b), whereas Ice margin lake SS906 had maximum fluorescence
Central lakes had the highest fluorescence intensities for all components of the three lake groups (Figure 2a). Lakes were most dominated by C2 and C1 but showed a distinct difference in the relative amount of C3. Highest to lowest mean FMax values were ranked differently among the lake groups. For the Coastal lakes, mean FMax values for C3 were lowest: C2 > C1 > C4 > C6 > C5 > C3. For the Central lakes, C3 was dominant: C3 > C2 > C1 > C4 > C5 = C6. For the Ice margin lakes, C3 and microbial humic-like C4 were of equal intensity, while these lakes contained much less C6 fluorescence: C2 > C1 > C3 = C4 > C5 > C6. The components were ranked similarly for the soil DOM extracts: C1 = C2 > C4 > C5 > C3 = C6 (Figure 2b). C3 was notably lacking in the soil DOM extracts from the Central and Ice margin regions compared to those regions’ lakes, suggesting that C3 very likely was indicative of transformation and degradation of DOM within the lakes. Median C3 values were significantly greater in the Coastal lakes compared to other lake groups (P < 0.001). Thus, the Central lakes appeared to have the most transformed or degraded DOM compared to DOM in the Ice margin or the Coastal lakes, exhibiting the least similarity to the soil DOM extracts.

**3.3. δ13C-DOC Values**

Stable carbon isotope values of DOC (δ13C-DOC) were generally consistent across lake groups. The median δ13C-DOC value for the Coastal lakes was (−26.20‰), which was isotopically depleted compared to the Central lake group (−24.53‰) and the Ice margin lake group (−24.71‰), the latter of which has the widest range of values, including a highly depleted δ13C-DOC value of −32.85‰ for lake SS911. A Kruskal-Wallis test revealed that median δ13C-DOC values for the Coastal lakes were significantly depleted relative to the Central lakes (P < 0.001) and the Ice margin lakes (P < 0.01), whereas the median δ13C-DOC values for the Central and Ice margin lakes were not significantly different.

**Table 1**

<table>
<thead>
<tr>
<th>Variable</th>
<th>Lake group</th>
<th>Coasal (n = 10)</th>
<th>Central (n = 46)</th>
<th>Ice margin (n = 40)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DOC (mg C L⁻¹)</td>
<td>Median</td>
<td>2.14</td>
<td>34.92</td>
<td>3.73</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>1.02</td>
<td>15.31</td>
<td>0.98</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>4.90</td>
<td>61.30</td>
<td>13.17</td>
</tr>
<tr>
<td></td>
<td>Median</td>
<td>9.36</td>
<td>96.10</td>
<td>22.47</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>3.71</td>
<td>38.75</td>
<td>1.58</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>30.41</td>
<td>302.89</td>
<td>57.86</td>
</tr>
<tr>
<td>a254 (m⁻¹)</td>
<td>Median</td>
<td>17.07</td>
<td>23.33</td>
<td>20.71</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>13.00</td>
<td>18.79</td>
<td>12.57</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>24.57</td>
<td>27.32</td>
<td>25.10</td>
</tr>
<tr>
<td>S (μm⁻¹)</td>
<td>Median</td>
<td>0.99</td>
<td>1.29</td>
<td>1.28</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>0.53</td>
<td>0.90</td>
<td>0.56</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>1.49</td>
<td>1.63</td>
<td>1.63</td>
</tr>
<tr>
<td>SUVA254 (L mg C⁻¹ m⁻¹)</td>
<td>Median</td>
<td>2.23</td>
<td>1.18</td>
<td>1.40</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>1.18</td>
<td>0.57</td>
<td>0.64</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>3.21</td>
<td>2.96</td>
<td>2.30</td>
</tr>
<tr>
<td>a375* (L mg C⁻¹ m⁻¹)</td>
<td>Median</td>
<td>0.73</td>
<td>0.17</td>
<td>0.23</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>0.19</td>
<td>0.06</td>
<td>0.07</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>0.93</td>
<td>0.65</td>
<td>0.72</td>
</tr>
<tr>
<td>δ13C-DOC (% vs VPDB)³</td>
<td>Median</td>
<td>−26.20</td>
<td>−24.53</td>
<td>−24.71</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>−27.39</td>
<td>−25.49</td>
<td>−32.85</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>−25.79</td>
<td>−23.05</td>
<td>−22.08</td>
</tr>
<tr>
<td>BIX</td>
<td>Median</td>
<td>0.52</td>
<td>0.68</td>
<td>0.68</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>0.45</td>
<td>0.49</td>
<td>0.50</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>0.62</td>
<td>0.87</td>
<td>1.38</td>
</tr>
<tr>
<td>HIX</td>
<td>Median</td>
<td>4.63</td>
<td>4.81</td>
<td>3.13</td>
</tr>
<tr>
<td></td>
<td>Min.</td>
<td>2.56</td>
<td>2.16</td>
<td>1.17</td>
</tr>
<tr>
<td></td>
<td>Max.</td>
<td>6.41</td>
<td>11.13</td>
<td>8.37</td>
</tr>
</tbody>
</table>

³No δ13C-DOC values available for the lakes sampled during 2002 and 2003, in which case number of observations for the Coastal, Central, and Ice margin lake groups were n = 7, n = 30, and n = 24, respectively.

Indices of biological production (BIX) and humification (HIX) also varied among lake groups (Table 1) (Birdwell & Engel, 2010). The Coastal lakes had lower BIX values than the Central or Ice margin lakes, and in each case, the differences were significant (P < 0.001 for each). Median BIX values for the Central and Ice margin were not significantly different (P > 0.05). Median HIX values showed very similar degrees of humification among the three lake groups (Table 1). The Coastal lakes had the narrowest range of HIX values, while the Central lakes had the highest. Only the Central and Ice Margin lake groups were significantly different from one another with respect to HIX (P = 0.002).
3.4. Controls on Greenland Lake DOM Quantity and Quality

Ordination by PCA of DOC-normalized absorbance and fluorescence results (SUVA and SUVF, respectively) along with qualitative indicators (S, S0, BIX, and HIX) allowed us to view hydrological connectivity, internal DOM production, and photodegradation as the key processes describing the quality of DOM in these lakes (Figure 3). The first three components explained 79% of the variability in the data; we then correlated the PC axis values against CDOM variables (Table 2) and against conductivity data for the lakes (2013 data only; Table S4).

Principal component 1 (PC1) accounted about 40% of the variability in the CDOM lake data and was also positively correlated with m.a.p. (Pearson’s $r = 0.874, P < 0.001, n = 14$) and negatively correlated to conductivity (2013 samples only, Pearson’s $r = -0.595, P = 0.025; n = 14$; Table S4). Thus, PC1 represented increasing hydrological connectivity between lakes and their catchments. Values for SUVF$_{C1}$, SUVF$_{C2}$, SUVA$_{254}$, and $a^s_{275}$ had very high Spearman’s rho ($\rho$) correlation values to PC1, which supported our hypothesis that hydrological connectivity mobilized allochthonous aromatic-rich DOM into the lakes (Table 2 and Figures 3a and 3c). Moreover, $S_0$ was negatively correlated with PC1 ($\rho = -0.586$; Table 2), which indicated that samples having high scores on PC1 also had comparatively higher molecular weight. These effects on the linkage of hydrological connectivity and soil-derived DOM were strongest for the Coastal lakes which also had the highest m.a.p.

PC2 (~26% variation explained) had the highest significant and positive $\rho$ values found for BIX, SUVF$_{C6}$, and SUVF$_{C6}$, all indicators of recent biological sources of DOM (Table 2) (Birdwell & Engel, 2010). These substances are enriched in lakes having moderate to high amounts of primary and secondary production, including macrophytes (Lapiere & Frenette, 2009; Tank et al., 2011; Zhang et al., 2013). PC2 was negatively correlated to m.a.p. ($r = -0.866; P < 0.0001; n = 14$) and positively correlated with conductivity ($r = 0.574, P = 0.032, n = 14$; Table S4), which suggested accrual of internally produced DOM in the lakes with evapoconcentration. This result is consistent with a decline in hydrological connectivity, and hence in the influence of soil-derived DOM.

Scores on PC3 (~13% variation explained) had the highest negative correlation with m.a.p. ($r = 0.923; P < 0.0001; n = 14$) and positive correlation with conductivity ($r = 0.743, P = 0.002, n = 14$) (Table S4). Further, the scores on PC3 had positive correlations with the humification index (HIX) and SUVF$_{C3}$, and a significant negative correlation with $S_0$ (Table 2). These correlations combined indicate that this PC axis reflected a combination of microbial and photochemical processing of lake DOM.

Scores for samples coded by lake group showed clear separation among the three climatic regions when PC1 was plotted against PC2 or PC3 (Figures 3b and 3b). Scores from the cool-wet Coastal region cluster in the bottom right of each scatterplot and reflect the higher degree of hydrological connectivity as postulated by Anderson and Stedmon (2007). In Figure 3b, scores from both the cool-dry Ice margin region and warm-dry Central region dominate the top part of the PC1-PC2 biplot, which suggests that internally produced DOM is important in those lakes. Scores from the warm-dry Central lake region dominate the top part of the PC1-PC3 biplot (Figure 3d); most scores are negative on PC1 but also positive on PC3. Loadings for $S$, $S_0$, and BIX all plot negatively on PC1, but notably $S$ and $S_0$ plot positively on PC3 and nearly directly opposite the indicators for soil DOM (e.g., SUVA$_{254}$, $a^s_{275}$, and SUVF$_{C2}$). Further, SUVF$_{C3}$, SUVF$_{C6}$, and HIX all cluster in the top-right region of the loadings biplot, positive on both PC1 and PC3. HIX is higher in environments with more humified DOM, but HIX values also decrease with photodegradation (Birdwell & Engel, 2010; Catalán et al., 2013). Moreover, lakes from the warm-dry Central region had higher scores on PC3 for these variables than did lakes from the cool-dry Ice margin region. These results indicate that substantial microbial and photochemical degradation, both of which can be enhanced by retention time, exert a major secondary control of DOM in these lakes as a consequence of drier conditions and loss of hydrological connectivity.
It is important to note that several lakes from different lake groups overlap with respect to where their scores plot in this analytic space. We interpret this overlap to represent the gradient of temperature and precipitation that distinguishes the cool-wet from the warm-dry and cool-dry lake regions. This climatic gradient is reflected in conductivity values of the lakes (Anderson et al., 2001; Ryves et al., 2006).

### 3.5. Seasonal Effects of DOM Quality

Seasonal variability of DOM quality in the lakes was revealed the SUVF\textsubscript{C1} and SUVA\textsubscript{254} data from the Central and Ice margin lakes in 2014, and the soil extract data, also from 2014 (Figure S4). A linear mixed effects (lme) regression model fit to these data, with log SUVA\textsubscript{254} as the predictor variable and with a random intercept that varied by month, revealed two groups of samples: those in June and August, which trend along a slope with the soil extracts, and those largely from April, which have a similar slope but a different intercept (Table S5). The model was substantially improved by inclusion of the soil extract data ($r^2 = 0.95; P < 0.001; n = 34$), compared to their exclusion ($r^2 = 0.87; P < 0.0002; n = 25$), yet no significant difference in slope value was found with or without the soil extract data (unpaired $t$ test with Welch correction: $P = 0.8858$).

Figure 3. (a and c) Loadings and (b and d) scores plots of lake CDOM properties from PCA. PC1 represented hydrologic connectivity, PC2 represented internal production of DOM, and PC3 represented degradation. Refer to Table 1 for variable definitions.
4. Discussion

4.1. Hydrological Connectivity Drives Soil-Derived DOM Inputs Into SW Greenland Lakes

Our results suggested that DOM quality in lakes across SW Greenland exhibits shifts in both sources and processing along decreasing precipitation gradients that weaken hydrological connectivity between lakes and the soils in their watersheds. The results complement the Sobek et al. (2007) paradigm of hierarchical regulation of DOC concentration in lakes by showing that at higher m.a.p., there is a greater influence of catchment soils on DOM quality, even at low DOC concentrations. This explanation is consistent with our observations from the Coastal lakes, situated in a region with higher m.a.p., which indicated a stronger hydrological connectivity between the Coastal lakes and their watersheds, despite lower median DOC concentrations. Coastal lakes had lower $\delta^{13}$C-DOC values, larger SUVA254 values, and a larger relative amount of C1 and C2 fluorescence relative to C3 fluorescence (Table 1). Further, the distribution pattern of PARAFAC components for the Coastal lakes more closely resembled the soil extracts, which were dominated by C1 and C2 fluorescence (Figure 2).

The low median DOC concentration in the Coastal lakes ($2.14 \text{ mg L}^{-1}$) was likely due to the wetter climatic regime in the Coastal lake region but also perhaps to the soil properties of each lake’s catchment (Table 1) (Nielsen, 2010; cf. Mulholland, 2003). Flushing of soil OM into streams and catchments varies considerably with rainfall dynamics and soil properties, and higher rainfall does not always equate to higher DOC concentrations (Raymond & Saiers, 2010; Sobek et al., 2007) despite evidence that high DOC concentrations in lakes resulted from higher connectivity (Martin & Soranno, 2006). Flushing implies short retention time; thus, we expect younger DOM in the Coastal lakes (Anderson & Stedmon, 2007).

Key terrestrial DOM quality parameters for each lake group, such as SUVA254 specific UV fluorescence for C1 (SUVF$_{C1}$), and $\delta^{13}$C-DOC values, illustrated the first-order control on lake DOM properties exerted by hydrological connectivity, using m.a.p. as a proxy (Figures 4a–4c). Each parameter changed with decreasing m.a.p. in a way which indicated that loss of hydrological connectivity of a lake to its watershed diminished the influence of soil-derived DOM sources in the lakes. For example, SUVA254 values indicated less aromatic DOM as m.a.p. decreased ($r^2 = 0.17; P < 0.001; n = 96$), while SUVF$_{C1}$ (normalized to DOC as for a254) indicated a loss of soil humic fluorescence across this environmental gradient ($r^2 = 0.08; P = 0.005; n = 96$) (Figures 4a and 4b). These weaker correlation coefficients reflect the fact that mean values were significantly different between Coastal lakes and Inland and Ice margin lakes, but not significantly different between Ice margin and Central lakes. This result can be explained by the larger difference in m.a.p. between Coastal lake group and the Ice margin or Central groups (141 and 227 mm, respectively) than between the Central and Ice margin groups (86 mm). Moreover, as m.a.p. decreases from the coast inward, $\delta^{13}$C values of lake DOM increased from approximately $-26\%$ to approximately $-24.5\%$ ($P < 0.001$; Table 1); which reflected a shift from soil-derived terrestrial DOM, which is rapidly transferred to the lakes, as at the coast, to processed DOM in the Central and Ice margin lakes (Figure 4c). Typical $\delta^{13}$C-DOC values for terrestrial DOM fall between $-25$ and $-29\%$ (Bauer, 2002). Thus, the Coastal lakes exhibited the strongest hydrological connectivity, reflected in the largely soil-like quality of their DOM (Figure 3).
The gradient of hydrological connectivity demonstrated on the quality of DOM in the SW Greenland lakes follows earlier work that proposed a model based on $a_{375}$ values plotted against $S$ values, which showed regional separation between Coastal lakes (which had high values of $a_{375}$ and low values of $S$) and Central and Ice margin lakes (which had low $a_{375}$ values and high $S$ values) (Anderson & Stedmon, 2007). We combined the Anderson and Stedmon (2007) data from 2002 to 2003 with the 2009–2014 data from the present study and included the soil extracts as a model of a purely terrestrial end-member source of DOM in the lakes (Figure 5). The regional differences are clear in that the Coastal lakes have $a_{375}$ values of 0.5 to 1.0 L mg C$^{-1}$ m$^{-1}$ and $S$ values between 14 and 20 μm$^{-1}$. The soil extracts have higher $a_{375}$ values than the cluster of Coastal lakes, as would be expected from the humic substances recently extracted from soil organic matter (Jaffé et al., 2008). However, the $S$ values of the soil extracts fall within the range observed for the Coastal lakes. Therefore, our results support the hypothesis of Anderson and Stedmon (2007) that this separation was caused by the degree of the hydrological connectivity whereby lakes that are better connected to their watersheds in the cool, wet Coastal region has “fresher” CDOM than lakes that are disconnected in the drier Central and Ice margin regions.

By contrast, Central and Ice margin lakes are markedly different from the soil extracts with $a_{375}$ values between 0.06 and 0.75 L mg C$^{-1}$ m$^{-1}$ and $S$ values generally above 18 μm$^{-1}$. High values for $S$ and low $a_{375}$ suggest CDOM, which has since been photodegraded and is comparable to prairie lakes also influenced by evapoconcentration and extensive solar photodegradation (Osburn et al., 2001, 2011; Waiser & Robarts, 2004). The significant differences in these CDOM qualitative parameters between the low m.a.p., arid Central and Ice margin lakes, and the high m.a.p. Coastal lakes indicate that alternative processes beyond simple dilution of soil-sourced DOM exert a major second-order control on DOM quality and quantity in these Arctic lakes. The largest changes occur in the Central lakes, which are the most disconnected from their watersheds and have the longest retention times under conditions of decreasing m.a.p. and increasing evapoconcentration. Thus, drier climates of the Central and Ice margin lake groups also result in higher stocks of refractory DOC than the wetter Coastal lake group, due to decreasing m.a.p. and, as a result, a longer retention time allowing for a greater influence of internal processes within those lake groups.

**4.2. Internal Processes in SW Greenland Lakes Modify and Amend Inputs of Soil-Derived DOM**

The negative relationship between PC2 and m.a.p. supported our hypothesis that the deviation of DOM quality in the SW Greenland lakes away from characteristics consistent with soil sources was mainly caused by processes resulting from a lack of hydrological connectivity. This reduces the supply of terrestrial, soil-derived DOM and enhances evapoconcentration, promoting accumulation of both allochthonous and autochthonous DOM (Osburn et al., 2011). As a result of declining m.a.p., both internal DOM production and photodegradation appear to modulate the soil-derived DOM signature in Ice margin and Central lake groups.

Internal DOM production may be dominated by lateral inputs from littoral submerged and emergent vegetation. The SW Greenland lakes in this study are oligotrophic: chlorophyll $a$ concentration is <1 μg L$^{-1}$, total $N$ ranges from 0.1 to 1.5 mg L$^{-1}$, and total phosphorus ranges from 5 to 10 μg L$^{-1}$ (Anderson et al., 2001; Saros et al., 2015; Whiteford et al., 2016). It is thus unlikely that annual water column primary production alone could support the DOC concentrations in these lakes, e.g., roughly 35 mg L$^{-1}$ for the Central lakes. However, considering that the Central lake group exhibited the driest conditions (lowest m.a.p.), we hypothesize that macrophytes and/or aquatic mosses or benthic microalgae growing along the littoral zone supplement the autochthonous input.

In macrophyte-rich temperate lakes, strong protein- and C2-like fluorescence has been observed similar to the macrophyte-rich waters of the Florida Everglades, in which a PARAFAC component was found to be a statistical match to C2 in the SW Greenland lakes model (Lapierre & Frenette, 2009; Yamashita et al., 2010; Zhang et al., 2013). Although we had no measurements of macrophyte exudates, we suspect, based on field
Large intra-annual variability in the DOM quality of the SW Greenland lakes in 2014 was described by the lme regression model (Table S5 and Figure S4). Indeed, this seasonal variability in SW Greenland lake DOM properties suggested a “pulse-process” dynamic as the arctic summer creates a complex environment in which hydrology, climate, and sunlight modulate DOM sources and cycling in these lakes. Mean values (± standard error of the mean) of SUVA254 and SUVFC1 illustrate this dynamic (Figure 6). Mean values of monthly data (April, June, and August) for 2014 from the lme model were combined with mean values of the three lake groups for 2013 to represent the climatic gradient of decreasing m.a.p. The mean soil extract values also presented to represent strong hydrological connectivity to the landscape, as for Figure 5.

April samples were taken under ice, representing conditions prior to the open water period and the lowest influence of soil DOM in the lakes, hence the lowest mean values of both SUVA254 and SUVFC1. We
interpret these results to represent the annual accrual of processed DOM in the lakes, in other words, the DOM "state" of the lakes after the end of the arctic summer and during the period of under-ice microbial processing over the winter (Bertilsson et al., 2013). Mean soil extract values and April under-ice values thus represented end-points of fresh "pulsed" DOM and degraded “processed” DOM, respectively, defined by the dotted regression line in Figure 6. June and August mean values deviate from this line. April was significantly lower than both June (MANOVA: $P < 0.001$) and August ($P < 0.0001$); August was significantly different than June ($P = 0.02$). While both months shared characteristics with soil DOM (a similar intercept in the lme model), their deviations toward higher log SUVF$_{C_1}$ are consistent with an increase in structurally complex humic DOM, such that might arise not only from soil sources seeping into the lakes after ice melt but also from internal inputs of macrophytes or Chara (Tank et al., 2011). We have no observations of these lakes between August and early autumn during ice-formation, but such observations might clarify the seasonal patterns described here. Along with extensive and parallel processing of autochthonous and allochthonous DOM, via photochemical and/or microbial degradation as seen in other studies, we hypothesize that June and August DOM properties return to values similar to April DOM by autumn during ice-formation (Guillemette et al., 2013; Osburn et al., 2011).

The seasonal "pulse-process” dynamic described here is similar to DOM dynamics observed in the Green Lakes Valley catchment in the Colorado Rocky Mountains, USA (Miller & McKnight, 2010). Under-ice DOM in alpine and subalpine lakes exhibited C3-like fluorescence and was notably lacking in C1-like fluorescence. However, the summer snowmelt, which dominates the lakes’ hydrologic cycles, shifts DOM quality to more complex humic material with C1-like fluorescence. Later in the season, the fluorescence signal shifted back to C3-like fluorescence, while SUVA$_{254}$ values declined from ~4.0 to 1.5 L mg C$^{-1}$ m$^{-1}$ (Miller & McKnight, 2010). The consistency between seasonal patterns in these two regions is supported by the pulse of snowmelt driving the hydrologic cycles in these two very different environments.

The mean values for the Coastal lakes fell along the “pulse-process” line, indicating the pulses of soil DOM from surface soil horizon thaw and subsequent seepage into the lakes due to strong hydrological connectivity in these lakes but also its subsequent degradation after the initial pulse. Across the climatic gradient of decreasing m.a.p., the Coastal lakes were significantly different from Ice margin (MANOVA: $P = 0.018$) and from Central lakes (MANOVA: $P = 0.0033$); Ice margin and Central lake groups were not significantly different. This indicated that internal processes ongoing in those lakes during the open water period were more similar to one another than to the Coastal lakes. Thus, despite relatively low input fluxes of soil DOM after ice melt, evapoconcentration in these arid regions can both concentrate DOM but also facilitate its photochemical degradation (Mulholland, 2003). Based on the SUVA$_{254}$ and SUVF$_{C_1}$ results in Figure 6, the lowest m.a.p., highest DOC Central lakes exhibit the strongest effects of these parallel processes, especially photodegradation.

### 4.4. Implications for Carbon Cycling in Arctic Lakes

Results of our study have important implications for lake C cycling in the Arctic. First, the climatic gradient under which the SW Greenland lakes currently exist provide a means of predicting future Arctic lake C cycling scenarios with respect to DOM. Recent work on the Arctic hydrological cycle has revealed that while many Arctic regions currently are in a water surplus in fluvial systems driven by permafrost melt, other regions may become drier (Bring et al., 2016). This means that circumpolar lakes situated in cool wetter climatic regimes with strong hydrological connectivity (e.g., Toolik) could have reduced connectivity under a drier future climate and evolve toward an end-point along the hydrological connectivity-C processing gradient represented by the Central lakes in the present study. Moreover, Šmejkalová et al. (2016) used Moderate Resolution Imaging Spectroradiometer satellite observations from 2000 to 2013 to suggest that ice-out is occurring sooner in Arctic lakes. Early ice-out would result in longer periods of open water while altered precipitation patterns might cause longer residence times in Arctic lakes. Earlier ice-out and longer residence times create opportunities for photochemical and biological degradation in lakes (Curtis & Schindler, 1997; Osburn et al., 2011; Waiser & Robarts, 2004) but also cause lakes to accumulate recalcitrant DOM reaching the high mean concentrations ($>$30 mg L$^{-1}$) we have observed in the Central lake group in SW Greenland.

Second, strong evidence from this and prior work indicates that sunlight plays a key role in shaping lake DOM properties (Anderson & Stedmon, 2007). It is well known that Arctic freshwater DOM is highly reactive to both sunlight and bacteria (Cory et al., 2014; Mann et al., 2012; Osburn et al., 2009). Despite the clear evidence for the importance of photochemistry in carbon cycling via the production of CO$_2$ from DOM (e.g., Cory et al,
Acknowledgments
This work was funded by the UK Natural Environment Research Council grant NE/NE/1022063/1. We thank Tania Cresswell-Maynard, John Green, Scott Warren, and Mark Breckles for help with sample collection and Stephen Richardson, Roxane Bowden, Brittany Adams, Daniel Rojas-Jiménez, and Lauren Handels for technical assistance in the laboratory. J.F. Lapierre and an anonymous reviewer are thanked for their comments which improved this manuscript. Readers can access the data in this manuscript from the University of Essex Research Data Repository: http://dx.doi.org/10.5526/ERDR-00000067.

References

2014; Osburn et al., 2001; Spencer et al., 2009), macrophyte-dominated lakes may in fact act as CO2 sinks (Tank et al., 2011). Thus, further work is required to elucidate for the SW Greenland lakes, their ultimate role in CO2 evasion or sequestration.

Third, the role of microbial activity in degrading DOM in SW Greenland lakes is virtually unknown. Our results clearly show dynamic changes in the quality of DOM on a seasonal basis (Figure 6). The trend in these changes appears to be driven by environmental factors such as seasonal evapoconcentration and photodegradation, but perhaps modulated by biological processing. The arid environment of SW Greenland makes runoff less important than littoral or pelagic processes mediated by aquatic microbes. The oligotrophic nature of these lakes (Chl a < 1 μg L−1; Whiteford et al., 2016) suggests that planktonic production of DOM is low. Knowledge of bacterial community composition, enzyme activities, rates of bacterial production, and rates of respiration are key important biogeochemical information sorely needed for these lakes.

In summary, we have examined the DOM concentration and quality for lakes in SW Greenland over a climatic gradient of cool, wet coastal conditions to a warm, dry inland interior, to cool, dry climate close to the present margin of the Greenland ice sheet. We found large variability that results in rapid changes to DOC and CDOM concentration and quality, which appears to result from changing source inputs (phytoplankton and macrophytes) and rapid microbial and photochemical cycling (Cory et al., 2014). DOC-specific absorbance (SUVA254) and fluorescence (SUVA F675) elucidated distinctions among soil inputs of DOM, internal DOM production, and photochemical (and possibly microbial) degradation, insightful metrics to evaluate lake ecosystems with respect to DOM quality along gradients of hydrological connectivity and climate. The age of the organic matter, the relative abundance of terrestrial biomarkers such as lignin, bacterial activity, and the importance of benthic production in these lakes are knowledge gaps that could provide further insight into the rate of DOM input, cycling and, by extension, CO2 sequestration or evasion. Given the large number of lakes across the de-glaciated landscape of SW Greenland, our results support prior findings that Greenland’s inland waters could be an important regulator of its regional carbon cycling (e.g., Anderson et al., 2009; Cole et al., 2007; Saros et al., 2015; Tranvik et al., 2009).


