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Published in:
Frontiers in Microbiology

Link to article, DOI:
10.3389/fmicb.2016.00576

Publication date:
2016

Document Version
Publisher's PDF, also known as Version of record

Link back to DTU Orbit

Citation (APA):
Sequence-Based Characterization of Tn5801-Like Genomic Islands in Tetracycline-Resistant Staphylococcus pseudintermedius and Other Gram-positive Bacteria from Humans and Animals

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Antibiotic resistance in pathogens is often associated with mobile genetic elements, such as genomic islands (GI) including integrative and conjugative elements (ICEs). These can transfer resistance genes within and between bacteria from humans and/or animals. The aim of this study was to investigate whether Tn5801-like GIs carrying the tetracycline resistance gene, tet(M), are common in Staphylococcus pseudintermedius from pets, and to do an overall sequences-based characterization of Tn5801-like GIs detected in Gram-positive bacteria from humans and animals. A total of 27 tetracycline-resistant S. pseudintermedius isolates from Danish pets (1998–2005) were screened for tet(M) by PCR. Selected isolates (13) were screened for GI- or ICE-specific genes (intTn5801 or xisTn916) and their tet(M) gene was sequenced (Sanger-method). Long-range PCR mappings and whole-genome-sequencing (Illumina) were performed for selected S. pseudintermedius-isolates (seven and three isolates, respectively) as well as for human S. aureus isolates (seven and one isolates, respectively) and one porcine Enterococcus faecium isolate known to carry Tn5801-like GIs. All 27 S. pseudintermedius were positive for tet(M). Out of 13 selected isolates, seven contained Tn5801-like GIs and six contained Tn916-like ICEs. Two different Tn5801-like GI types were detected among S. pseudintermedius isolates (Tn5801 and GI6287) - both showed high similarity compared to GenBank sequences from human pathogens. Two distinct Tn5801-like GI types were detected among the porcine E. faecium and human S. aureus isolates (Tn6014 and GI6288). Tn5801-like GIs were detected in GenBank-sequences from Gram-positive bacteria of human, animal or food origin worldwide. Known Tn5801-like GIs were divided into seven types. The results showed that Tn5801-like GIs appear to be relatively common in tetracycline-resistant S. pseudintermedius in Denmark. Almost identical Tn5801-like GIs were identified in different Gram-positive species of pet and human origin, suggesting that horizontal transfer of these elements has occurred between S. pseudintermedius from pets and human pathogens, including S. aureus.

Keywords: tet(M), integrative and conjugative elements, ICEs, GIs, transmission, horizontal gene transfer, Tn9f6, mobile genetic elements
INTRODUCTION

The emergence of antibiotic resistance among pathogenic bacteria is a general problem which is regarded as a threat to public health in many countries – both human and veterinary infections caused by antibiotic resistant pathogens pose a challenge to treatment and have high economic costs (Marshall and Levy, 2011; Wernli et al., 2011; Wegener, 2012; U.S. Department of Health and Human Services - Centers for Disease Control and Prevention [CDC], 2013). 

Resistant bacteria can develop in a human or animal host by evolutionary selection due to the presence of antibiotics, e.g., in connection with antibiotic therapy (Davies and Davies, 2010; Marshall and Levy, 2011). In addition, a host can acquire resistant bacteria by transmission from an outside source, e.g., via contact with other people and/or animals or via environmental exposure (Guardabassi et al., 2004a; Wegener, 2012; Da Costa et al., 2013). Transmissions of multiple antibiotic-resistant Staphylococcus pseudintermedius between dogs and humans has been reported (Guardabassi et al., 2004a). *S. pseudintermedius* is the most important staphylococcal pathogen in dogs where it is primarily associated with skin and ear infections, but it rarely causes infections in humans. Occasionally, this species is isolated from skin infection in cats (Bannoehr and Guardabassi, 2012).

Antibiotic resistance in pathogens is often associated with mobile genetic elements (MGEs) that can transfer antibiotic resistance genes within and between bacteria from human and/or animal hosts (Burrous et al., 2002; Bennett, 2008; Roberts and Mullany, 2011). MGEs include plasmids, transposable elements, prophages and different types of genomic islands (GIs) such as integrative and conjugative elements (ICEs) (Wozniak and Waldor, 2010).

Tn5801-like GIs belong to the family of Tn916-like ICEs. Tn916 was originally identified in the late 1970s as an 18-kb conjugative transposon from *Enterococcus faecalis* DS16 (Franke and Clewell, 1981). Many Tn916-like ICEs have a broad host range and are responsible for dissemination of the tetracycline resistance gene tet(M) in Gram-positive bacteria from humans and animals (Franke and Clewell, 1981; Rice, 1998; Roberts and Mullany, 2011). Tn5801 is a putative ICE (25.3 kb), however, this has not been proven. Like Tn916, Tn5801 is also associated with the tet(M) gene. The tet(M) gene encodes a ribosomal protection protein conferring resistance by protecting the ribosome against the action of tetracyclines (Chopra and Roberts, 2001). Tn5801 was first described in a clinical *S. aureus* isolate, Mu50, from a Japanese boy (Kuroda et al., 2001). Tn5801-like GIs have subsequently been identified in other human *S. aureus* and *Streptococcus* isolates, mainly in pathogenic, but also in commensal strains (Kuroda et al., 2001; de Vries et al., 2009; Denapaite et al., 2010; Holden et al., 2010; Mingoa et al., 2013; Benciani et al., 2014). Comparative genome hybridization analysis recently showed Tn5801 to be specifically associated with human methicillin-resistant *S. aureus* (MRSA) in China (Li et al., 2013). Only a few Tn5801-like elements from animal-associated bacterial isolates have been reported so far: a partially sequenced element, CW459tet(M) in *Clostridium perfringens* isolated from porcine feces, and an element detected in an *E. faecium* isolate CICTY-205 from a healthy pig in Spain (Roberts et al., 2001; Jurado-Rabadán et al., 2014). For the latter, only tet(M) was sequenced. Finally, a Tn5801-like GI has recently been reported in a fully sequenced genome from a methicillin resistant *S. pseudintermedius* (MRSP) strain ED99 isolated from a dog in the UK (Ben Zakour et al., 2012).

The importance of GIs/ICEs associated with tet(M) was highlighted in a recent study. Previous extensive clinical use of tetracycline and corresponding integration of Tn916- and Tn5801-like elements in *Streptococcus agalactiae* was shown to have had great significance for the dissemination of specific pathogenic *S. agalactiae* clones in humans (Da Cunha et al., 2014).

Characteristic of MGEs, including the Tn916-like family, Tn5801-like elements have a modular organization (Figure 1A; Burrous et al., 2002; Toussaint and Merlin, 2002; Roberts and Mullany, 2009). Tn5801 contains ORFs similar to those of Tn916, but differs by containing an integrase gene (intTn916) different from the excisionase/integrase genes (xisTn916/intTn916) present in Tn916. The intTn5801 - and intTn916 genes are of different sizes and show very low DNA identity (<50%) – they encode different types of tyrosine recombinases. The intTn5801 gene is identical to the int459 gene from the element CW459tet(M) (Roberts et al., 2001). Both Tn5801 and CW459tet(M) are integrated at the same site, located at the 3‘end of an ORF, predicted to encode GMP synthetase, whereas Tn916 can integrate at random sites in most hosts, but preferentially into AT-rich sequences (Burrous et al., 2002). The tet(M) genes from Tn5801 and Tn916 show high similarity (97.7% DNA identity). Only one tet(M)Tn5801-allele has been detected so far, and this allele appears to be specifically associated with Tn5801-like GIs (de Vries et al., 2009).

The overall aim of this study was to investigate whether Tn5801-like elements carrying tet(M) are common in clinical *S. pseudintermedius* isolates from pets, where they may constitute a likely reservoir for human pathogens (Guardabassi et al., 2004a,b). Another aim of the study was to perform an overall sequence-based characterization of Tn5801-like GIs detected in Gram-positive bacteria from humans and animals in order to get an overview of different Tn5801-like GI types and the degree of Tn5801-like GI dissemination. The study was performed by screening selected tetracycline-resistant *S. pseudintermedius* from veterinary diagnostic isolates for tet(M) as well as for int or xis genes specific for Tn5801- or Tn916-like elements, respectively. PCR mapping and gene/genome sequencing was performed in order to characterize and compare different Tn5801-like GIs detected in pet-associated *S. pseudintermedius* isolates as well as in human *S. aureus* isolates and one pig *E. faecium* isolate from previous studies (de Vries et al., 2009; Jurado-Rabadán et al., 2014). We conducted an overall comparison of full length Tn5801-like GIs characterized in this study and detected in genome sequences submitted to GenBank.

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1 Centers for Disease Control and Prevention (CDC): http://cdc.gov/features/antibioticresistance/
de Vries et al. Tn
5801-Like GIs

FIGURE 1 | Schematic illustration of Tn
5801 and the strategy for PCR mapping of full-length Tn
5801-like elements. (A) Illustration of the 25 kb putative integrative-conjugative element (ICE) integrated into the 3′ end of a GMP synthetase gene guaA (black dotted arrow) in Staphylococcus aureus Mu50. Open reading frames (ORFs) are illustrated with arrows. Besides tet(M) (striped arrow), Tn
5801 consists of three structural domains containing genes associated with conjugation (dark gray arrows), regulation (light gray arrows), and recombination (white dotted arrow) typically of elements belonging to the Tn916-family. ORFs whose functions are unknown (SAV393, SAV404, SAV406, SAV410, and SAV413-415) are illustrated with white arrows. SAV415 is predicted to encode a transposase. Direct repeats of 11 bp located in both ends of Tn
5801 are chromosomal junctions and compose a putative core site for integration of the element (Ito et al., 2003). (B) Schematic illustration of primers (small arrows) designed based on the Tn
5801 sequence (BA000017) and predicted PCR-products (rectangles) with expected product size listed below (see Supplementary Tables S2 and S3). (C) Schematic illustration of results of the PCR mapping of Tn
5801-like genomic islands (GIs) for seven pet-associated S. pseudintermedius isolates, seven human S. aureus isolates and one porcine Enterococcus faecium isolate.

MATERIALS AND METHODS

Strains

Twenty-seven tetracycline-resistant isolates from dogs and one cat were sampled and identified as S. intermedius by biochemical tests, morphology and 16S sequencing (Supplementary Table S1). Based on host origin and PCR-screening, all isolates were further identified as S. pseudintermedius (Bannoehr and Guardabassi, 2012). All isolates were diagnostic submissions to The National Veterinary Institute, Technical University of Denmark. Thirteen S. pseudintermedius isolates selected to cover the whole isolation period, 1998–2005 were characterized further in regard to tet(M) and Tn
5801-like sequences (Table 1). In addition, one porcine E. faecium isolate and seven human S. aureus isolates identified in previous studies were also characterized (Table 1). All 13 S. pseudintermedius isolates, except one isolate, 200007910-1, were confirmed to be tetracycline-resistant by the use of Sensititre and MIC testing (Aarestrup et al., 2000a). Susceptibility MIC-testing using cefoxitin showed all 13 isolates to be methicillin sensitive S. pseudintermedius ( MSSP).

Besides tetracycline and cefoxitin, the isolates were also tested for susceptibility to penicillin, streptomycin, erythromycin, ciprofloxacin, spectinomycin, tiamulin, trimethoprim, chloramphenicol, florfenicol, gentamicin, and sulfamethoxazole by the Sensititre method as described previously (Aarestrup et al., 2000a).

PCR Screening and Sequencing of tet(M), intTn
5801 and xisTn916 in S. pseudintermedius

All 27 S. pseudintermedius isolates were screened for tet(M) by PCR as described previously (see Supplementary Table S2; Aarestrup et al., 2000b). Thirteen tet(M)-positive isolates (Table 1) were selected for tet(M)-gene-sequencing by the Sanger method in a manner similar to a previously described strategy (see Supplementary Information; de Vries et al., 2009). In addition, the 13 isolates were screened for intTn
5801 or xisTn916 genes by PCR, and selected screening products were sequenced by the Sanger method and compared to sequences
### TABLE 1 | Selected strains with Tn5801-like genomic islands (GIs) characterized in this study.

<table>
<thead>
<tr>
<th>Strains</th>
<th>Species (typing info)</th>
<th>AR phenotypes</th>
<th>Element type/[tet(M) type]</th>
<th>Source/country</th>
<th>Year</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>2001-08299-1</td>
<td><em>S. pseudintermedius</em> (5-7-2-3-1-1)</td>
<td>TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>2001</td>
<td>This study</td>
</tr>
<tr>
<td>2005-06729-1</td>
<td><em>S. pseudintermedius</em></td>
<td>PEN&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Cat/Denmark</td>
<td>1998</td>
<td>This study</td>
</tr>
<tr>
<td>98-41998-1</td>
<td><em>S. pseudintermedius</em> (1-3-2-20-11-1-1)</td>
<td>ERY&lt;sup&gt;R&lt;/sup&gt; PEN&lt;sup&gt;R&lt;/sup&gt; SPE&lt;sup&gt;R&lt;/sup&gt; SMX&lt;sup&gt;R&lt;/sup&gt; STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt; TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>1999</td>
<td>This study</td>
</tr>
<tr>
<td>99-07249-2</td>
<td><em>S. pseudintermedius</em></td>
<td>CHL&lt;sup&gt;R&lt;/sup&gt;ERY&lt;sup&gt;R&lt;/sup&gt; PEN&lt;sup&gt;R&lt;/sup&gt;STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>1998</td>
<td>This study</td>
</tr>
<tr>
<td>98-41778-1</td>
<td><em>S. pseudintermedius</em> (1-3-2-20-11-1-1)</td>
<td>CHL&lt;sup&gt;R&lt;/sup&gt;ERY&lt;sup&gt;R&lt;/sup&gt; PEN&lt;sup&gt;R&lt;/sup&gt;STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>1998</td>
<td>This study</td>
</tr>
<tr>
<td>2005-06768-1</td>
<td><em>S. pseudintermedius</em></td>
<td>CHL&lt;sup&gt;R&lt;/sup&gt;ERY&lt;sup&gt;R&lt;/sup&gt; PEN&lt;sup&gt;R&lt;/sup&gt;STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>1999</td>
<td>This study</td>
</tr>
<tr>
<td>2003-07869-1</td>
<td><em>S. pseudintermedius</em></td>
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<td>Tn5801/[tet(M)&lt;sub&gt;Tn5801&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>2003</td>
<td>This study</td>
</tr>
<tr>
<td>2000-7910-1</td>
<td><em>S. pseudintermedius</em></td>
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<td>Tn916-like ICE/[tet(M)&lt;sub&gt;Tn916&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>2000</td>
<td>This study</td>
</tr>
<tr>
<td>2001-08127-3</td>
<td><em>S. pseudintermedius</em></td>
<td>SMX&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn916-like ICE/[tet(M)&lt;sub&gt;Tn916&lt;/sub&gt;-k1e type -2]</td>
<td>Dog/Denmark</td>
<td>2001</td>
<td>This study</td>
</tr>
<tr>
<td>2001-08050-1</td>
<td><em>S. pseudintermedius</em></td>
<td>ERY&lt;sup&gt;R&lt;/sup&gt;PEN&lt;sup&gt;R&lt;/sup&gt; STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn916-like ICE/[tet(M)&lt;sub&gt;Tn916&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>2001</td>
<td>This study</td>
</tr>
<tr>
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<td>ERY&lt;sup&gt;R&lt;/sup&gt;PEN&lt;sup&gt;R&lt;/sup&gt;STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt; TMP&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn916-like ICE/[tet(M)&lt;sub&gt;Tn916&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>2005</td>
<td>This study</td>
</tr>
<tr>
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<td><em>S. pseudintermedius</em></td>
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<td>Tn916-like ICE/[tet(M)&lt;sub&gt;Tn916&lt;/sub&gt;-k1e type -1]</td>
<td>Dog/Denmark</td>
<td>1999</td>
<td>This study</td>
</tr>
<tr>
<td>CICYT-205</td>
<td><em>E. faecium</em> (1-9-5-1-20-3-ST 437)</td>
<td>ERY&lt;sup&gt;R&lt;/sup&gt;KAN&lt;sup&gt;R&lt;/sup&gt;MFX&lt;sup&gt;R&lt;/sup&gt; STR&lt;sup&gt;R&lt;/sup&gt;TET&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn5801</td>
<td>Pg/Spain</td>
<td>2005</td>
<td>Jurado-Rabadán et al., 2014</td>
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<tr>
<td>213</td>
<td><em>S. aureus</em> (0008/CC5)</td>
<td>PEN&lt;sup&gt;R&lt;/sup&gt;STR&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn6014/[tet(M)&lt;sub&gt;Tn6014&lt;/sub&gt;-k1e type -1]</td>
<td>Human/Denmark</td>
<td>1957</td>
<td>de Wied et al., 2009</td>
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<td>229</td>
<td><em>S. aureus</em> (0008/CC5)</td>
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<td>1957</td>
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<td>1742</td>
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<td>Tn6014/[tet(M)&lt;sub&gt;Tn6014&lt;/sub&gt;-k1e type -1]</td>
<td>Human/Denmark</td>
<td>1963</td>
<td>de Wied et al., 2009</td>
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<tr>
<td>33597</td>
<td><em>S. aureus</em> (0037/CC8)</td>
<td>CIP&lt;sup&gt;R&lt;/sup&gt;ERY&lt;sup&gt;R&lt;/sup&gt;GEN&lt;sup&gt;R&lt;/sup&gt;MET&lt;sup&gt;R&lt;/sup&gt;PEN&lt;sup&gt;R&lt;/sup&gt;STR&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Tn6014/[tet(M)&lt;sub&gt;Tn6014&lt;/sub&gt;-k1e type -1]</td>
<td>Human/Denmark</td>
<td>2000</td>
<td>de Wied et al., 2009</td>
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<td>34148</td>
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<td>Human/Denmark</td>
<td>2000</td>
<td>de Wied et al., 2009</td>
</tr>
</tbody>
</table>

CHL, chloramphenicol; CIP, ciprofloxacin; ERY, erythromycin; GEN, gentamicin; KAN, kanamycin; MET, methicillin; MXF, moxifloxacin; PEN, penicillin; SMX, sulfamethoxazole; SPT, Spectinomycin; STR, streptomycin; TMP, trimethoprim; a, resistant; ST, sequence type; MLST, multi locus sequence type; AR, antibiotic resistance.

<sup>a</sup> MLST allelic profile and ST type (if known) determined by submitting assembled contigs to MLST typing tool (1.7) from Center for Genomic Epidemiology<sup>2</sup> (Larsen et al., 2012).

<sup>b</sup>MIC-testing showed this isolate to have an MIC for tetracycline ≤0.5 μg/ml and was therefore not confirmed to be TET<sup>R</sup>. The tet(M)<sub>Tn5801</sub>-k1e type -1 gene contained 3 bp deletion compared to tet(M)<sub>Tn916</sub>-k1e type -2 which may explain the lack of TET<sup>R</sup> phenotype.

<sup>2</sup> Center for Genomic Epidemiology MLST-typing tool: MLST 1.7 (MultiLocus Sequence Typing): https://cge.cbs.dtu.dk/services/MLST/
from Tn5801 and Tn916 as described in former studies (Agero et al., 2002; de Vries et al., 2009). (See Supplementary Information, including Supplementary Tables S2 and S3 for further details).

**PCR Mapping of Tn5801-Like GIs from Pet-Associated S. pseudintermedius, Human S. aureus and a Porcine E. faecium**

The strategy for mapping the Tn5801-like elements is outlined in Figures 1A,B. Elements detected in pet-associated S. pseudintermedius from this study and previously detected elements in isolates from human S. aureus and porcine E. faecium were mapped (de Vries et al., 2009; Jurado-Rabadán et al., 2014). The mapping was performed by Long range PCR with the Phusion High Fidelity DNA Polymerase (Finnzymes, Finland) using the conditions recommended by the manufacturer. DNA Taq polymerase (Ampliqon, Denmark) was used to obtain PCR products expected to be <2000 bp. The PCR conditions and primers are listed in Supplementary Tables S2 and S3. S. aureus Mu50 was used as positive control and a S. pseudintermedius strain 200307768-1 that was shown to be negative for int<sub>Tn5801</sub> and positive forxis<sub>Tn5801</sub>, was used as negative control. Besides int<sub>Tn5801</sub>, selected PCR-products, ex3 and ex4, were fully or partially sequenced by the Sanger method and compared to corresponding sequences in Tn5801.

**Whole-Genome Sequencing of Selected S. pseudintermedius, S. aureus, and E. faecium**

Based on the PCR mapping of detected Tn5801-like elements and tet(M) sequence types 3 S. pseudintermedius strains (2001-08299-1, 9841998-1, and 9841787-1), S. aureus 1680 and E. faecium CICYT-205 were selected for whole-genome sequencing. Genomic DNA was purified by using the Easy-DNA gDNA Purification kit (Life Sciences). For each isolate, a sequencing library was constructed and pooled using Nextera DNA Sample Preparation Kit (Illumina). Sequencing was performed using the Illumina platform, MiSeq (paired-end reads). Paired-end reads were trimmed directly by the MiSeq in order to remove sequencing adaptor and primer contamination, and CLC Genomic Workbench (version 7.5.1) was used to remove low quality reads. De novo assembly was conducted using CLC Genomic Workbench (version 7.5.1). (See Supplementary Information including Supplementary Tables S4 and S5 for details regarding trimming and assembly). For all five isolates, assembled contigs of minimum 500 bp (for CICYT-205 cut-off was minimum 1000 bp contigs) with an average coverage >30x were submitted as a Whole Genome Shotgun (WGS) sequencing project to GenBank under bioProject: PRJNA264198; bioSamples: SAMN03120277, SAMN03120291, SAMN03120292, SAMN03120293, SAMN03120290; accession no: JTKN00000000, JTKO00000000, JTKP00000000, JTKQ00000000, JTKR00000000. The WGS submissions were annotated by NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP)<sup>3</sup> (Angiuoli et al., 2008).

**Bioinformatic Analysis of Sequenced Genomes with Focus on Tn5801-Like GIs**

For all the five WGS-sequences, species were confirmed by using the strain identification tool (EzTaxon server)<sup>4</sup> and/or the species finder tool (SpeciesFinder 1.0)<sup>5</sup> using annotated full length 16S rRNA sequences and/or assembled contigs as input, respectively (Larsen et al., 2014). Besides, all five WGS-sequences were analyzed for MLST allelic profile, ST type and the presence of antibiotic resistance genes using the online-tools MLST typing tool (1.7) and ResFinder<sup>6</sup> (Table 1 and Supplementary Table S6) (Zankari et al., 2012). Contigs containing Tn5801-like tet(M) and int<sub>Tn5801</sub> sequences from the sequenced genomes were detected by search annotations for “tet(M)” and “int” as well as sequences for identity to int<sub>Tn5801</sub>. For all staphylococci isolates a Tn5801-like element was detected in one contig, whereas for the E. faecium CICYT-205 isolate, specific Tn5801-like sequences were detected in two contigs that showed an overlap of 129 bp with 100% DNA identities. The two contigs were fused in Geneious (Trial version 8.0.4). Primers used in the PCR mapping were mapped to the selected contigs containing Tn5801-like elements, and expected PCR-product sizes were compared to product sizes obtained in the PCR mapping in order to verify correct assembly. Sequence analysis was performed using CLC Genomic Workbench (version 7.5.1 or 8.5.1). GenBank was searched for full length Tn5801-like GIs based on a BLASTp<sup>7</sup> search using the annotated Int protein of Tn5801 as query (GenBank accession no. BAB56554). A threshold of minimum 98% amino acid (aa.) identity was used, and 11 full-length elements for which sequences were published were selected for comparative sequence analysis together with elements sequenced in this study.

**RESULTS**

**The tet(M) Gene in S. pseudintermedius from Pets Was Detected on Tn5801- and Tn916-Like Elements**

Twenty seven tetracycline-resistant clinical S. pseudintermedius isolates were screened for the presence of the tet(M) gene. All 27 isolates were positive for the tet(M)-PCR screening (Supplementary Table S1). Of the 13 sequenced tet(M) genes, seven were highly similar to tet(M) of Tn5801 from S. aureus Mu50 (99.9–100% DNA identity), and six sequences were highly similar

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<sup>4</sup>E. Strain identification tool: http://www.ezbiocloud.net/eztaxon

<sup>5</sup>Center for Genomic Epidemiology SpeciesFinder 1.0 Server: https://cge.cbs.dtu.dk/services/SpeciesFinder/

<sup>6</sup>Center for Genomic Epidemiology Antibiotic resistance finder tool: https://cge.cbs.dtu.dk/services/ResFinder/

<sup>7</sup>NCBI BLAST: http://blast.ncbi.nlm.nih.gov/Blast.cgi
to tet(M)Tn916 from E. faecalis DS16 (99.8–100% DNA identity). Altogether two Tn5801-like tet(M)-sequence types were detected: tet(M)Tn5801 and tet(M)Tn5801-like—type-1 differing by 1/1920 bp. In addition, 3 Tn916-like-tet(M) sequence types were detected: tet(M)Tn916, tet(M)Tn916-like—type-1 and tet(M)Tn916-like—type-2 differing by 1–3/1920 bp. (See Table 1 and Supplementary Table S7 for details). PCR screenings confirmed that the 13 tested strains contained intTn5801 or xisTn916 genes. In addition, long PCR confirmed that the 7 Tn5801-like tet(M) genes were physically linked to detected intTn5801 genes (see Figures 1B,C).

### Two Different Tn5801-Like GIs Types Were Detected Among the S. pseudintermedius

PCR mapping and whole genome sequencing revealed two different Tn5801-like elements among the tested S. pseudintermedius isolates corresponding to the two different Tn5801-like tet(M) types detected (see Figures 1 and 2A).

For 5 S. pseudintermedius isolates, the PCR mapping showed that all amplified PCR products were of the same size compared to the control strain Mu50 (Figures 1B,C and Table 1). Besides tet(M)Tn5801, the intTn5801 screening product and sequenced ex4- and ex3 fragments from these strains were shown to be 100% identical to corresponding regions in Tn5801 (Figures 1B,C). The whole element present in these isolates was shown to be almost identical to Tn5801 from S. aureus Mu50 both in organization as well as on DNA sequence level (Table 2). In addition, all ORFs of this element were identical or highly similar to Tn5801 ORFs (intTn5801-sav415; Figure 2A).

For two other S. pseudintermedius strains, the PCR mappings were positive, except for int-guaA, ex4, P6 and P7. The size of P2, P3, ex3, and P5 differed from Tn5801 (Figures 1B,C). These strains contained the tet(M)Tn5801-like—type-1 gene. Compared to Tn5801, the intTn5801-like screening product showed 96% DNA identity. This element showed similar organization compared to Tn5801, but differed by being approximately 5000 bp smaller than Tn5801 corresponding to a missing upstream region containing sav413-sav415 of Tn5801 (Figure 2A and Table 2). Instead of Tn5801 sav404, this element contained a larger ORF with low similarity to sav404. Thus, this element was registered as a novel element, Gl6287 in the Transposon Nomenclature Database from the UCL Eastman Dental Institute, London (Roberts et al., 2008; Roberts and Mullany, 2011).

Both S. pseudintermedius Tn5801 and Gl6287 were found to be located in the 3’ end of putative GMP synthase genes (guaA) with high similarity (99.68% DNA identity) and downstream of a putative integrase pseudogene with high similarity (97.45% DNA identity). Both elements are flanked by 11 bp sequences that are almost identical to the direct repeats that form the core of the putative attachment sites of Tn5801 (Ito et al., 2003; Figure 2A).

### Two Other Tn5801-Like GI Types Were Characterized from the Human S. aureus Isolates and One E. faecium Isolate from Pig

Characterization of Tn5801-like GIs previously detected in human clinical S. aureus from Denmark and an E. faecium strain isolated from a healthy pig in Spain revealed 2 other Tn5801-like GI types different from the elements detected among the S. pseudintermedius isolates (see Figures 1 and 2; de Vries et al., 2009; Jurado-Rabadán et al., 2014).

The PCR mapping showed that for the seven S. aureus isolates, PCR fragment P1, P4, ex4, P5-P7 were of the same size as Tn5801 (Figures 1B,C). PCR products P2, P3, and ex3 were approximately 1.7-kb longer than the corresponding fragments detected in Mu50. As for tet(M)Tn5801, the intTn5801 screening product and the ex4 fragment showed 100% DNA identity with Tn5801. This element is referred to as Tn6014 as previously registered (de Vries et al., 2009). The whole Tn6014 sequence contained identical or highly similar ORFs and showed overall similarity compared to Tn5801 (intTn5801-sav415; Figure 2B and Table 2). In addition, Tn6014 contained an additional region of 1775 bp corresponding to two putative ORFs not present in Tn5801.

For the E. faecium isolate, all PCRs except for P6, P5 and int-guaA were shown to be of the same size as for Tn5801 (Figures 1B,C). Besides the tet(M)Tn5801 gene sequenced previously (Jurado-Rabadán et al., 2014), the intTn5801 screening product and the ex3 fragment showed 100% DNA identity with Tn5801. This element showed similarity on DNA level and a similar organization compared to Tn5801, except that it did not contain a sav415-like ORF that is predicted to encode a transposase in Tn5801 (Figure 2C and Table 2). This element was registered as a novel element: Gl6288.

Both Tn6014 and Gl6288 were found integrated into the 3’ end of guaA. The 11-bp sequences that correspond to the imperfect direct repeats included in the attL and attR sites that flank the elements were almost identical to those of Tn5801 (see Figures 2B,C).

### Sequence Analysis of S. aureus Tn6014

Further sequence analysis (BLASTn search in GenBank) of Tn6014 from the human S. aureus strain showed that most of the additional 1775 bp-region detected in Tn6014 was highly similar (99.89% DNA identity) to a region containing ORFs predicted to encode an integrase and a transposase in the genome of an E. faecalis strain 62 isolated from a stool sample of a healthy Norwegian infant (Brede et al., 2011). This isolate was reported to contain a Tn916 element at a different location (accession no. CP002491; 562138–582758) than the 1771 bp region similar to a region in Tn6014. However, we showed it to be a Tn5801-like element containing intTn5801 integrated into a guaA gene (see next paragraph). Thus, Tn6014 is shown to be a composite element that may have been formed within Gram-positives species, possibly Enterococcus.

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8http://www.ucl.ac.uk/eastman/tn/
9NCBI BLAST. http://blast.ncbi.nlm.nih.gov/Blast.cgi
FIGURE 2 | Illustration of Tn5801-like GIs from Gram-positive bacteria fully sequenced in this study. Gray arrows illustrate ORFs identical (sav393-sav397, and sav399-sav415) or similar (sav393-like-sav415-like) to corresponding ORFs in Tn5801, black arrows illustrate ORFs which were not annotated in Tn5801.

Striped arrow: tet(M), white dotted arrow: Tn5801-like integrase gene (int), black dotted arrow: guaA, tnp: transposase. Direct repeats of 11 bp located in both ends of Tn5801-like elements are chromosomal junctions and compose a putative core site for integration of the element (Ito et al., 2003). Underlined nucleotides differ from Tn5801. (A) Tn5801 from S. pseudintermedius 9841787-1 isolated from a dog and 9841998-1 isolated from a cat – sequenced GIs from the two isolates were 100% identical. GI6287 from S. pseudintermedius 200108299-1 isolated from a dog. (B) Tn6014 from human S. aureus 1980. (C) GI6288 sequenced from E. faecium CICYT205.

TABLE 2 | Comparison of whole Tn5801-like GI-sequences from this study with corresponding sequences from Tn5801, S. aureus Mu50.

<table>
<thead>
<tr>
<th>GI</th>
<th>Tn5801f</th>
<th>DNA identity (%)</th>
<th>Gaps (number of gaps/total bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. pseudintermedius Tn5801</td>
<td>25807/25814</td>
<td>99.97</td>
<td>2/25812</td>
</tr>
<tr>
<td>S. pseudintermedius GI6287</td>
<td>19172/25812</td>
<td>73.12</td>
<td>6009/25812</td>
</tr>
<tr>
<td>S. aureus Tn6014</td>
<td>25782/27589</td>
<td>93.45</td>
<td>1777/27589</td>
</tr>
<tr>
<td>E. faecium GI6288</td>
<td>24451/25812</td>
<td>94.72</td>
<td>1346/25812</td>
</tr>
</tbody>
</table>

a Determined by pairwise alignment and comparison using CLC Genomic Workbench (version 8.5.1).
b Corresponding to the largest of the compared GIs.
Comparison of Tn5801-Like GIs Shows Evidence of Horizontal Transfer between Gram-positive Species of Human and Animal Origin

Tn5801-like GIs were found to be present in GenBank sequences from different Gram-positive bacterial pathogens and a few commensals, mainly from humans, but also sequences from bacteria associated with animals and food in Europe, USA, Asia, and Australia were found (see Supplementary Table S8). A multiple alignment of 11 selected full-length Tn5801-like GIs from GenBank and the five Tn5801-like GIs characterized in this study divided the elements into seven predicted GI types. All Tn5801-like GIs characterized in this study except for Tn6014 fell into three different groups (groups 1–3; Figure 3). Pairwise comparisons of full-length elements within groups 1, 2, and 3 showed very high similarity. In addition to S. aureus Tn5801, S. pseudintermedius Tn5801 showed very high similarity to the Tn5801-like element from canine S. pseudintermedius ED99 within group 1 (99.98%). Besides, S. pseudintermedius Tn5801 showed high similarity to an element from a Lactococcus garvieae IPLA31405 strain isolated from cheese (99.40% DNA identities). Within group 2, E. faecium G16287 showed 99.88% DNA identity compared to an element from a human S. mitis B6 isolate. Within group 3, S. pseudintermedius G16287 was shown only to differ by 4 out of 20,621 bp from corresponding elements in human isolates, E. faecalis strain 62 and S. agalactiae COOH1. This comparison shows that almost identical full-length Tn5801-like GIs are present in Gram-positive species of animal and human origin, which strongly supports the occurrence of recent direct or indirect horizontal transfer of these elements between different species of different origin. These results in particular support occurrence of horizontal transfer of Tn5801 and G16287 between pet-associated S. pseudintermedius and Gram-positive pathogens of human origin.

DISCUSSION

Tetracycline resistance is common in S. pseudintermedius and is most often mediated by the tet(M) gene (Schwarz et al., 1998; Kim et al., 2005). In agreement with this, we found all the tetracycline-resistant S. pseudintermedius isolates in this study (27) to contain the tet(M) gene. We cannot completely rule out that tetracycline resistance could also partly be mediated by other genes, as this was not further investigated. However, this is unlikely, since no other tetracycline resistance genes were detected by the sequence analysis of the fully sequenced S. pseudintermedius genomes (Supplementary Table S6). For the thirteen isolates for which the tet(M) gene was sequenced, we showed that six and seven contained Tn916-like and Tn5801-like elements, respectively. This shows Tn5801-like GIs to be relatively common in tetracycline-resistant pet-associated S. pseudintermedius in Denmark, and the prevalence appears to be on the same level as for Tn916-like ICEs. Previously, Tn5801-like GIs (Tn6014) were detected in 7 out of 20 tet(M)-positive human S. aureus isolates from Denmark (1957–2001; de Vries et al., 2009). As opposed to this, 17 porcine S. aureus strains from the same study only contained tet(M) associated with Tn916-like ICEs. A Tn5801-like element was recently reported in a MRSP canine strain (Schwarz et al., 1998; Ben Zakour et al., 2012). However, this study is the first to address the prevalence of Tn5801-like elements among tetracycline-resistant S. pseudintermedius and the first report on Tn5801-like elements within MSSP.

Previous studies have suggested that specific tet(M)-alleles or subtypes of tet(M) are associated with a specific element type or element subgroups within the Tn916-like ICE family as well as other ICE families (Agero et al., 2006; de Vries et al., 2009; Jurado-Rabadán et al., 2014). Among the tested S. pseudintermedius isolates with Tn5801-like GIs two different subtypes were detected corresponding with two highly similar tet(M) sequence types that only differed by 1/1920 bp corresponding to a difference of 1/640 aa. The two tet(M) subtypes were shown both by Sanger and Illumina sequencing. This is the first report of a new Tn5801-like tet(M) subtype. This subtype may be specifically associated with G16287, however, the almost identical Tn5801-like GIs that are present in E. faecalis strain 62 and S. agalactiae COOH1 contain the known Tn5801-tet(M) – provided that the difference does not result from a sequencing mistake.

Tn5801 was detected in the genome of S. pseudintermedius isolates from four dogs and a one cat, including two isolates of the same unknown sequence type (ST). S. pseudintermedius Tn5801 was shown to be almost identical compared to the element detected in the MRSP S. pseudintermedius ED99 (UK, ST type 25) and the Tn5801 element from the Japanese human S. aureus strain (Mu50). As mentioned above, S. pseudintermedius G16287 was shown to be highly similar to elements present in human Streptococcus and Enterococcus strains from US and Norway, respectively. The fact that almost identical GIs can be found in different species of animal and human pathogens supports that horizontal transfer of these elements has occurred between the S. pseudintermedius and human pathogens – the direction of transfer cannot be determined, however. Alternatively, the different species may have received these elements from a common bacterial source.

The mechanism of transfer for Tn5801 is unknown, but the similar organization of Tn5801 and Tn916 may suggest that Tn5801, like Tn916, transfer horizontally by conjugation with a rolling circle (Rice, 1998; Mullany et al., 2002). The only evidence of horizontal transfer of an Tn5801-like element is presented in a previous study (de Vries et al., 2009). Tn6014 from human S. aureus strain 1680 was shown to transfer at very low frequencies to a recipient strain - actually only one transconjugant was shown to have received the Tn6014-tet(M) gene physically linked to the intTn5801. Attempts to detect proof of circularization of Tn6014 by PCR did not succeed (data not shown). In this study we were not able
FIGURE 3 | Comparison of full length Tn5801-like GIs from this study (5) and full-length Tn5801-like elements detected in sequences from GenBank (11).

A multiple alignment of selected Tn5801-like sequences constructed and visualized with CLC (version 7.5.1) revealed overall seven Tn5801 GI types based on similar organization and DNA identity >99%. Element sizes are shown at the end of the sequences (to the right). EFA, E. faecalis; SAG, Streptococcus agalactiae; SA, Staphylococcus aureus; SP: LG, Lactococcus garvieae; SM, Streptococcus mitis. (EF_TX0133C/Bahaman: two strains TX0133C and TX0133B).

to transfer Tn5801-like elements from S. pseudintermedius 200108299-1 or 9841998-1 to recipient strains, E. faecalis JH2-2, E. faecium BH4105, S. aureus 8794RF or S. aureus 4220RF, by filter-mating (data not shown). This was also the case in a study reporting a Tn5801-like element from S. agalactiae (Tn5801.Sag), for which circularization and conjugal transfer could not be shown (Mingoia et al., 2013). Apparently, horizontal transfer of Tn5801-like GIs occurs at extremely low frequencies and/or under highly specific conditions. These elements do not contain xis-like genes which are required for excision. Thus Tn5801-like elements are most likely mobilizable genomic islands (MGIs) that need a helper element providing them with a factor allowing their excision (Bellanger et al., 2014). Tn5801-like elements could also turn out to be true ICEs that encode their own excision, transfer by conjugation and their integration. However, we can not rule out that these elements may have spread in bacterial populations by transformation or transduction.

Overall, different types of Tn5801-like GIs were found to be present in publicly available sequences from Enterococcus-, Lactobacillus-, Lactococcus-, Staphylococcus-, Streptococcus-, and Clostridium species associated with humans, animals or foods in Europe, USA, Asia, and Australia. This does show that successful dissemination of different types of this element has occurred among Gram-positive species world-wide. The observed diversity among the Tn5801-like GIs suggests plasticity of this element-type within Gram-positives, which is supported by the characterization of Tn6014 in S. aureus. Tn6014 contains a smaller region with a predicted integrase and a transposase gene, which may derive from Enterococcus.

Tn5801-like GIs can be dated back to the 1950’s in human S. aureus (de Vries et al., 2009). In Enterococcus, a Tn5801-like GI is present in the human E. faecium E1636 strain isolated in 1961 (Supplementary Table S8; van Schaik et al., 2010). This is not many years after tetracycline resistance was first observed, and considering the complexity of these GIs it is highly unlikely that these elements should have evolved within only a few years. Thus, Tn5801-like (and Tn916-like) elements may have evolved long before tetracycline was introduced into the clinical practice and have transferred into human pathogenic species early in the so called “antibiotic era.” Tn5801-like GIs could also have existed in human pathogens or reservoirs of pathogens prior to the “antibiotic era” and evolved by acquiring the tet(M) gene.

CONCLUSION

This study presents evidence which supports that pet-associated S. pseudintermedius is a likely reservoir of Tn5801-like GIs detected in human pathogens (e.g., S. aureus). However, larger full-genome sequencing and metagenomic sequencing studies of animal-associated isolates/samples from an expanded time-period (including the pre-antibiotic era) should be
conducted in order to make a final conclusion regarding the origin of Tn5801-like GIs in human pathogens such as S. aureus. Besides, the mechanism of transfer for Tn5801-like elements remains to be shown.

AUTHOR CONTRIBUTIONS

LV and YA designed the study and did most analysis as well as data interpretation of the work. HH contributed substantially to analysis and data interpretation regarding the genome sequencing. SR did experimental work and data interpretation regarding PCR-screenings. LV drafted the paper and all co-authors (YA, HA, and SR) have critically revised it.

FUNDING

The study was supported by Metropolitan University College, Department of Technology and by the Center for Genomic Epidemiology\(^\text{10}\) grant 09-067103/DSF from the Danish Council for Strategic Research.

ACKNOWLEDGMENTS

Parts of this study were presented at the EMBO/EMBL Symposium: New Approaches and Concepts in Microbiology 2015. We want to thank Jacob D. Jensen, Hanne Mordhorst, Christina Aaby Svendsen, and Inge M. Hansen for excellent technical assistance and to Jan S. Knudsen for proofreading the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: http://journal.frontiersin.org/article/10.3389/fmicb.2016.00576

\(^{10}\) Center for Genomic Epidemiology: http://www.genomicepidemiology.org/


Roberts, A. P., Johansen, P. A., Lyra, D., Mullany, P., and Rood, J. I. (2001). Comparison of Tn5397 from Clostridium difficile, Tn916 from Enterococcus faecalis and the CW459tet(M) element from Clostridium perfringens shows that they have similar conjugation regions but different insertion and excision modules. Microbiology 147, 1243–1251. doi: 10.1099/00221287-147-5-1243


Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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