Modelling the glucose-insulin-glucagon dynamics after subcutaneous administration of native glucagon and a novel glucagon analogue in dogs

Wendt, Sabrina Lyngbye; Boye Knudsen, Carsten; Jørgensen, John Bagterp; Madsen, Henrik; Haidar, Ahmad

Publication date: 2016

Document Version
Publisher's PDF, also known as Version of record

Link back to DTU Orbit

Citation (APA):
Advanced Technologies and Treatments for Diabetes (ATTD) 2016

Topic: Closed-loop system and algorithm

Presentation preference: E-Poster

“I confirm that I am aware of conflicts of interest in my presentation. I agree to declare this conflict of interest at the beginning of my presentation during the meeting.”

Contact details of presenting author, Sabrina Lyngbye Wendt:

Email: slw@zealandpharma.com
Address: Zealand Pharma A/S, Smedeland 36, DK-2600 Glostrup, Denmark
Phone number (daytime): +45 88 77 36 43

Sabrina Lyngbye Wendt1,2,3, Carsten Boye Knudsen1, John Bagterp Jørgensen2, Henrik Madsen2, Ahmad Haidar3,4

1Department of Bioanalysis & Pharmacokinetics, Zealand Pharma A/S, Glostrup, Denmark
2Department of Applied Mathematics and Computer Science, DTU Compute, Technical University of Denmark, Kongens Lyngby, Denmark
3Institut de Recherches Cliniques de Montréal, Montreal, Quebec, Canada
4Division of Experimental Medicine, McGill University, Montreal, Quebec, Canada
MODELLING THE GLUCOSE-INSULIN-GLUCAGON DYNAMICS AFTER SUBCUTANEOUS ADMINISTRATION OF NATIVE GLUCAGON AND A NOVEL GLUCAGON ANALOGUE IN DOGS

Background and aims:
Simulation and prediction based verification are required for glucagon bolus strategies for treatment of severe hypoglycemia in diabetes patients and for glucagon administration strategies by a dual-hormone closed-loop system. We aim to develop an improved simulation model of the complex glucose-insulin-glucagon dynamics.

Method:
We adopted a physiological model of endogenous glucose production with multiplicative effects of insulin and glucagon and combined it with the Hovorka model of glucose and insulin. We estimated model parameters using data from 10 experiments in 5 dogs who received two subcutaneous bolus injections of a novel Zealand Pharma glucagon analogue (ZP-GA-1) with increased stability in liquid formulation (20 and 120 nmol/kg). Model parameters were also estimated using data from 20 experiments in 10 dogs who received two subcutaneous bolus injections of native glucagon (20 and 120 nmol/kg or 10 and 50 nmol/kg). We used the Bayesian approach to estimate model parameters by Maximum a Posteriori (MAP) estimation given priors reported in the literature.

Results:
We report posterior probability distributions for each of the model parameters. Based on visual inspection, the model fitted data satisfactorily for both glucagons. Paired t-tests confirmed that the estimated parameters were similar after SC administration of native glucagon and ZP-GA-1.

Conclusion:
The new model enables more realistic simulations of the glucose-insulin-glucagon dynamics. In a future study, we will estimate human model parameters so the model can be used for comparative simulations of native glucagon and the glucagon analogue in humans, which will benefit the development of dual-hormone closed-loop systems for treatment of diabetes.
MODELLING THE GLUCOSE-INSULIN-GLUCAGON DYNAMICS AFTER SUBCUTANEOUS ADMINISTRATION OF NATIVE GLUCAGON AND A NOVEL GLUCAGON ANALOGUE IN DOGS

Background and Aims

Simulation and prediction based verification are required for glucagon bolus strategies for treatment of severe hypoglycemia in diabetes patients and for glucagon administration strategies by a dual-hormone closed-loop system. We aim to develop an improved simulation model of the complex glucose-insulin-glucagon dynamics.

Method

We adopted a physiological model of endogenous glucose production with multiplicative effects of insulin and glucagon and combined it with the Hovorka model of glucose and insulin. We estimated model parameters using data from 10 experiments in 5 dogs who received two subcutaneous bolus injections of a novel Zealand Pharma glucagon analogue (ZP-GA-1) with increased stability in liquid formulation (20 and 120 nmol/kg). Model parameters were also estimated using data from 20 experiments in 10 dogs who received two subcutaneous bolus injections of native glucagon (20 and 120 nmol/kg or 10 and 50 nmol/kg). We used the Bayesian approach to estimate model parameters by Maximum a Posteriori (MAP) estimation given priors reported in the literature.

Results

We report posterior probability distributions for each of the model parameters. Based on visual inspection, the model fitted data satisfactorily for both glucagons. Paired t-tests confirmed that the estimated parameters were similar after SC administration of native glucagon and ZP-GA-1.

Conclusion

The new model enables more realistic simulations of the glucose-insulin-glucagon dynamics. In a future study, we will estimate human model parameters so the model can be used for comparative simulations of native glucagon and the glucagon analogue in humans, which will benefit the development of dual-hormone closed-loop systems for treatment of diabetes.